

From THE DEPARTMENT OF MEDICINE, HUDDINGE,
Karolinska Institutet, Stockholm, Sweden

**IDENTIFICATION OF NOVEL FACTORS REGULATING HUMAN ADIPOCYTE
FUNCTION AND THEIR LINK TO METABOLIC HEALTH**

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Stockholm 2019

Cover photo: Painting representing mature adipocytes surrounded by extracellular matrix and blood vessels.

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Published by Karolinska Institutet.

Printed by Eprint AB 2019

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ISBN 978-91-7831-364-8

IDENTIFICATION OF NOVEL FACTORS REGULATING HUMAN
ADIPOCYTE FUNCTION AND THEIR LINK TO METABOLIC HEALTH
THESIS FOR DOCTORAL DEGREE (Ph.D.)

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“Education is not the learning of facts, but the training of the mind to think”

-Albert Einstein

Till Meri och Lenox

ABSTRACT

The white adipose tissue (WAT) regulates energy homeostasis by storing and releasing energy in the form of fat as well as functioning as an endocrine organ secreting a myriad of different peptides. The heterogeneous WAT consists of various cell types including the lipid-storing cells termed adipocytes. The energy-storing capacity of WAT is challenged by the rapid worldwide changes in diet and physical activity. This thesis aimed to identify novel factors regulating adipocyte function and to assess their impact on metabolic health.

The transcription factor Early B Cell Factor 1 (EBF1) has previously been shown to regulate WAT morphology (adipocyte size and number). Low expression/activity is associated with WAT hypertrophy (few but large adipocytes), a metabolically detrimental phenotype. **Study I** aimed to determine if the expression and activity of EBF1 associated with metabolic risk markers. Results suggested that EBF1 expression and activity associated with parameters of the metabolic syndrome.

Adipocytes in WAT is continuously renewed however, the turnover is irreversibly increased when a person gains weight. In fact, fat cell number is increased during weight gain and kept constant after weight loss. **Study II** aimed to identify the factor/s contributing to the maintenance of the high adipocyte number. Prospective analyses in clinical cohorts identified a set of growth factors that were highly expressed in obese compared to never-obese and were kept high after weight loss. Among these, transforming growth factor beta 3 (TGFB3) induced immature adipocyte proliferation. The WAT was studied in a mouse model expressing half of the gene expression and the results displayed a reduced proliferative capacity of immature adipocytes, hypertrophic WAT and glucose intolerance.

The hypertrophic WAT is characterized by changes in several biological processes including inflammation. The chronic low grade inflammation in obesity is one of the processes believed to cause insulin resistance and type 2 diabetes mellitus. **Study III** focused on identifying upstream regulators of adipocyte inflammation. This led to the identification of *SLC19A1*, a gene encoding a cell membrane bound folate transporter. Folate is metabolized by the one-carbon-cycle, an important pathway for DNA-methylation. We linked the inflammatory effects of reduced *SLC19A1* expression to increased global DNA-methylation. In particular, methylation of a glucocorticoid receptor binding site in the promotor of the pro-inflammatory gene *CCL2* regulated its expression.

Altogether, this work contributes to the characterization of a dysfunctional WAT. Furthermore, the clinical relevance of the reported regulators of WAT function was evaluated. This knowledge confirms an emerging theory, that an important link between obesity and metabolic disease is limited WAT expansion. Therapies resolving WAT expansion exists and this knowledge could contribute in making these more effective. However, whether such therapies should replace interventions targeting behavior (nutrition and physical activity) warrants ethical appraisal and discussion.

LIST OF SCIENTIFIC PAPERS

- I. **Petrus P**, Mejhert N, Gao H, Bäckdahl J, Arner E, Arner P, Rydén M. Low early B-cell factor 1 (EBF1) activity in human subcutaneous adipose tissue is linked to a pernicious metabolic profile. *Diabetes Metab.* 2015 Dec;41(6):509-12. PMID: 25791133
- II. **Petrus P**, Mejhert N, Corrales P, Lecoutre S, Li Q, Maldonado E, Kulyté A, Lopez Y, Campbell M, Acosta JR, Laurencikiene J, Douagi I, Gao H, Martínez-Álvarez C, Hedén P, Spalding KL, Vidal-Puig A, Medina-Gomez G, Arner P, Rydén M. Transforming Growth Factor- β 3 Regulates Adipocyte Number in Subcutaneous White Adipose Tissue. *Cell Reports.* 2018 Oct. PMID: 30332637
- III. **Petrus P**, Bialesova L, Checa A, Kerr A, Naz S, Bäckdahl J, Gracia A, Toft S, Dahlman-Wright K, Hedén P, Dahlman I, Wheelock CE, Arner P, Mejhert N, Gao H, Rydén M. Adipocyte Expression of SLC19A1 Links DNA Hypermethylation to Adipose Tissue Inflammation and Insulin Resistance. *J Clin Endocrinol Metab.* 2018 Feb 1. PMID: 29121255

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LIST OF ABBREVIATIONS

BMI	Body Mass Index
CCL2	C-C Motif Chemokine Ligand 2
DNA	Deoxyribonucleic Acid
EBF1	Early B cell Factor 1
GLP-1	Glucagon-Like Peptide 1
Kcal	Kilo-calories
PPAR γ	Peroxisome Proliferator-Activated Receptor Gamma
SLC19A1	Solute Carrier Family 19 Member 1
TGFB3	Transforming Growth Factor Beta 3
WAT	White Adipose Tissue

1 BACKGROUND

1.1 OBESITY

The relevance of this thesis is signified by the obesity pandemic¹. It has never been more vital to understand the mechanisms driving obesity and its subsequent co-morbidities including type 2 diabetes, cardio vascular disease and cancer².

