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IDENTIFICATION OF THE SUSCEPTIBILITY GENES IN TYPE 1 DIABETES AND DIABETIC NEPHROPATHY

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To my family with love

ABSTRACT

Genetic susceptibility plays an important role in the pathogenesis of type 1 diabetes (T1D) and diabetic nephropathy (DN). The present study aims to investigate the association between single nucleotide polymorphisms (SNPs) in candidate genes and DN. Three candidate genes, the intercellular adhesion molecule-1 (*ICAM-1*), adiponectin (*AdipoQ*) and neuropeptide Y (*NPY*), have been selected based upon the approaches: i) from the specific chromosomal regions predicted as important for development of DN by the previous genome wide scan and linkage studies; ii) from the main regulatory steps in the pathways participating in development of diabetic microvascular complications. The studied subjects include 445 Swedish T1D patients with and without DN and 1282 American T1D patients with and without DN selected from the Genetics of Kidneys in Diabetes (GoKinD) study. Genotyping experiments were performed using dynamic allele specific hybridization (DASH). Pyrosequencing and direct sequencing techniques were used for confirmation experiments.

Both *ICAM-1* and *AdipoQ* genes are the strong positional and biological candidates for genetic association study in T1D and DN. We first carried out a genetic association study of the *ICAM-1* gene in Swedish subjects and found that SNP rs5498 E469K(A/G) had a high heterozygous index. Frequencies of the allele G in this SNP were decreased gradually from non-diabetic controls, to T1D patients without DN and the patients with DN. Direct sequencing analysis for the subjects with high heterozygous index was performed, but no duplication in the genomic region around the SNP was found. We further evaluated association between the *ICAM-1* genetic polymorphisms and DN with the GoKinD subjects, and found that the allele G in SNP rs5498 E469K(A/G) was significantly associated with the decreased risk susceptibility of DN in female T1D patients. To evaluate the association of *AdipoQ* genetic polymorphisms with DN in T1D, we have identified 4 binding sites of transcriptional stimulatory protein (SP1) in the *AdipoQ* putative promoter. The allele G of SNP -11377C/G in the promoter altered the sequence for one of SP1 binding sites. This promoter polymorphism and its common diplotype (haplotypic genotype) constructed with this SNP and -11391G/A are found to be associated with DN in female T1D patients among the GoKinD population. Leu7Pro polymorphism in the *NPY* gene is found to be associated with DN and coronary heart disease in Finnish women with T1D. We replicated a genetic association study of this polymorphism in both Swedish and GoKinD subjects. The allele C frequency of Leu7Pro polymorphism in Swedish T1D was higher than in the GoKinD population. This polymorphism was found to be significantly associated with DN in Swedish female T1D patients.

The present study provides evidence that *ICAM-1*, *AdipoQ* and *NPY* genetic polymorphisms are associated with DN in T1D. Genetic susceptibility of the polymorphisms may be influenced by many factors, including gender specificity and ethnic stratification.

Key words: Type 1 Diabetes, Diabetic Nephropathy, Single Nucleotide Polymorphism, Genetic Association

LIST OF PUBLICATIONS

- I. **Ma J**, Mollsten A, Prazny M, Falhammar H, Brismar K, Dahlquist G, Efendic S, Gu HF. Genetic influences of the intercellular adhesion molecule 1 (ICAM-1) gene polymorphisms in development of Type 1 diabetes and diabetic nephropathy.
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- IV. Zhang D, **Ma J**, Brismar K, Efendic S, Gu HF. Genetic Influence of the Adiponectin Promoter Polymorphisms in the Patients with Type 1 Diabetes and Diabetic Nephropathy.
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- V. **Ma J**, Nordman S, Mollsten A, Falhammar H, Brismar K, Dahlquist G, Efendic S, Gu HF. Distribution of neuropeptide Y Leu7Pro polymorphism in patients with type 1 diabetes and diabetic nephropathy among Swedish and American populations.
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LIST OF ABBREVIATIONS

ACE	angiotension-converting enzyme
ACR	urine albumin/urine creatinine ratio
AdipoQ	adiponectin
AGE	advanced glycation end-product
AGT1R	angiotensin II type 1 receptor
AQP1	aquaporin 1
AR	aldose reductase
BCL2	B-cell leukemia/lymphoma 2
BMI	body mass index
CAT	catalase
CTLA-4	cytotoxic T-lymphocyte associated 4
DASH	dynamic allele specific hybridization
DBP	diastolic blood pressure
DM	diabetes mellitus
DN	diabetic nephropathy
DNA	deoxyribonucleic acid
ELMO1	engulfment and cell motility 1
ESRD	end-stage renal disease
FAD	flavin adenine dinucleotide
FFA	free fatty acid
FTO	fat mass and obesity-associated
GAD	glutamic acid decarboxylase
GAPDH	glyceraldehyde phosphate dehydrogenase
GFR	glomerular filtration rate
GH	growth hormone
GoKinD	the Genetics of Kidneys in Diabetes study
GPX1	glutathione peroxidase 1
GWA	genome-wide association
GWS	genome wide scan
HbA1c	hemoglobin A1c
HDL	high-density lipoprotein
HHEX	haematopoietically expressed homeobox
HLA	human leucocyte antigen
HWE	Hardy-Weinberg equilibrium
IA-2	insulinoma-associated protein 2
ICA	islet cell autoantigen
ICAM-1	intercellular adhesion molecule-1
IDDM	insulin-dependent diabetes mellitus
IFIH1	interferon-induced helicase 1
Ig	immuno-globulin
IGF1	insulin-like growth factor 1
INS	insulin
IL2RA	interleukin 2 receptor alpha
KCNJ11	potassium inwardly-rectifying channel, subfamily J, member 11

LAMA4	laminin, alpha 4
LAMC1	laminin, gamma 1
LD	linkage disequilibrium
MHC	major histocompatibility complex
MSVs	multisite variations
NADH	nicotinamide adenine dinucleotide
NPY	neuropeptide Y
OR	odds ratio
PCR	polymerase chain reaction
PKC	protein kinase C
PPi	pyrophosphate
PTPN22	protein tyrosine phosphatase, non-receptor type 22
PVT	plasmacytoma variant translocation
RNA	ribonucleic acid
ROS	reaction oxygen species
SBP	systemic blood pressure
SD	standard deviation
sICAM-1	soluble intercellular adhesion molecule-1
SLC12A3	solute carrier family 12 (sodium/chloride transporters), member 3
SMAD3	mothers against DPP homolog 3
SNP	single nucleotide polymorphism
SP1	stimulatory protein
STZ	streptozotocin
sVCAM-1	soluble vascular adhesion molecule-1
TCA	tricarboxylic acid
TDT	transmission-disequilibrium test
TESS	transcription element search system
TFC7L2	transcription factor 7-like 2
TGFBR2	transforming growth factor, beta receptor II
TGFBR3	transforming growth factor, beta receptor III
TIMP3	tissue inhibitor of metalloproteinase 3
T1D	type 1 diabetes
T2D	type 2 diabetes
UAER	urinary albumin excretion rates
USF1	upstream transcription factor 1
VCAM	vascular cell adhesion molecule-1
VEGF	vascular endothelial growth factor

