Department of Biosciences and Nutrition Karolinska Institutet, Stockholm, Sweden

Role of p21-Activated Kinase 4 in Cell Migration

Zhilun Li

AKADEMISK AVHANDLING

som för avläggande av medicinsk doktorsexamen vid Karolinska Institutet offentligen försvaras på engelska språket i Seminar room 6L at Alfred Nobels Allé 8,

Onsdag den 8 December, 2010, kl. 9.15



Huvudhandledare: Fakultetsopponent:

Prof. Staffan Strömblad Dr. Christoph Ballestrem

Karolinska Institutet University of Manchester, Manchester, UK

Bihandledare: Betygsnämnd:

Dr. Hongquan Zhang Prof. Lars Holmgren, Karolinska Institutet
Karolinska Institutet Prof. Roger Karlsson, Stockholm University

Prof. Ulf Hedin, Karolinska Institutet

Stockholm 2010

Abstract

Cell migration is a cellular process that plays a critical role in various physiological and pathological phenomena, including in cancer metastasis. Understanding at a fundamental level how cancer cells migrate and invade will help to delineate potential targets for the directed development of anti-metastatic therapeutic agents. For example, αν integrins are up-regulated or activated in many migratory cells, and are essential to the processes of wound healing, angiogenesis, and metastasis. Similarly, integrin ανβ5, a vitronectin (VN) receptor, is expressed in most patient carcinoma specimens and is functionally involved in growth factor-induced carcinoma cell migration in vitro and metastasis in vivo. However, the mechanisms of integrin ανβ5-mediated cell migration are not fully understood. In this project, we aimed to identify proteins that interact with the cytoplasmic tail of integrin β 5, and to study their role in cell motility. Firstly, we employed a yeast two-hybrid screening of a mouse embryo cDNA library and thereby identified six proteins specifically interacting with the human integrin \beta 5 cytoplasmic domain. One of the integrin \beta 5interacting proteins was p21-activated kinase 4 (PAK4), which, in addition to its direct interaction with the integrin β5 cytoplasmic tail, also appeared to functionally regulate ανβ5-mediated migration of the human MCF-7 breast carcinoma cells. Importantly, engagement of integrin ανβ5 by cell attachment to VN led to a redistribution of PAK4 from the cytosol to dynamic lamellipodial structures where PAK4 co-localized with integrin ανβ5. Functionally, PAK4 induced integrin ανβ5mediated, but not integrin β1-mediated MCF-7 cell migration, without affecting the cell surface levels of integrin αvβ5.

In addition, we found that PAK4 was activated by cell attachment to VN mediated by the PAK4 binding partner integrin $\alpha\nu\beta5$, and that active PAK4 induced accelerated integrin $\alpha\nu\beta5$ turn-over within adhesion complexes. Accelerated integrin turn-over was the likely cause of additionally observed PAK4-mediated effects, including inhibited integrin $\alpha\nu\beta5$ clustering, reduced integrin to F-actin connectivity and perturbed maturation of cell adhesion complexes. These specific outcomes were ultimately associated with reduced cell adhesion capacity and increased cell motility. We thus demonstrate a novel mechanism deployed by cells to tune cell adhesion levels through the auto-inhibitory regulation of integrin-mediated adhesion.

Furthermore, we identified a unique PAK4-binding membrane-proximal β 5-SERS-motif in the cytoplasmic tail of β 5, and demonstrated a key role for this motif in controlling cell adhesion and migration. We mapped the integrin β 5-binding within PAK4 and observed that PAK4 binding to integrin β 5 was not sufficient to promote cell migration; instead the PAK4 kinase activity was required for PAK4 promotion of cell motility. In fact, PAK4 specifically phosphorylated the integrin β 5 subunit at Ser 759 and Ser 762 within the β 5-SERS-motif. Importantly, point mutation of these two serine residues abolished PAK4-mediated promotion of cell migration, indicating a functional role for these phosphorylations in cell migration.

In conclusion, our results demonstrate that PAK4 interacts with and selectively phosphorylates integrin $\alpha\nu\beta5$ and thereby promotes $\alpha\nu\beta5$ -mediated cell migration, a functional outcome paralleled by a concurrent decrease in total cellular adhesion to VN. Given our finding that PAK4 is activated by $\alpha\nu\beta5$ ligation to VN, these results delineate an auto-inhibitory negative feedback loop that is initiated by cell adhesion to VN. Binding of integrin $\alpha\nu\beta5$ to VN drives translocation and activation of PAK4, leading to phosphorylation of $\alpha\nu\beta5$ and ultimately the limiting of total adhesion between cells and VN and increased cell migration. Thus, our findings provide a new mechanistic characterization of PAK4's role in the functional regulation of integrin $\alpha\nu\beta5$. This knowledge may ultimately be important for understanding vascular permeability, angiogenesis and cancer dissemination.