Obesity is defined as $BMI > 30 \text{ kg/m}^2$, *i.e.* a large body-weight in relation to the height. It is obvious that changes in the former variable drives the obesity pandemic. In the common population, the excess weight is constituted by fat stored in the WAT, which characterizes individuals with obesity. Why is obesity so prevalent today? Albert Einstein published the mass-energy equivalence formula 1905 which revealed the association between energy and mass. Hence, the chemical energy humans consume in form of food is stored in the WAT or converted to kinetic and thermal energy which manifest in body movement and heat, respectively. Today, it is common knowledge that body weight and fat mass can be reduced by eating less and moving more. Yet, understanding this concept has proven insufficient to reverse the increasing prevalence of obesity. In this section, I will discuss the origin of obesity, why it is a problem and how it can be prevented/treated.

1.1.1 Evolutionary aspects of obesity

A classical theory explaining the obesity pandemic has been the thrifty and/or the drifty genotype³. It suggests that the human body has evolved to store excess energy to survive times of famine (thrifty genotype) and/or the lack of exposure to predators (drifty genotype). Hence, human biology, which is viewed as static during the relatively short period of industrialization, is not suited to prosper in modern environments that promote an overbalance in energy⁴. Although genetics explain a fraction of the variations in BMI⁵, emerging knowledge suggest that the thrifty/drifty genotype theory is an oversimplification⁶. In fact, human biology is continuously evolving and maternal as well as paternal environments influence their own- and their offspring phenotype which is evident already in the F1 generation⁷. Hence, to understand the evolutionary aspects of obesity we must first appreciate the complexity of evolution.

It is not controversial that the modern environment is a major contributor to the obesity pandemic by encouraging high caloric intake and sedentary behavior⁸. However, not everybody becomes obese in this environment. This spurs the inevitable “nature vs nurture” question; how much of the phenotype is hardwired in human nature and how much can be modified by the environment? In reality, all life forms are constantly adapting to their environments and the acquired adaptation can be transferred transgenerationally^{6,7,9}. The epigenome, defined as modifications of the chromatin that are not changes in the nucleotide sequence, function as a bridge between the environment and biological function, including human nature¹⁰⁻¹³. For instance maternal and/or paternal diet pre-conception influence the body weight and WAT biology in the offspring^{6,9}. Thus, the influence from human nature on a trait, such as obesity, cannot be separated from the environment and *vice versa* because both

are variables, and they are interwoven. Just to give an example of this interplay; an individual with a “thrifty genotype” is more likely to become obese but being obese also influence the environment of the person via exposures to stimuli such as social stress¹⁴ and reduced healthcare quality¹⁵ which may contribute to drive the weight gain further¹⁶.

Finally, phenotypic heterogeneity in body weight is observed in genetically identical organisms raised in the same environment suggesting that there is a stochastic aspect to obesity^{17,18}. Thus, obesity may be acquired just by chance independent of the genes or environment.

1.1.2 The link between obesity and disease

Obesity is associated with a myriad of non-communicable diseases such as type 2 diabetes and cancer^{2,19–21}. Several processes have been linked to the pathophysiology of obesity including inflammation and ectopic lipid accumulation^{22,23}. Both processes are associated with a hypertrophic WAT (few but large adipocytes in relation to fat mass)^{24,25}. These observations in combination with observations linking WAT expandability and adipocyte size to metabolic disease^{25–29} indicate that obesity causes disease by exceeding the fat storage capacity. This theory suggests that the problem is not a large fat mass *per se* but rather the inability to further expand³⁰. Hence, therapies treating obesity associated metabolic disease should aim at allowing further adipose tissue expansion via mechanisms such as adipogenesis²⁹. This will be discussed in more detail in section 1.2.

1.1.3 Treatment/prevention of obesity and its comorbidities

As previously discussed, obesity becomes a problem when the body is not able to handle further energy supply. There are three solutions to the problem. The first is to reduce the energy supply which could be achieved by reducing food intake³¹. Several drugs has been used for this purpose but the most effective ones are the GLP-1 analogues³² which has several other positive antidiabetic effects as well. Another effective method to limit caloric intake is by changing the gastrointestinal anatomy via bariatric surgery³³. A second therapeutic method is to increase energy expenditure³¹. Physical exercise and browning of adipose tissue are potential therapies that could be used to convert the chemical energy to movement and heat, respectively^{24,34}. The third therapeutic target, which also is the most relevant to the work in this thesis, is to expand the storage capacity of the WAT to avoid ectopic lipid deposition and disease^{19,24,29}. An effective way to do so is by activating the master regulator of adipocyte development (adipogenesis) PPAR γ which has indeed been used as an antidiabetic therapy³⁵. However, metabolic disease is not the only factor that reduces the quality of life in individuals with obesity. Excess body fat affects other features of life quality such as social acceptance^{14–16}.

The feasibility, cost effectiveness and the ethical consideration of these therapies warrants further discussion. In my opinion, the most reasonable strategy is to change the initial problem which is how modern societies are structured⁸. It is not at all necessary to live like hunter gatherers in order to reverse and prevent the increasing prevalence of obesity. In fact,

the oversupply of energy that accumulates and result in excess body fat over time could be prevented by walking as little as 2000 to 2500 extra steps per day⁸ or eating about 10-25 less kcal of energy per day³⁶ which is equivalent to 1-2 cubes of sugar (~3-6g). Nevertheless, the WAT storage capacity needs to be enhanced if interventions aiming at behavioral changes are unsuccessful. Such strategies, in combination with elimination of the weight-stigma, could lead to a healthy obese world population.