1 INTRODUCTION

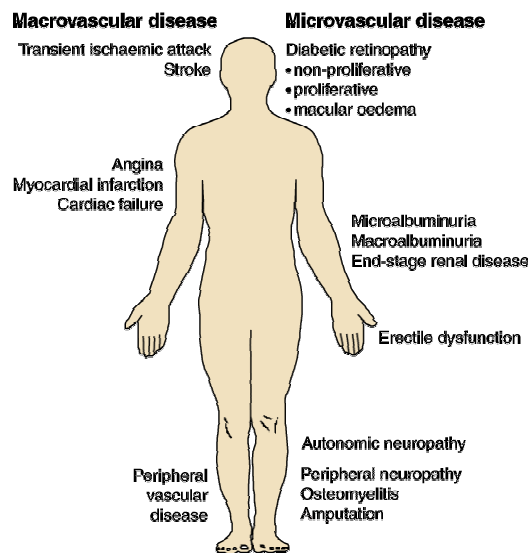
1.1 TYPE 1 DIABETES

Diabetes mellitus (DM) includes two major types: type 1 and type 2 diabetes. Type 1 diabetes (T1D) patients are characterized by low or absent levels of circulating endogenous insulin and thereby dependent on injected insulin to prevent ketosis and sustain life. T1D is an autoimmune disorder in which the immune system attacks the pancreas, rendering it unable to produce insulin. Auto-antibodies against specific islet cell antigens (ICA, GAD, IA-2 and insulin) are presented in the majority of patients at the onset of T1D. Furthermore, certain human leukocyte antigens (HLA, DR3 and/or DR4) are strongly associated with the disease (Nerup et al., 1974; Palmer et al., 1983; Baekkeskov et al., 1990; Payton et al., 1995; Di et al., 2007). T1D onsets predominantly in youth but can occur at any age. Although the etiology of T1D has not been understood fully, evidence indicates that genetic and environmental factors are involved in the development of this disease (Adeghate et al., 2006).

1.2 DIABETIC MICRO-VASCULAR COMPLICATIONS

Patients with T1D often develop diabetic vascular complications (Figure 1). The diabetic macro-vascular complications include coronary artery, cerebrovascular and peripheral vascular diseases (Bate, Jerums, 2003). The diabetic micro-vascular complications consist of diabetic retinopathy, nephropathy and neuropathy, which are caused by damage to affecting capillaries and small blood vessel in the eyes, kidneys and nerves (Bate, Jerums, 2003).

Figure 1. Diabetic vascular complications



From Bate and Jerums 2003

Chronic hyperglycemia is the most important cause for development of micro-vascular complications (Bate, Jerums, 2003). There are several hypotheses about the direct toxic effects of glucose, which causes diabetic micro-vascular complications: activation of protein kinase C (PKC), increased advanced glycation end-product (AGE) formation (Parving, 2001); increased flux through hexosamine pathway (Wild et al., 2004) and

increased polyol pathway. In particular, hyperglycaemia and increased free fatty acid (FFA) levels result in the production of reaction oxygen species (ROS), such as the superoxide anion, which causes oxidative stress. The generation of oxidative stress results in endothelial dysfunction and a reduction in flow-mediated vasodilation, which may be central to the development of not only micro- but also macro-vascular complications. Hyperglycaemia increases the flux of glucose through the tricarboxylic acid (TCA) cycle, leading to increased levels of the electron donors nicotinamide adenine dinucleotide (NADH) and flavin adenine dinucleotide (FADH₂). The increased mitochondrial membrane potential inhibits electron transport at complex III and also extends the half-life of free radical intermediates, which increases the production of superoxide from O₂. Central to this activation is the inhibition of glycerolaldehyde-3-phosphate dehydrogenase (GAPDH) by superoxide. This causes an over-utilisation of upstream glycolytic intermediates.

In addition, diabetes and hypertension frequently coexist, and hypertension accelerates the course of micro-vascular complications in the patients with T1D (Schutta, 2007). The development of proteinuria is paralleled in most cases by a gradual rise in systemic blood pressure (SBP).

1.2.1 Diabetic nephropathy

Diabetic nephropathy (DN) was discovered by British physician Clifford Wilson (1906-1997) and Germany-born American physician Paul Kimmelstiel (1900-1970), and description of the disease was published for the first time in 1936. DN, also known as Kimmelstiel-Wilson syndrome and intercapillary glomerulonephritis, is a progressive kidney disease caused by angiopathy of capillaries in the kidney glomeruli. It is characterized by nephrotic syndrome and nodular glomerulosclerosis. Due to longstanding diabetes, the kidney becomes damaged and more protein than normal collects in the urine. As the disease progresses, more of the kidney is destroyed. Over time, the ability of kidney to function starts to decline, which eventually leads to chronic kidney failure.

The cumulative incidence of overt DN in T1D is approximately 20-30% (Marshall, 2003). Diabetes is the leading cause of treated end-stage renal disease (ESRD), accounting for almost half of the new cases each year (Jones et al., 2005). Among European Americans with T1D approximately 30% develop severe DN that leads to ESRD (Krolewski et al., 1996).

1.3 GENETIC STUDIES OF DIABETIC NEPHROPATHY IN TYPE 1 DIABETES

It is widely accepted that the development and progression of DN result from a combination of genetic predisposition, metabolic and hemodynamic abnormalities, and environmental factors (Brownlee, 2001; He, King, 2004; Chow et al., 2005; Jones et al., 2005; Liu, Freedman, 2005; Ng, Krolewski, 2005; Ritz, 2005). There are substantial evidences for a genetic contribution to DN. For instance, in a study of 110 families who

had multiple siblings with T1D, the cumulative risk of DN was 71.5% in those who had a sibling with DN, but just 25.4% for those in which siblings did not show the complications ($P < 0.001$) (Quinn et al., 1996). Epidemiologic studies have shown that DN is strongly clustered in families and that race has a major effect on DN susceptibility and rate of progression (Seaquist et al., 1989; Iyengar et al., 2002; Ng, Krolewski, 2005). Therefore, genetic study may provide useful information for better understanding the pathogenesis of DN.

1.3.1 Genome wide scan and linkage study

Genome wide scan (GWS) and linkage analysis approach has been used to reveal evidence for DN susceptibility loci in chromosomal regions. With this research approach, polymorphic genetic markers for instance micro-satellites with known positions and spaced in smaller intervals along the entire genome are used. The markers distributed across the genome are genotyped in large family pedigrees. Markers that occur more commonly in family members with DN may indicate the presence of a nearby susceptibility loci along the same chromosome. In the recent years, several GWS and linkage analyses have been reported. For instance, chromosomal region 3q is found to link with DN and T1D (Mein et al., 1998; Moczulski et al., 1998; Chistiakov et al., 2004; Osterholm et al., 2007). The linkage peak in this chromosomal region is also observed in T2D with DN (Bowden et al., 2004). Furthermore, two microsatellites D10S558 and D10S1435 in chromosome 10 are found to be significantly associated with DN in T1D (McKnight et al., 2006). These evidences of chromosomal regions linked to DN and T1D provide information for further investigations, including not only fine mapping and positional cloning, but also positional candidate gene single nucleotide polymorphism (SNP) genotyping analyses.

1.3.2 Candidate gene-based association study

Candidate gene-based association studies have been the most common approach employed to identify susceptibility genes for DN. The strategy is the use of either population-based or family-based approaches. Many factors may contribute to variable association studies: multiple hypothesis testing, ethnic stratification, population-specific linkage disequilibrium (LD) between markers and causal variants, inadequate statistical power, publication bias, and gene-gene and gene-environmental interactions. The population-based approach is used to determine whether gene variants are found more or less commonly in patients with DN (cases) than in patients without DN (controls). The family-based approach, such as the transmission-disequilibrium test (TDT), is used to test the frequency of transmission of variants from heterozygous parents to their offsprings in comparison with the expected 50:50 ratio (Spielman et al., 1993). However, TDT study requires analysis of both parents. In the parents of patients with DN, it is difficult to recruit sufficient family trios to provide adequate power (Conway et al., 2006). Using the approach of TDT, Ewens et al. have tested 115 candidate genes in 72 T1D families of European descent. Of the candidates, several genes, including aquaporin 1 (*AQP1*), B-cell leukemia/lymphoma 2 (*BCL2*), catalase (*CAT*), glutathione peroxidase 1 (*GPX1*), insulin-like growth factor 1 (*IGF1*), laminin, alpha 4 (*LAMA4*),

laminin, gamma 1 (*LAMC1*), mothers against DPP homolog 3 (*SMAD3*), transforming growth factor, beta receptor II (*TGFBR2*), transforming growth factor, beta receptor III (*TGFBR3*), tissue inhibitor of metalloproteinase 3 (*TIMP3*) and upstream transcription factor 1 (*USF1*), are found to be associated with DN (Ewens et al., 2005).

A number of genetic association studies have been reported. Aldose reductase (*AR*) is the first and rate-limiting enzyme in the pathway that catalyses the reduction of glucose to sorbitol. Several studies have indicated that the polymorphisms of a micro-satellite located at 5' strand of the aldose reductase gene are associated with DN. Further association studies have recently demonstrated that a promoter polymorphism C-106T in this gene confers genetic susceptibility to DN in both T1D and T2D (Moczulski et al., 2000; Neamat-Allah et al., 2001). Angiotensin II type 1 receptor (*AGT1R*) is expressed in the vascular smooth muscle cells of the afferent and efferent arterioles of the kidney, glomerular mesangial cells, proximal tubule cells of the kidney and adrenal glomerulosa cells. The polymorphism C1166A of the *AGT1R* gene is strongly associated with renal hemodynamic function in early T1D (Miller et al., 2000). Vascular endothelial growth factor (*VEGF*) is a potent angiogenic and vascular permeability factor and is implicated in DN. An insertion/deletion C-460 polymorphism in the promoter region of the *VEGF* gene has been found to be associated with promotion of neovascularization (Ray et al., 2004). The relationship between the angiotensin-I-converting enzyme (*ACE*) insertion/deletion polymorphism and DN in T1D was established (Rudberg et al., 2000; Hadjadj et al., 2001).

1.3.3 Genome wide association study

With the improvement of genotyping technologies and the exponentially growing number of SNPs, case-control genome-wide association (GWA) studies have become a useful tool for identification of the susceptibility genes in diabetes and DN. A recent GWA study on seven diseases, including T1D, has been performed, and the results have demonstrated association between T1D and six chromosomal regions 12q24, 12q13, 16p13, 18p11, 12p13 and 4q27 (The Wellcome Trust Case Control Consortium, 2007). Several genes, including *ERBB3* in chromosome 12q13, *C12orf30* in 12q24, *KIAA0350* in 16p13 and *PTPN2* in 18p11, are found to be associated with T1D (Todd et al., 2007). Furthermore, Hakonarson et al. have also performed a GWA study and indicated that the *KIAA0350* gene might be involved in the pathogenesis of T1D (Hakonarson et al., 2007). In addition, the solute carrier family 12 (sodium/chloride transporters), member 3 (*SLC12A3*) and engulfment and cell motility 1 (*ELMO1*) genes are found to be associated with DN in T2D (Shimazaki et al., 2005; Maeda et al., 2007). Hanson et al. have recently identified that the plasmacytoma variant translocation (*PVT1*) gene is a susceptibility gene for ESRD in T2D (Hanson et al., 2007). Recent successes in identifying the susceptibility genes in T1D, T2D and DN in T2D suggest that GWA is an useful approach to identify the susceptibility gene for DN in T1D as well.

2 THE PRESENT STUDY

2.1 AIMS

The research project is focused on genetic association study between the candidate genes and DN in T1D. The specific aims are:

- To evaluate the association between SNPs in the intercellular adhesion molecule-1 (*ICAM-1*) gene and DN in T1D;
- To investigate the genetic influence of adiponectin (*AdipoQ*) variation in the development of DN and T1D;
- To study the association of neuropeptide Y (*NPY*) Leu7Pro polymorphism and DN in T1D.

The results may provide useful information for a better understanding pathogenesis of DN in T1D.