1.2 WHITE ADIPOSE TISSUE

The WAT is the most plastic organ in the human body and a central regulator of energy homeostasis²⁴. The tissue consists of several different cell-types including macrophages, T-cells, fibroblasts, endothelial cells and adipocytes. The latter is characterized by its energy storing ability. The energy is stored as triglycerides in a lipid droplet. The adipocyte fraction comprises the majority of the WAT volume. Furthermore, the WAT is an endocrine organ that secretes a number of hormones that may act in para- and/or endocrine fashion. Many of the secreted peptides are pro-inflammatory cyto- and chemokines. In metabolically healthy individuals, the WAT regulate whole body homeostasis by buffering energy and secreting hormones (adipokines) that modulate functions in other organs. However, in an metabolically unhealthy state, often observed in individuals with obesity, these functions becomes dysfunctional and the WAT is instead characterized by inefficient expansion, inefficient lipid mobilization and a low grade chronic release of pro-inflammatory adipokines²⁴.

1.2.1 White adipose tissue expansion

The plastic ability of the WAT is regulated by the energy balance in the body. Hyper-caloric conditions demand WAT expansion whereas hypo-caloric conditions induce the release of energy from WAT and thus, resulting in a reduction of its mass²⁴. The expansion of WAT occurs via two processes, increased adipocyte size or increased adipocyte number³⁷. The latter is constantly renewed with a 10% annual turnover-rate³⁷. The inter-individual variation in the morphology for any given fat mass is large but on average, moving from low fat mass to higher is mainly associated with increased adipocyte size until a plateau of about 800 pL where the tissue starts to expand mainly via increased cell numbers³⁷. A hyperplastic WAT (many small fat cells for a given fat mass) is associated with good metabolic health^{24,26} probably because it is permissive for additional caloric over-supply. Several processes have been implicated in tissue expansion such as adipogenesis, angiogenesis, extracellular remodeling and inflammation^{24,25,28,29}. In addition, proliferation of adipocyte progenitors (undifferentiated adipocytes) is also an important mechanism as the aforementioned processes are dependent of the availability of these cells. In contrast, weight loss is only associated with reduced fat cell size but no reduction in number, at least after rapid weight loss post bariatric surgery^{37,38}. This phenomenon would suggest that a post obese state is metabolically beneficial in relation to a never obese state at a given fat mass. In fact, this notion has recently been confirmed³⁸. The underlying factors determining WAT adipocyte cellularity are poorly understood and warrant further investigation. Stimuli during gestation and early life has been proposed to be of vital importance⁶. Other mechanisms involve

specific transcription factor activity³⁹ and disturbed circadian rhythms⁴⁰. Increased understanding of the mechanisms regulating WAT expansion will allow us to develop strategies to maintain a high consumption of food combined with physical inactivity without disturbing metabolic health.

1.2.2 White adipose tissue inflammation

The intimate association between metabolism and the immune system is well established⁴¹ and has set the foundation of a new term, namely immunometabolism⁴². It is well known that obesity is associated with an inflammatory state and that the WAT contribute to the systemic inflammation^{22,43–46}. The pro-inflammatory state in WAT during obesity has been known in about two and a half decades⁴⁷ and its causal effect on insulin resistance is well studied^{41,43,44,48}. Hence, it makes a lot of sense to treat metabolic disease by lowering inflammation. However, such treatments have not been successful⁴⁹ suggesting that the inflammation may have beneficial functions in maintaining metabolic homeostasis. In fact, it has been suggested that chronic insulin exposure and insulin resistance in adipocytes precedes the inflammation^{46,50}. One could speculate that the insulin resistance is a result of energy oversupply and that the subsequent inflammation is activated to remodel the WAT to make room for the surplus energy. This model is supported by findings suggesting that transient activation of inflammation is essential for the WAT to expand and the lack of inflammation results in lipodystrophy⁵¹. Furthermore, inflammation is believed to cause insulin resistance through induced lipid mobilization from the WAT to ectopic deposition⁵². This may be true however; lipid mobilization is needed in some instances such as during physical exercise. Inhibition of inflammation during physical exercise limits visceral fat loss⁵³. Taken together, inflammation has long been considered as a metabolically detrimental process, a view that is being challenged with emerging reports. Hence, the interplay between metabolism and the immune system seem to be more dynamic and context-dependent than previously thought. Understanding this complexity could help find strategies to fine-tune it and restore immunometabolic homeostasis.

1.3 FROM ENVIRONMENT TO PHENOTYPE: THE ROLE OF THE EPIGENOME AND THE TRANSCRIPTOME

As discussed in section 1.1.1 of this thesis, biology cannot be viewed separate from its environment and they are not necessarily more or less plastic than each other. Charles Darwin's theory of natural selection may have contributed to a static view of biology but today we know that individual organisms constantly adapt to their environments, a phenomenon termed phenotypic plasticity⁵⁴. Environmental stimuli alter the structure of the chromatin itself which results in an altered gene expression¹². The metabolism functions as a bridge between the environment and the epigenome which result in an altered gene expression^{11,12}. We know of thousands of metabolites that can function as substrates or co-substrates to regulate hundreds of epigenetic modification and/or other post translational modifications in the cell¹³ whereas the most well-studied is DNA-methylation. This knowledge has helped us understand that the nucleotide order of the DNA-molecule is

insignificant without an environment. Altogether, it is important to study epigenetics and gene expression in order to fully understand the mechanisms regulating WAT dysfunction.

2 AIMS

2.1 GENERAL AIM

The overarching aim was to identify novel regulators of adipocytes and link them to traits of the metabolic syndrome. This was achieved by starting with WAT biopsies from well-phenotyped clinical cohorts and study associations between gene expression and various clinical measures or WAT phenotypes. Subsequent functional analyses of these factors were studied *in vitro* in various cell types and/or *in vivo* in a mouse model.

2.2 SPECIFIC AIMS

2.2.1 Study I

The aim was to characterize the clinical relevance of EBF1, a transcription factor previously described to regulate adipogenesis and WAT morphology.