2.2 SELECTION OF CANDIDATE GENES

We have selected the candidate genes for our study based upon two approaches:

- i) From the specific chromosomal regions predicted as important for development of DN by the previous genome wide scan and linkage studies.
- ii) From the main regulatory steps in the pathways participating in the development of endothelial dysfunction and micro-angiopathy in the patients with DN.

2.2.1 Intercellular adhesion molecule-1

ICAM-1 is a 90-kD cell surface glycoprotein of the immuno-globulin (Ig) superfamily involved in the firm attachment of leukocytes to endothelium (Staunton et al., 1988; van, van, 1996). It is normally expressed at low levels on the surface of arterial endothelial cells. But, its expression can be induced by multiple factors, including inflammatory cytokines, reactive oxygen species and shear stress (Koulu et al., 2004; Nordman et al., 2005). Particularly, ICAM-1 expression at both mRNA and protein levels is found to be increased by hyperglycemia (Kado et al., 2001; Omi et al., 2002).

The *ICAM-1* gene is located in chromosome 19p13. GWS has predicted that the T1D susceptibility genes may reside in this chromosomal region (Mein et al., 1998). The levels of plasma adhesion molecules are elevated either in T1D or T2D patients with DN and relate to renal dysfunction (Lampeter et al., 1992; Hirata et al., 1998; Clausen et al., 2000; Mohamed-Ali et al., 2001; Guler et al., 2002; Nelson et al., 2005). Vascular cell adhesion molecule-1 (VCAM) is elevated in the setting of moderately decreased glomerular filtration rate (GFR) in T2D patients and may be the link between renal insufficiency and increased risk for cardiovascular events in American population (Lin et al., 2006). Soluble vascular adhesion molecule-1 (sVCAM-1) and intercellular

adhesion molecule-1 (sICAM-1) are increased in T1D patients with DN (Nelson et al., 2005). Furthermore, it has been proposed that the increased levels of plasma adhesion molecules may be the sign of pre-clinical T1D in children (Toivonen et al., 2004).

The experimental studies using T1D animal models demonstrate that the increased ICAM-1 expression accompanies the progression of T1D (Balasa et al., 2000) and DN (Matsui et al., 1996; Sugimoto et al., 1997). Furthermore, ICAM-1 expression in glomeruli and tubulointerstitium is significantly elevated in the streptozotocin (STZ)-diabetic rats (Qi et al., 2006). Increased mesangial matrix, glomerular hypertrophy, and albuminuria are found to significantly suppressed in diabetic ICAM-1(-/-) mice compared with diabetic ICAM-1(+/+) mice (Okada et al., 2003). Thus, up-regulation of kidney ICAM-1 appears to be a common response to diabetic renal injury. Therefore, the *ICAM-1* gene is a strong positional and biological candidate for genetic association study in T1D and DN.

2.2.2 Adiponectin

Adiponectin is a hormone exclusively secreted by adipose tissue. Serum adiponectin concentrations are decreased in the patients with T2D and in the subjects with obesity (Arita et al., 1999; Statnick et al., 2000). In contrast, serum adiponectin levels are increased in the patients with T1D (Imagawa et al., 2002; Diez, Iglesias, 2003; Lindstrom et al., 2006) and also in T1D patients with micro-angiopathy (Frystyk et al., 2005; Hadjadj et al., 2005) and DN (Saraheimo et al., 2005). The mechanism behind an increase of circulating adiponectin in DN is not fully understood. The recent report indicates that renal insufficiency and tubular injury possibly play a contributory role in increases of serum and urinary adiponectin levels in overt DN (Fujita et al., 2006).

The adiponectin gene is encoded as *AdipoQ* (adipocyte C1q and collagen domain containing). The alternative names for this gene are *GBP28*, *ACRP30*, *APM1* and *ACDC*. The *AdipoQ* gene is located on chromosome 3q27, within the chromosomal region linked with T2D, obesity and hypertension (Kissebah et al., 2000; Vionnet et al., 2000; Francke et al., 2001; Zacharova et al., 2005), and also with DN in T1D and T2D (Imperatore et al., 1998; Moczulski et al., 1998). Several reports have indicated that the *AdipoQ* genetic polymorphisms are associated with T2D (Hara et al., 2002; Vasseur et al., 2002; Gu et al., 2004; Vasseur et al., 2005; Schwarz et al., 2006). In order to investigate the susceptibility of the *AdipoQ* genetic polymorphisms in T1D and DN, we have conducted a genetic association study with Swedish T1D patients with and without DN. A recent study has suggested that the *AdipoQ* promoter polymorphisms may confer the risk susceptibility to the development of DN in T1D patients in Danish but not in French and Finnish populations (Vionnet et al., 2006). We thus further evaluate the association between the *AdipoQ* promoter polymorphisms and DN with patient material from the Genetics of Kidneys in Diabetes (GoKinD) study.

2.2.3 Neuropeptide Y

NPY is a 36-amino acid polypeptide that is present in the central and peripheral nervous system, several peripheral organs and plasma (Jamal et al., 1991; Wang et al., 1994; Bennet et al., 1996; Myrsen-Axcrona et al., 1997). NPY has multiple functions and mainly plays a role in the regulation of satiety, ingestive behaviors, energy balance and energy expenditure (Billington et al., 1994; Levine et al., 2004). NPY also stimulates lipoprotein lipase activity in the adipose tissue (Billington et al., 1991) and regulates insulin release (Moltz, McDonald, 1985).

NPY exists at a high concentration in the hypothalamus and increased hypothalamic NPY and noradrenaline overflow may contribute to diabetic hyperphagia (Morris, Pavia, 2004). NPY release is found to be increased from the hypothalamus in STZ-diabetic rats (Gozali et al., 2002; Morris, Pavia, 2004). Furthermore, NPY has been shown to regulate renal blood flow (Bischoff, Michel, 1998). Stimulation of the NPY receptors in the kidney, using active agonists, may decrease GFR, aldosterone concentration and plasma renin activity, but increase sodium excretion in the kidney (Playford et al., 1995). NPY is also a potent vasoconstrictor of renal arteries (Michel et al., 1992) and decreases renal blood flow and glomerular filtration (Echtenkamp, Dandridge, 1989). Interestingly, the Leu7Pro polymorphism in the *NPY* gene is found to be associated with high blood lipid concentrations and accelerated rate of atherosclerosis as well as diabetic retinopathy. The subjects with Leu7/Pro7 genotype of this polymorphism had 54% higher maximal increases in the plasma GH concentrations compared to the subjects carrying with Leu7/Leu7 genotype (Kallio et al., 2001). NPY may play a role in the pathogenesis of diabetes and DN.

3 SUBJECTS AND METHODS

3.1 SUBJECTS

Patient material in the present study consists of two sets of subjects, i.e. Swedish and the GoKinD populations. T1D patients were diagnosed according to the World Health Organization criteria. The studies were approved by the local ethics committees, and informed consent was obtained from all participants.

3.1.1 Swedish subjects

This set of subjects includes 445 (male 210/ female 235) Swedish T1D patients (Table 1). Most of T1D patients were collected from Department of Endocrinology, Metabolism and Diabetes, Karolinska University Hospital, while 146 T1D patients were collected from Umeå University Hospital. T1D patients had at least 10 years of diabetes duration. All patients had the urinary albumin excretion rate (UAER) measured in at least two consecutive overnight urine samples. The patients with UAER <20 µg/min were considered as being free from DN (n=249). The patients with UAER of 20-200 µg/min and of >200 µg/min in at least 2 out of 3 consecutive overnight samples were considered as microalbuminuria (incipient nephropathy, n=105) and macroalbuminuria (overt nephropathy, n=91. 22 of them were ESRD), respectively. The patients without DN had never been treated with ACE inhibitors when UAER was measured. In addition, 187 non-diabetic healthy individuals, with no family history of diabetes, were also included. All subjects were Swedish Caucasians.

Table 1. Swedish subjects

	T1D without DN			T1D with DN		
	All	Male	Female	All	Male	Female
N	249	117	132	196	93	103
Age (years)	44±12	45±12	43±12	46±12	49±12	43±12
Duration (years)	30±10	29±10	31±10	29±11	34±13	34±11
BMI (kg/m ²)	24.6±3.1	24.9±2.2	24.4±3.6	25.8±3.7	25.7±3.7	25.9±3.6
SBP (mmHg)	127±15	129±14	125±16	138±21	140±22	136±18
DBP (mmHg)	74±7	75±7	72±7	78±10	79±10	77±10
HbA1c (%)	7.0±1.1	6.9±1.0	7.2±1.2	7.4±1.4	7.2±1.4	7.6±1.3

All data=means±SD.

3.1.2 The GoKinD subjects

This set of subjects was selected from the GoKinD study (Table 2) and collected by the Juvenile Diabetes Research Foundation in collaboration with the Joslin Diabetes Center and George Washington University, and the United States Centers for Diabetes Control and Prevention (Mueller et al., 2006). Totally, 620 T1D patients without DN and 662 patients with DN were included. Among the patients, 1177 (91.8%) were of European descent, while 105 (8.2%) patients were Americans of Black, Asian, Hispanic or Indian descent. T1D patients were diagnosed before 31 years of age. Treatment with insulin was instituted within one year of diagnosis, and has been uninterrupted since then. The

patients with DN had T1D for at least 10 years and had persistent proteinuria or ESRD (not due to condition other than diabetes). Persistent proteinuria was defined as 2 out of 3 tests positive for albuminuria (at least 1 month apart), i.e. dipstick (Albustix or Multistix) at least 1+ or Urine Albumin/Urine Creatinine Ratio (ACR) value exceeding 300 μg albumin/mg of urine creatinine. Patients without DN had persistent normalalbuminuria despite duration of T1D for at least 15 years and had never been treated with ACE inhibitors. Persistent normoalbuminuria was defined as at least 2 out of 3 of ACR measurements (at least 1 month apart) in random urine specimens being less than 20 μg albumin/mg of urine creatinine. If 3 ACR measurements were needed, the highest must also be less than 40 μg albumin/mg of urine creatinine. ESRD is recognized as the patients need chronic dialysis or kidney transplant treatment to maintain their life. The onset of ESRD is defined as the date of the first dialysis or kidney transplant, whichever occurred first. Other kidney diseases in cases were excluded (Mueller et al., 2006).

Individuals were excluded from the study if they did not meet the inclusion criteria just described or if any of the following exclusion criteria were met: unable or unwilling to give informed consent; unable to communicate with staff; major psychiatric disorder such as schizophrenia; exclusion in relation to medication: any antihypertensive medication for controls; infectious disease: self-reported HIV positivity and active tuberculosis; pregnant women (although they may be reconsidered 3 months after delivery); other kidney disease due to condition other than diabetes.

Table 2. The GoKinD subjects

	T1D without DN			T1D with DN		
	All	Male	Female	All	Male	Female
N (M/F)	620	251	369	662	350	312
Age (years)	40 \pm 8	40 \pm 8	40 \pm 9	44 \pm 7	44 \pm 6	43 \pm 7
Duration (years)	26 \pm 8	26 \pm 8	26 \pm 8	31 \pm 8	32 \pm 8	31 \pm 8
BMI (kg/m^2)	26.1 \pm 4.3	26.6 \pm 3.7	25.6 \pm 4.6	25.8 \pm 5.4	26.1 \pm 4.9	25.5 \pm 6.0
SBP (mmHg)	118 \pm 12	122 \pm 12	116 \pm 12	132 \pm 19	134 \pm 18	130 \pm 20
DBP (mmHg)	71 \pm 8	74 \pm 8	70 \pm 8	74 \pm 11	76 \pm 11	72 \pm 11
HbA1c (%)	7.5 \pm 1.1	7.4 \pm 1.1	7.5 \pm 1.1	7.4 \pm 1.9	7.4 \pm 1.7	7.4 \pm 2.1

All data=means \pm SD.

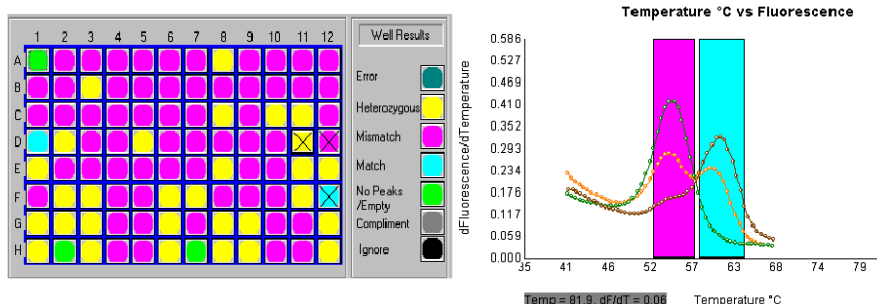
3.2 METHODS

3.2.1 Dynamic allele specific hybridization

Dynamic allele specific hybridization (DASH) is a high throughput genotyping method for scoring SNPs and small insertion/deletions. This method is based upon hybridization with a specific probe. To most readily interpret DASH results, the negative first derivatives can be shown on the computer screen. The sample mismatched to the probe by one base pair produces a single lower temperature peak. The sample matched to the probe produces a single high-temperature peak. The heterozygous sample undergoes two phases of denaturation (one for each allele present)

and therefore produces two peaks in the negative first derivative (Figure 2) (Howell et al., 1999).