2.2.2 Study II

The primary aim was to identify regulators of the induced and irreversible adipocyte cellularity in WAT of obese individuals.

2.2.3 Study III

The aim was to study the regulation of DNA-methylation in adipocytes and its link to adipocyte function and insulin resistance in individuals suffering from obesity.

3 METHODOLOGICAL CONSIDERATIONS

3.1 HUMAN COHORTS

All studies in this thesis include human cohorts. The study-individuals are grouped depending on the research question (such as comparisons between lean and obese or obese and post-obese etc.). Characteristics of the WAT, including the transcriptome, is mapped and correlated with clinical characterization of the study-subjects.

WAT biopsies are taken in the mornings after an overnight fast. This minimizes the influence of dietary and/or circadian factors on the results. However, the timing of the last meal and the misalignment between biological and astronomical circadian rhythms may still constitute as confounders⁵⁵. In addition, differences between groups are context dependent and may be- or not be evident in certain settings such as in the presence of insulin⁵⁶.

The study-participants are mainly females and the data may not be extrapolated to males. It warrants particular consideration when studying WAT as there are remarkable differences in its expansion and distribution between genders^{24,57}. Furthermore, the fat distribution is not taken into account when subdividing the BMI groups (*i.e.* lean, overweight or obese). Inter-individual differences in body-weight are influenced by muscle mass. However, the study participants are not athletes or body builders thus; fat mass is the main variable explaining the variations in BMI. The total fat mass is less important than the body-fat distribution as a predictor of metabolic health^{23,57}. The subdivision according to BMI was still used as it is common practice in the clinic.

The transcriptome of WAT was compared between groups to identify new factors that may influence its function. The gene expression level *ex vivo* may not be a perfect representation of the state *in vivo*. The procedure in which the biopsy is taken may induce a stress response locally in the tissue as well as systemically if the individual experience fear/nervousness when being exposed to the needle. Extraction of the different cell fractions of the WAT requires collagenase treatment and centrifugation which most likely alter the transcriptome. A better representation of the WAT transcriptome/proteome may be assessed when novel techniques are developed and made available.

3.2 CELL CULTURES

Cell cultures were used in study I and II. Differentially expressed transcripts identified in the clinical cohorts were studied *in vitro* in primary human and murine WAT-derived cells or mouse embryonic fibroblasts. These systems allow us to study the cells in an isolated setting and manipulate gene expression or the micro-environment in the conditioned media. However, adipocytes crosstalk with other cell types to facilitate tissue function *in vivo*^{42,58}. Thus, some factors may not have effects *in vitro* but still be relevant for WAT function.

As mentioned in chapter 1, WAT dysfunction is characterized by *inter alia* chronic inflammation, reduced adipogenic ability and increased basal lipolysis. Hence, these readouts

are commonly used in the *in vitro* system to characterize the role of a gene/protein on WAT function. The processes are subdivided into “good” and “bad” *i.e.* a gene that is highly expressed in a metabolically unhealthy state is expected to induce “bad” pathways and *vice versa* for the opposite. As discussed previously, these processes are dynamic and context-dependent *in vivo* and the “good/bad” subdivision of tissue function may mislead and limit our understanding of biology.

3.3 ANIMAL MODELS

A transgenic mouse model was used in study II to test the causal role of *Tgfb3* *in vivo*. It was of particular importance to use an animal model in this study as the data *in vitro* suggested that TGFB3 regulate progenitor proliferation. The effects on WAT cellularity is also dependent on adipogenesis and hence, induced proliferation may not necessarily translate into increased adipocyte cellularity.

The way humans and mice store fat is different. Mice store fat mainly in the visceral depot and humans in the subcutaneous⁵⁹. The differences between depots, at least in mice, seem to be dependent of the microenvironment of the depot and not intrinsic in the cells⁶⁰. Although mice are not equal to humans, they constitute a model organism that can be used to identify qualitative effects on WAT function.

4 RESULTS AND DISCUSSION

4.1 STUDY I

In study I, we demonstrated that EBF1 levels and activity in WAT are associated with several measures of metabolic health. As EBF1 is a well-established regulator of WAT function (regulating adipogenesis, inflammation and lipolysis)^{39,61,62}, these data support the notion that WAT function is relevant for overall metabolic health. The causal link between EBF1 expression and adipose tissue function has been established prior to this study³⁹ but they did not report the association to clinical parameters in humans. This study in combination with the previous studies on EBF1 function in WAT suggests that therapies aiming at targeting its transcriptional activity could be used to treat metabolic disease.

How is EBF1 activity regulated and how can its activity be targeted? Specific ligands that activates EBF1 activity remain unknown however, inflammation (*i.e.* TNF α stimulation *in vitro*) reduce *EBF1* expression in adipocytes³⁹. This is not likely mediated via direct mechanisms as TNF α stimulation perturbs the characteristics of adipocytes, including high *EBF1* expression. It is tempting to just imagine the DNA strand as a two-dimensional static molecule that EBF1 bind to, at specific motifs, after receiving a signal in form of a ligand or similar. Today we know that the chromatin is a three-dimensional structure and it is dynamic⁶³. It can move specific regions of DNA into machineries of proteins involved in specific nuclear functions such as gene transcription. The three-dimensional chromatin structure is in large parts shaped by the cellular metabolic state via epigenetic mechanisms¹¹. Thus, it is conceivable that EBF1 activity is regulated by specific metabolic states which shape the chromatin allowing EBF1-motifs to be accessible for gene transcription.

In mice and humans EBF1 activity is associated with adipocyte cellularity independent of fat mass³⁹ suggesting that interventions increasing EBF1 activity could increase adipocyte number without influencing total fat mass. However, in individuals with behavioral patterns resulting in a caloric oversupply, the WAT will continue to expand and a threshold for a hypertrophic phenotype will be reached at a larger fat mass. This suggests that potential therapies targeting EBF1 activity should be combined with intervention managing behavioral patterns.