Figure 2. An example from DASH genotyping experiments

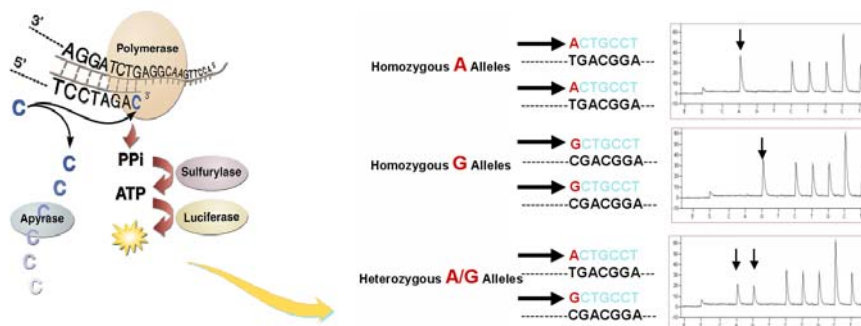


From the paper I in the present study

3.2.2 Pyrosequencing

Pyrosequencing is based on the detection of released pyrophosphate (PPi) during synthesis of DNA sequence. The synthesis starts with a nucleic acid polymerization reaction in which inorganic PPi is released as a result of nucleotide incorporation by DNA polymerase. The released PPi is subsequently converted to ATP by ATP sulfurylase, which provides the energy to luciferase to oxidize luciferin and generate light. Because the added nucleotide is known, the sequence of the template can be determined (Figure 3). This method can be used not only for sequencing analysis in short DNA or RNA templates but also for SNP genotyping (Ronaghi et al., 1998).

Figure 3. Pyrosequencing and SNP genotyping



From <http://www.pyrosequencing.com/DynPage.aspx?id=8583&mn1=1366&mn2=1370>

3.2.3 Direct sequencing analysis

Segmental duplications (duplicons) with >90% similarity between the copies comprise at least 5% of the human genome, which may cause specific allelic and genotypic diversities such as high heterozygous index in complex diseases (Venter et al., 2001; Shaw, Lupski, 2004). In order to detect the possible duplicons in the sequences that the studied SNPs reside, the direct sequencing analysis with ABI BigDye terminator cycle sequencing kit and protocol was used. The kit and protocol are suitable for performing fluorescence-based cycle sequencing reactions on large single-stranded or double-

stranded DNA templates, and on polymerase chain reaction (PCR) fragments and on large templates.

3.2.4 Bioinformatics

Sequences of the studied genes are recorded in the GenBank of NCBI. The studied SNPs are selected from public SNP databases, including dbSNP (<http://www.ncbi.nlm.nih.gov/SNP/>) and HGVbase (<http://hgvdbase.cgb.ki.se/>) according to their types, locations, validation status in HapMap (<http://www.hapmap.org/>) and information from the previous reports.

For each studied SNP, the surrounding approximately 50 bp sequences in 5'- or 3'-direction were examined for repeats and duplicated sequences (<http://repeatmasker.genome.washington.edu/cgi-bin/RepeatMasker>) and blasted in human genome sequences (<http://www.ncbi.nlm.nih.gov/blast>).

To design DASH assays, the possible structures of PCR products for DASH genotyping experiments were analyzed with mFold (version 3.1; <http://www.bioinfo.rpi.edu/applications/mfold/old/dna/>). To search for the possible transcriptional binding factors in the sequence of the AdipoQ promoter, Transcription Element Search System (TESS) is used for predicting transcription factor binding sites in DNA sequences (<http://www.cbil.upenn.edu/cgi-bin/tess/tess>).

3.2.5 Hardy-Weinberg equilibrium test

Deviation from the Hardy-Weinberg equilibrium (HWE) for alleles in each SNP was assessed using the appropriate Chi-square distributed statistics.

In the general population the genotype distribution of a SNP should always be in HWE. The law of HWE was founded in 1908, by Godfrey H. Hardy and Wilhelm Weinberg and is basically stating that in a randomly mating population the relative frequencies of heterozygotes and homozygotes are mathematically related and that allele frequencies will remain constant from generation to generation provided that none of the alleles is under positive or negative selection. Several criteria must be fulfilled for a SNP to be in HWE, namely random mating, absence of selection and gene flow, a large enough population, no change of mutation frequency and finally no chromosomal rearrangements.

HWE can easily be tested by using the equation: $(p+q)^2 = p^2+2pq+q^2 = 1$, where p is the allele frequency of allele 1 and q is the allele frequency of allele 2 and $2pq$ is the frequency of heterozygotes. The observed allele frequencies are calculated and used to find the expected genotype frequencies. The observed and expected genotype frequencies are compared using the Chi-square test. A P value that is <0.05 , indicates

that a polymorphism is not in HWE, which means that the observed and expected genotype distribution values differ significantly.

If a SNP does not follow the HWE law in the patient group it could be interpreted to strengthen the evidence of association between the SNP and the phenotype because of increased frequency of a certain genotype that is associated with the phenotype. On the other hand, HWE must always be obtained in the control group when performing association studies. One other explanation why a SNP does not follow the HWE law could be complex SNP related sequence variations, a recently discovered form of genome polymorphisms, called multisite variations (MSVs) that usually do not follow the HWE. An MSV can masquerade as a SNP, but may in reality be a duplicon marker that arises from segmental duplications of a sequence (Fredman et al., 2004).

3.2.6 Single marker association analysis

To detect single marker association between cases and controls, two models are tested comparing either allele frequencies in 2×2 contingency tables or genotypes in 3×2 contingency tables. If heterozygous individuals confer significant risk to the carriers with non-risk homozygosity, recessive mode of inheritance is rejected. Additive and dominant modes are considered to be used. If homozygous individuals confer significant risk to heterozygous carriers, additive and dominant modes are not suitable. The recessive mode is then used. Statistical powers are calculated using a software of PowerSampleSize (PS version 2.1.31, <http://biostat.mc.vanderbilt.edu/twiki/bin/view/Main/PowerSampleSize>). Tests for association between genotypes and quantitative traits are performed using Kruskal–Wallis analysis of ranks for traits with non-normal distributions, or alternatively, ANOVA for normally distributed traits. For association estimation, odds ratios (OR) and 95% confidence intervals (CI) are estimated with unconditional logistic regression models between cases and controls. All statistical analyses are performed using the BioMedical Data Program (BMDP) version 1.12 and/or STATISTICA version 7.0.