4.2 STUDY II

Study II was performed to identify regulators of WAT adipocyte cellularity. A novel regulator of adipocyte cellularity, namely TGFB3 was identified. This factor is proposed to induce fat cell number in obesity by regulating pre-adipocyte proliferation. When obese individuals lose weight (termed post-obesity) *TGFB3* expression is unaltered indicating that it contributes and explains part of the maintenance of elevated and irreversible adipocyte turnover once an individual becomes obese. Mice expressing half the mRNA levels compared to wild-type littermates displayed perturbed glucose metabolism when fed with a high fat diet

but not on a regular chow diet. These findings support that WAT expandability is important for metabolic health.

The factors inducing *TGFB3* expression during weight gain remain unknown but it is possible that, as discussed above, the metabolic changes induce its expression via epigenetic mechanisms. Another explanation could be that the progenitor pool is induced during hypercaloric conditions and the accumulation of this cell type maintains a high *TGFB3* expression in the WAT. Nevertheless, instead of understanding the mechanisms regulating its expression, recombinant TGF β 3 can be injected in patients. In fact, this has already been tested for anti-scarring therapies under the trademark Avotermin⁶⁴. However, the timing of TGF β 3 injections in relation to adipogenic signals need to be investigated in order to expand the tissue efficiently as a chronic induction of progenitor proliferation may inhibit adipocyte differentiation.

4.3 STUDY III

A novel regulator of inflammation, namely *SLC19A1*, was identified and characterized in Study III. We observed that the expression of several genes involved in folate and methionine metabolism (one carbon cycle) was altered in WAT of individuals with obesity. *SLC19A1* expression displayed the strongest association to pro-inflammatory pathways. We linked the reduced expression of this gene to an induced global DNA-methylation. In particular, we showed that DNA-methylation in the promoter of the pro-inflammatory gene *CCL2* induced its gene expression by modulating glucocorticoid receptor activity. These findings contribute to the understanding of the link between metabolism and adipocyte function.

An important controversy of this study is that knockdown of *SLC19A1* *in vitro* resulted in increased DNA methylation even though folate and the universal methyl donor S-adenosylmethionine levels decreased. In fact, whole cellular metabolism is inter-woven and the one carbon cycle is not an isolated system⁶⁵. In this study we reduced the expression of one gene encoding a folate transporter. The intracellular folate is connected to a myriad of metabolic pathways which involves several other intermediary metabolites as well as other enzymes and transporters⁶⁵. Furthermore, S-adenosylmethionine and DNA methylation is just one out of hundreds of epigenetic modifications that may be influenced by the metabolome¹³. Understanding the complexity of such systems is made possible with omics tools and deeper understanding of these systems will probably be elucidated in the near future.

This study suggest that folate metabolism is important for adipocyte function and metabolic health which is supported by a randomized control trial in humans⁶⁶. However, folate alone is not likely enough to treat insulin resistance but it may constitute an ingredient in a mixture of metabolites. In fact, several metabolites are altered in individuals with obesity and metabolic disease⁶⁷.

4.4 FUTURE PERSPECTIVE

The studies in this thesis report three regulators of adipocyte function and metabolic health. Where do we go from here? Before moving on discussing future perspectives we need to define what the end goal is. The answer should be obvious; to reduce human suffering and improve the quality of life for humankind. If this is the goal, future research should focus on identifying therapies that can regulate WAT function. These interventions could involve treatment with drugs targeting specific mechanisms in adipocyte or behavioral interventions. The latter would require understanding of what behavioral patterns that causes an unhealthy WAT phenotype (hypertrophy and inflammation).

The next step is to evaluate if therapies targeting WAT expansion and/or inflammation can be used to treat metabolic disease. If such therapies are successful, the impact on the subjective improvement in patient life-quality needs to be evaluated.

5 CONCLUSION

The structure of modern societies promotes an imbalance between energy intake and energy expenditure resulting in increased fat mass and obesity over time. This spurred scientists to map WAT function in health and disease in order to counteract obesity itself as well as its comorbidities. Collectively, decades of characterizations suggests that the problem with obesity is not that the WAT expands but rather that its expansion is limited and that once this limit is reached, individuals become metabolically sick. Chronic low grade inflammation is one characteristic of WAT that has reached its expansion. The work herein supports the existing theory and contributes to the understanding of the underlying mechanisms. The first two studies and the last study link metabolic health to tissue remodeling/expansion and inflammation, respectively. Taken together, this suggests that therapies to treat metabolic disease by mechanisms in WAT should aim at allowing it to further expand. The future will tell if humankind will solve the problem with obesity by modifying the fat storage capacity of the body, adjust the behavior to adapt to the environment or re-format the environment to adapt to the existing behavior.

“There are very few new things in this world, very few. That’s why people that are young, if they’re smart, try to profit from the experience of an older guy so they won’t have to go through all the pain and suffering. But a certain amount of pain and suffer is good, because it makes a person think they’ve learned.”

– Cus D’Amato

6 ACKNOWLEDGEMENTS

Intuitively, I would think that this thesis was accomplished mainly as a result of my hard work, intelligence and smart choices in life. But after reading *Outliers* by Malcolm Gladwell and the work of Paul Piff, I realized that this egocentric view is extremely ignorant and that my own contribution is probably very small. This would not be possible without the opportunities and help provided by Sweden and the people mentioned below.

Mikael Rydén – My main supervisor. Your knowledge, pedagogic skills and willingness to always explain make you a great supervisor. However, this is not what I appreciated the most under your supervision. You have always taken my input into consideration and if it has been bad, you have taken your time to explain why. This improved my self-confidence and I would probably not be able to develop the skill to think creatively in science without it. I also appreciate that I could be myself at work; it made the whole PhD experience much more fun.