3.2.7 Linkage disequilibrium and haplotype analysis

SNPs are positioned on the same chromosome can segregate together more often than expected and are then said to be in LD. LD can be calculated and is dependent on allele frequencies as well as of recombination. The two most common measures of LD are Lewontin's LD parameter D' , which is a measure of recombination, and r^2 , which is a measure of correlation. When $D'=1$ the two SNPs are in total LD, meaning that there is no recombination and when $r^2=1$ the allele frequencies of the SNPs are equal and there is no recombination meaning that the SNPs are in total LD. The two LD measures do not have to follow each other, i.e. $D'=1$, but $r^2=0.6$ for the same two SNPs means that the SNPs segregate together more often than expected, but the allele frequencies are not equal. LD is an association between polymorphisms at the population level, where alleles in close proximity of each other are often inherited together, since they originate from a common chromosome. A polymorphism can be in LD with another

polymorphism on the same chromosome, which is associated with a trait and thereby act as a marker for the particular disease.

The estimation of pair-wise LD values and haplotype frequencies was performed using Arlequin program version 2.0 (<http://lgb.unige.ch/arlequin/>). Haplotype prevalence at >5% was further used for haplotype association analysis.

4 RESULTS AND DISCUSSION

4.1 ASSOCIATION STUDIES OF SNPs IN THE ICAM-1 GENE AND DN IN T1D (PAPERS I AND II)

We carried out a genetic association study of the *ICAM-1* gene in T1D. A total of 8 SNPs, including these two non-synonymous polymorphisms rs5498 E469K(A/G) and rs1799969 R241G(A/G) in the *ICAM-1* gene, were validated with 32 DNA samples from Swedish subjects. Of the studied SNPs, three had allele frequencies at <1% and were therefore excluded from further analyses. Genotyping results indicated that SNPs rs281432 (C/G) and rs5498 E469K(A/G) were significantly associated with T1D. Frequencies of the C allele in SNP rs281432 (C/G) and the A allele in SNP rs5498 E469K(A/G) increased gradually from non-diabetic controls, to T1D patients without DN and T1D patients with DN among Swedish T1D patients. However, no significant association of these two polymorphisms with DN was detected. We then evaluated the association between the *ICAM-1* genetic polymorphisms and DN in the GoKinD population. We found that the significant association of heterozygosity and allele G positivity in SNP rs5498 E469K(A/G) with DN in female T1D patients. The allele G of this polymorphism might confers the decreased risk susceptibility to the development of DN.

Rs5498 E469K(A/G) resides in the 5th immunoglobulin-like domain of ICAM-1. This domain may play a role in an immunodominant epitope of B lymphocytes and dendritic cells (Joling et al., 1994). Furthermore, a recent study has demonstrated that this SNP affects ICAM-1 mRNA splicing pattern and TPA-induced apoptosis (Iwao et al., 2004). Interestingly, the genotype distribution of SNP rs5498 E469K(A/G) was in HWE, and this polymorphism had high heterozygous index, which was not reported previously. By communication with the previous reporters, we found that the phenomenon of high heterozygous index of this SNP existed in all populations studied. Segmental duplications (duplicons) in the human genome may cause specific allelic and genotypic diversities such as high heterozygous index in complex diseases (Venter et al., 2001; Shaw, Lupski, 2004). To ascertain whether SNP rs5498 E469K(A/G) is involved in a duplicon, we have confirmed the genotyping data by using another method called pyrosequencing, and also performed direct sequencing analyses. Results indicated that no duplicon was involved in the genomic sequences around this SNP. The present study thus provide evidence that the heterozygous genotype of SNP rs5498 E469K(A/G) in the *ICAM-1* gene may have genetic influence in the pathogenesis of DN and T1D.

4.2 GENETIC ASSOCIATION ANALYSES OF THE ADIPOQ GENE IN T1D AND DN (PAPERS III AND IV)

We first performed validation for 13 SNPs in the *AdipoQ* gene with Swedish subjects. Of the studied SNPs, 3 non-synonymous polymorphisms in exon 3, including G84R(G/C), G90S(G/A) and Y111H(T/C), had low allele frequencies (<3%). These 3

polymorphisms were excluded for further analysis. We then found that SNPs +45G15G(T/G) and +276(G/T) were in a LD block. These two polymorphisms and their common haplotypes were associated with T1D. Comparison analysis among T1D patients carrying three different genotypes in SNP +276G/T indicated that the GG carriers had lower levels of serum adiponectin. However, no significant association of the *AdipoQ* genetic polymorphisms with DN was found most likely due to the limitation of sample size.

A recent study demonstrated that an *AdipoQ* promoter polymorphism -11391G/A was associated with DN in T1D patients among Danish, but not French, Finnish populations (Vionnet et al., 2006). We thus further studied the association between *AdipoQ* promoter polymorphisms, including -11377C/G, -11391G/A and -11426A/G, and DN with the GoKinD subjects. We identified 4 binding sites of transcriptional stimulatory protein (SP1) in the *AdipoQ* promoter, and found that the allele G of SNP-11377C/G alters the sequence for one of SP1 binding sites. This polymorphism was significantly associated with DN in female T1D patients. Additionally, two promoter polymorphisms -11377C/G and -11391G/A are located in a LD block and the common diplotype (haplotypic genotype) H1/H1, constructed with these two polymorphisms was found to be significantly associated with DN in females T1D patients.

Genetic variation in the promoter may alter the binding sequences for transcriptional elements and consequently influence gene regulation and expression. It has been demonstrated that SP1 belongs to the SP family of transcriptional factors and is ubiquitously expressed in mammalian cells (Krikun et al., 2000; Li et al., 2004). Barth et al. have previously reported that SP1 binding activity is enhanced during adipocyte differentiation, and has stimulatory effects on the *AdipoQ* promoter activity (Barth et al., 2002), but number and location of SP1 binding sites are unknown. Bouatia-Naji et al. have recently investigated the functional properties of SNP -11377C/G with the transfection approach and electrophoretic mobility shift assays. The data demonstrated that the allele G had lower, but not significantly, transcriptional activity than the “wild-type” allele C (Bouatia-Naji et al., 2006). Taking together, we suggest that the promoter polymorphisms in the *AdipoQ* gene may have genetic and biological influence related to the development of DN in T1D. However, we still look forward to having the possibility to examine the serum adiponectin levels in T1D patients with or without DN according to the genotypes of the *AdipoQ* promoter polymorphisms.