I also want to thank you as a friend. The first time I met you I thought to myself “who is this spoiled, arrogant brat” and I did never think that we would get along. It turned out that I was completely wrong in many ways. Today I consider you as a semi-humble and very considered person.

Peter Arner – A living legend. I am amazed by your enthusiasm and love for doing research. You are a true inspiration and it has been an honor to work with you.

Hui Gao – Thank you for always being available to help. Your wide range of knowledge in different methods and bioinformatics has been very useful. The discussions with you have helped me view things from different perspectives.

Ulf Risérus – I am very grateful that you gave me the opportunity to do science in the first place and for taking my own research interest into consideration. It was a great experience working with you and it truly boosted my love for science. Thank you!

Niklas Mejhert – You have been paving the way for me, advising me throughout my PhD studies and teaching me what it requires to become a successful scientist. I hope that we will get the opportunity to work together in the future.

Simon Lecoutre – I have learned a lot by working with you. Your knowledge and hard work resulted in long discussions combined with long sessions of pipetting. I feel like I fitted another PhD during the year we worked together.

Eva Sjölin - Thank you for teaching me how to behave in the lab. **Gaby Åström** - You have always been ready to help. I do not know how many times you saved my experiments from failing. **Lisa Dungner** - Thank you for taking care of me when I started and for finding my samples every time I lost them. **Kerstin Wählén** - You made it very easy to work in the lab. So thank you for everything that I know you helped me with and for everything I took for granted (like always having all the buffers ready to use). **Thais De Castro Barbosa** and **Ana-Maria Suzuki** – I thought that Lipid lab would go under when the ladies above retired but

you proved me wrong. I am glad that I got the opportunity to work with you, Muito Obrigado. **Lena Lindberg** – For helping me with my Achilles heel, administration. **Ingrid Dahlman** – thank you for bringing me to the lipid lab and for sharing all the arrays. **Alastair Kerr** – You came with a fresh breeze of enthusiasm and curiosity which contributed to a nice work-environment. It has been great working with you. **Kelvin Kwok** – Thank you for your valuable scientific input. I am glad that I got you as a colleague and not a reviewer. I also want to thank you for creating an obesogenic environment behind me in the office. **Jesper Bäckdahl** – Thank you for all the discussions about science and life in general. We will maybe get the opportunity to answer some of these questions in the future. **Daniel Andersson & Daniel Eriksson-Hogling** – Piff & Puff – Your enthusiasm for obesity research has been inspiring. It is never a boring moment in your company. **Agne Kulyté** – Thank you for being a great neighbor in Novum and for keeping the order in the lab. **Jurga Laurencikiene** – Your expertise about the non-adipocyte cells in the adipose tissue has been extremely valuable, thank you for sharing your data and knowledge. **Bea Tavira** – Thank you for helping out with analyses in a very organized way. **Veronica Lundbäck** – I thought that I was calm under stressful situations until I met you. It has been inspiring seeing you finish your PhD.

All previous and current colleagues and co-authors – I got the opportunity to meet and work with many different people and every single one of them has contributed to my scientific development. Thank you all!

Järfälla Boxing Club – This place have been part of my life since I was 12 years old and it has had a huge impact on me and the person I am today. The members of this gym are very unique and the most special person is my coach **Timo Karjalajnen**. It cannot be explained, it can only be experienced. Being part of this team has made me, among other things, self-confident, fearless and genuine. Thank you!

Finally, I want to thank the most important people in my life. My **parents** for giving me a very comfortable life and for making sure that I had all pre-requisites needed to finish my education. **Mother** – You have always placed your kids first, you gave us endless love and you have sacrificed a lot for making our lives comfortable. I am glad that my son also got the chance to experience your love. **Father** – You have never doubted my ability to accomplish my goals and you have therefore always been proud but never impressed. Nobody knows how to live life like you. You never forced this knowledge on me but you gave me all the tools to figure it out by myself so thank you for giving me the ability to enjoy life. “**Little brother** – You are one of the special people in the boxing gym mentioned above. Thank you for all your support and for always being ready to help. **Wife** – You have always been proud over my dedication to science and supported me throughout. This has been extremely helpful to keep my motivation up during hard times. I also want to thank you complementing the lack of my social intelligence. **My son** – Thank you for bringing joy and happiness to our lives.

7 REFERENCES

1. Trends in adult body-mass index in 200 countries from 1975 to 2014: a pooled analysis of 1698 population-based measurement studies with 19·2 million participants. *Lancet* **387**, 1377–1396 (2016).
2. Kopelman, P. G. Obesity as a medical problem. *Nature* **404**, 635–43 (2000).
3. Prentice, A. M., Hennig, B. J. & Fulford, A. J. Evolutionary origins of the obesity epidemic: natural selection of thrifty genes or genetic drift following predation release? *Int. J. Obes. (Lond.)* **32**, 1607–10 (2008).
4. Hill, J. O. & Peters, J. C. Environmental Contributions to the Obesity Epidemic. *Science (80-.)* **280**, 1371–1374 (1998).
5. Locke, A. E. *et al.* Genetic studies of body mass index yield new insights for obesity biology. *Nature* **518**, 197–206 (2015).
6. Lecoutre, S., Petrus, P., Rydén, M. & Breton, C. C. *Trends Endocrinol. Metab.* **29**, 675–685 (2018).
7. Wang, Y., Liu, H. & Sun, Z. Lamarck rises from his grave: parental environment-induced epigenetic inheritance in model organisms and humans. *Biol. Rev.* **92**, 2084–2111 (2017).
8. Hill, J. O. *et al.* Obesity and the environment: where do we go from here? *Science* **299**, 853–5 (2003).
9. Öst, A. *et al.* Paternal diet defines offspring chromatin state and intergenerational obesity. *Cell* **159**, 1352–64 (2014).
10. Jirtle, R. L. & Skinner, M. K. Environmental epigenomics and disease susceptibility. *Nat. Rev. Genet.* **8**, 253–262 (2007).
11. Berger, S. L. & Sassone-Corsi, P. Metabolic Signaling to Chromatin. *Cold Spring Harb. Perspect. Biol.* **8**, a019463 (2016).
12. Etchegaray, J. P. & Mostoslavsky, R. Interplay between Metabolism and Epigenetics: A Nuclear Adaptation to Environmental Changes. *Mol. Cell* **62**, 695–711 (2016).
13. Reid, M. A., Dai, Z. & Locasale, J. W. The impact of cellular metabolism on chromatin dynamics and epigenetics. *Nat. Cell Biol.* **19**, 1298–1306 (2017).
14. Richardson, S. A., Goodman, N., Hastorf, A. H. & Dornbusch, S. M. Cultural Uniformity in Reaction to Physical Disabilities. *Am. Sociol. Rev.* **26**, 241 (1961).
15. Phelan, S. M. *et al.* Impact of weight bias and stigma on quality of care and outcomes for patients with obesity. *Obes. Rev.* **16**, 319–326 (2015).
16. Tomiyama, A. J. *et al.* How and why weight stigma drives the obesity ‘epidemic’ and harms health. *BMC Med.* **16**, 123 (2018).
17. Gärtner, K. A third component causing random variability beside environment and genotype. A reason for the limited success of a 30 year long effort to standardize laboratory animals? *Int. J. Epidemiol.* **41**, 335–341 (2012).
18. Dalgaard, K. *et al.* Trim28 Haploinsufficiency Triggers Bi-stable Epigenetic Obesity.

Cell **164**, 353–364 (2016).

19. Lengyel, E., Makowski, L., DiGiovanni, J. & Kolonin, M. G. Cancer as a Matter of Fat: The Crosstalk between Adipose Tissue and Tumors. *Trends in Cancer* **4**, 374–384 (2018).
20. Font-Burgada, J., Sun, B. & Karin, M. Obesity and Cancer: The Oil that Feeds the Flame. *Cell Metab.* **23**, 48–62 (2016).
21. Samuel, V. T. & Shulman, G. I. The pathogenesis of insulin resistance: integrating signaling pathways and substrate flux. *J. Clin. Invest.* **126**, 12–22 (2016).
22. Donath, M. Y. & Shoelson, S. E. Type 2 diabetes as an inflammatory disease. *Nat. Rev. Immunol.* **11**, 98–107 (2011).
23. Tchernof, A. & Després, J.-P. Pathophysiology of human visceral obesity: an update. *Physiol. Rev.* **93**, 359–404 (2013).
24. Rosen, E. D. & Spiegelman, B. M. What We Talk About When We Talk About Fat. *Cell* **156**, 20–44 (2014).
25. Crewe, C., An, Y. A. & Scherer, P. E. The ominous triad of adipose tissue dysfunction: inflammation, fibrosis, and impaired angiogenesis. *J. Clin. Invest.* **127**, 74–82 (2017).
26. Acosta, J. R. *et al.* Increased fat cell size: a major phenotype of subcutaneous white adipose tissue in non-obese individuals with type 2 diabetes. *Diabetologia* **59**, 560–570 (2016).
27. Lönn, M., Mehlig, K., Bengtsson, C. & Lissner, L. Adipocyte size predicts incidence of type 2 diabetes in women. *FASEB J.* **24**, 326–331 (2010).
28. Sun, K., Kusminski, C. M. & Scherer, P. E. Adipose tissue remodeling and obesity. *J. Clin. Invest.* **121**, 2094–101 (2011).
29. Ghaben, A. L. & Scherer, P. E. Adipogenesis and metabolic health. *Nat. Rev. Mol. Cell Biol.* (2019). doi:10.1038/s41580-018-0093-z
30. Carobbio, S., Pellegrinelli, V. & Vidal-Puig, A. Adipose Tissue Function and Expandability as Determinants of Lipotoxicity and the Metabolic Syndrome. in *Advances in experimental medicine and biology* **960**, 161–196 (2017).
31. Wyatt, H. R. Update on treatment strategies for obesity. *J. Clin. Endocrinol. Metab.* **98**, 1299–306 (2013).
32. Gupta, V. Glucagon-like peptide-1 analogues: An overview. *Indian J. Endocrinol. Metab.* **17**, 413–21 (2013).
33. Sjöström, L. Review of the key results from the Swedish Obese Subjects (SOS) trial - a prospective controlled intervention study of bariatric surgery. *J. Intern. Med.* **273**, 219–234 (2013).
34. Wiklund, P. The role of physical activity and exercise in obesity and weight management: Time for critical appraisal. *J. Sport Heal. Sci.* **5**, 151–154 (2016).
35. Oakes, N. D. *et al.* A new antidiabetic agent, BRL 49653, reduces lipid availability and improves insulin action and glucoregulation in the rat. *Diabetes* **43**, 1203–10

(1994).

36. Arner, P., Andersson, D. P., Bäckdahl, J., Dahlman, I. & Rydén, M. Weight Gain and Impaired Glucose Metabolism in Women Are Predicted by Inefficient Subcutaneous Fat Cell Lipolysis. *Cell Metab.* **28**, 45–54.e3 (2018).
37. Spalding, K. L. *et al.* Dynamics of fat cell turnover in humans. doi:10.1038/nature06902
38. Hoffstedt, J. *et al.* Long-term Protective Changes in Adipose Tissue After Gastric Bypass. *Diabetes Care* **40**, 77–84 (2017).
39. Gao, H. *et al.* Early B Cell Factor 1 Regulates Adipocyte Morphology and Lipolysis in White Adipose Tissue. *Cell Metab.* **19**, 981–992 (2014).
40. Bahrami-Nejad, Z. *et al.* A Transcriptional Circuit Filters Oscillating Circadian Hormonal Inputs to Regulate Fat Cell Differentiation. *Cell Metab.* **27**, 854–868.e8 (2018).
41. Gökhane, S. Hotamisligil. Inflammation and metabolic disorders. *Nature* **444**, 860–867 (2006).
42. Mathis, D. & Shoelson, S. E. Immunometabolism: an emerging frontier. *Nat. Rev. Immunol.* **11**, 81–83 (2011).
43. Ouchi, N., Parker, J. L., Lugus, J. J. & Walsh, K. Adipokines in inflammation and metabolic disease. *Nat. Rev. Immunol.* **11**, 85–97 (2011).
44. Xu, H. *et al.* Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J. Clin. Invest.* **112**, 1821–1830 (2003).
45. Lee, Y. S., Wollam, J. & Olefsky, J. M. An Integrated View of Immunometabolism. (2018). doi:10.1016/j.cell.2017.12.025
46. Shimobayashi, M. *et al.* Insulin resistance causes inflammation in adipose tissue. *J. Clin. Invest.* **128**, 1538–1550 (2018).
47. Hotamisligil, G. S., Shargill, N. S. & Spiegelman, B. M. Adipose expression of tumor necrosis factor-alpha: direct role in obesity-linked insulin resistance. *Science* **259**, 87–91 (1993).
48. Hotamisligil, G. S., Uysal, K. T., Wiesbrock, S. M. & Marino, M. W. Protection from obesity-induced insulin resistance in mice lacking TNF-alpha function. *Nature* **389**, 610–614 (1997).
49. Gao, Z. & Ye, J. Why do anti-inflammatory therapies fail to improve insulin sensitivity? *Acta Pharmacol. Sin.* **33**, 182–8 (2012).
50. Pedersen, D. J. *et al.* A major role of insulin in promoting obesity-associated adipose tissue inflammation. *Mol. Metab.* **4**, 507–518 (2015).
51. Wernstedt Asterholm, I. *et al.* Adipocyte Inflammation Is Essential for Healthy Adipose Tissue Expansion and Remodeling. *Cell Metab.* **20**, 103–118 (2014).
52. Arner, P. & Langin, D. Lipolysis in lipid turnover, cancer cachexia, and obesity-induced insulin resistance. *Trends Endocrinol. Metab.* **25**, 255–262 (2014).

53. Wedell-Neergaard, A.-S. *et al.* Exercise-Induced Changes in Visceral Adipose Tissue Mass Are Regulated by IL-6 Signaling: A Randomized Controlled Trial. *Cell Metab.* **0**, (2018).
54. Schmid, M. & Guillaume, F. The role of phenotypic plasticity on population differentiation. *Heredity (Edinb)*. **119**, 214–225 (2017).
55. McHill, A. W. *et al.* Later circadian timing of food intake is associated with increased body fat. *Am. J. Clin. Nutr.* **106**, ajcn161588 (2017).
56. Rydén, M. *et al.* The Adipose Transcriptional Response to Insulin Is Determined by Obesity, Not Insulin Sensitivity. *Cell Rep.* **16**, 2317–2326 (2016).
57. Gesta, S., Tseng, Y.-H. & Kahn, C. R. Developmental origin of fat: tracking obesity to its source. *Cell* **131**, 242–56 (2007).
58. Macdougall, C. E. *et al.* Visceral Adipose Tissue Immune Homeostasis Is Regulated by the Crosstalk between Adipocytes and Dendritic Cell Subsets. *Cell Metab.* **27**, 588–601.e4 (2018).
59. Cleal, L., Aldea, T. & Chau, Y.-Y. Fifty shades of white: Understanding heterogeneity in white adipose stem cells. *Adipocyte* **6**, 205–216 (2017).
60. Jeffery, E. *et al.* The Adipose Tissue Microenvironment Regulates Depot-Specific Adipogenesis in Obesity. *Cell Metab.* **24**, 142–150 (2016).
61. Griffin, M. J. *et al.* Early B-cell Factor-1 (EBF1) Is a Key Regulator of Metabolic and Inflammatory Signaling Pathways in Mature Adipocytes. *J. Biol. Chem.* **288**, 35925–35939 (2013).
62. Akerblad, P., Lind, U., Liberg, D., Bamberg, K. & Sigvardsson, M. Early B-cell factor (O/E-1) is a promoter of adipogenesis and involved in control of genes important for terminal adipocyte differentiation. *Mol. Cell. Biol.* **22**, 8015–25 (2002).
63. Cremer, T. & Cremer, C. Chromosome territories, nuclear architecture and gene regulation in mammalian cells. *Nat. Rev. Genet.* **2**, 292–301 (2001).
64. Durani, P., Occleston, N., O’Kane, S. & Ferguson, M. W. J. Avoterm: A Novel Antiscarring Agent. *Int. J. Low. Extrem. Wounds* **7**, 160–168 (2008).
65. Ducker, G. S. & Rabinowitz, J. D. One-Carbon Metabolism in Health and Disease. *Cell Metab.* 1–16 (2016). doi:10.1016/j.cmet.2016.08.009
66. Solini, A., Santini, E. & Ferrannini, E. Effect of short-term folic acid supplementation on insulin sensitivity and inflammatory markers in overweight subjects. *Int. J. Obes.* **30**, 1197–1202 (2006).
67. Cirulli, E. T. *et al.* Profound Perturbation of the Metabolome in Obesity Is Associated with Health Risk. *Cell Metab.* **29**, 488–500.e2 (2019).