4.3 DISTRIBUTION OF NPY LEU7PRO POLYMORPHISM IN T1D PATIENTS IN SWEDISH AND GOKIND POPULATIONS (PAPER V)

NPY Leu7Pro (T1128C) is a non-synonymous polymorphism and the allele C may have an impact on NPY processing on circulating NPY levels, on growth hormone secretion, and on serum cholesterol and triglyceride levels (Karvonen et al., 1998; Kallio et al., 2001; Mattevi et al., 2002; Pihlajamaki et al., 2003; Karvonen et al., 2006). Furthermore, this polymorphism distributes a geographical north to south

gradient of decreasing frequency (Ding, 2003), suggesting that it may be a population-specific causal variant.

We have genotyped this polymorphism with the subjects from both Swedish and GoKinD populations. Results indicated that the allele C frequency of *NPY* Leu7Pro polymorphism in Swedish T1D was significantly higher than in the GoKinD population. Interestingly, this polymorphism was found to be associated with DN in Swedish female T1D patients, but not in the patients from the GoKinD population. Pettersson-Fernholm et al. reported that the allele C frequency was 7.5% in Finnish T1D patients, and the allele C had the risk susceptibility for DN and coronary heart disease (CHD) in Finnish female but not in male patients with T1D (Pettersson-Fernholm et al., 2004). The allele C frequency in Swedish T1D patients is 6.3%, which is slightly lower than in the Finnish T1D patients (Pettersson-Fernholm et al., 2004) but significantly higher than in the American T1D patients of the GoKinD study (4.0%). The results are consistent with other observations that frequency of the allele C was highest in Finnish, second high in Swedish (Karvonen et al., 1998; Ding, 2003; Ding et al., 2005; Nordman et al., 2005) but low in the GoKinD and other populations (Makino et al., 2001; Ding, 2003).

5 CONCLUSIONS

- SNP rs5498(E469K) in the *ICAM-1* gene, with a high heterozygous index, is found to be associated with DN in female T1D patients among the GoKinD population. Although frequencies of the allele A in this polymorphism is found to be increased gradually from non-diabetic controls, to T1D patients without DN and T1D patients with DN in Swedish population, no statistically significant association with DN is detected.
- We have identified four SP1 binding sites in the promoter region of the *AdipoQ* gene. The promoter SNP -11377C/G is found to be associated with DN among female T1D patients of the GoKinD study. This polymorphism may have a role in the regulation of *AdipoQ* promoter transcription activity. Additionally, SNPs +45G15G(T/G) and +276(G/T) in the *AdipoQ* gene are found to be associated with T1D in Swedish Caucasians.
- Frequency of the allele C in *NPY* Leu7Pro polymorphism is higher in Swedish T1D patients than in American GoKinD subjects. This polymorphism is found to be associated with DN in Swedish female T1D patients.

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7 REFERENCES

- Adeghate E, Schattner P, Dunn E. 2006. An update on the etiology and epidemiology of diabetes mellitus. *Ann N Y Acad Sci JT - Annals of the New York Academy of Sciences*. 1084:1-29.
- Arita Y, Kihara S, Ouchi N. et al. 1999. Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem Biophys Res Commun JT - Biochemical and biophysical research communications*. 257(1):79-83.
- Baekkeskov S, Aanstoot HJ, Christgau S. et al. 1990. Identification of the 64K autoantigen in insulin-dependent diabetes as the GABA-synthesizing enzyme glutamic acid decarboxylase. *Nature JT - Nature*. 347(6289):151-6.
- Balasa B, La Cava A, Van Gunst K. et al. 2000. A mechanism for IL-10-mediated diabetes in the nonobese diabetic (NOD) mouse: ICAM-1 deficiency blocks accelerated diabetes. *J Immunol JT - Journal of immunology (Baltimore, Md: 1950)*. 165(12):7330-7.
- Barth N, Langmann T, Scholmerich J. et al. 2002. Identification of regulatory elements in the human adipose most abundant gene transcript-1 (apM-1) promoter: role of SP1/SP3 and TNF-alpha as regulatory pathways. *Diabetologia JT - Diabetologia*. 45(10):1425-33.
- Bate KL, Jerums G. 2003. 3: Preventing complications of diabetes. *Med J Aust JT - The Medical journal of Australia*. 179(9):498-503.
- Bennet WM, Wang ZL, Jones PM. et al. 1996. Presence of neuropeptide Y and its messenger ribonucleic acid in human islets: evidence for a possible paracrine role. *J Clin Endocrinol Metab JT - The Journal of clinical endocrinology and metabolism*. 81(6):2117-20.
- Billington CJ, Briggs JE, Grace M. et al. 1991. Effects of intracerebroventricular injection of neuropeptide Y on energy metabolism. *Am J Physiol JT - The American journal of physiology*. 260(2 Pt 2):R321-7.
- Billington CJ, Briggs JE, Harker S. et al. 1994. Neuropeptide Y in hypothalamic paraventricular nucleus: a center coordinating energy metabolism. *Am J Physiol JT - The American journal of physiology*. 266(6 Pt 2):R1765-70.
- Bischoff A, Michel MC. 1998. Renal effects of neuropeptide Y. *Pflugers Arch JT - Pflugers Archiv : European journal of physiology*. 435(4):443-53.
- Bouatia-Naji N, Meyre D, Lobbens S. et al. 2006. ACDC/adiponectin polymorphisms are associated with severe childhood and adult obesity. *Diabetes JT - Diabetes*. 55(2):545-50.
- Bowden DW, Colicigno CJ, Langefeld CD. et al. 2004. A genome scan for diabetic nephropathy in African Americans. *Kidney Int JT - Kidney international*. 66(4):1517-26.

- Brownlee M. 2001. Biochemistry and molecular cell biology of diabetic complications. *Nature JT - Nature*. 414(6865):813-20.
- Chistiakov DA, Savost'anov KV, Shestakova MV. et al. 2004. Confirmation of a susceptibility locus for diabetic nephropathy on chromosome 3q23-q24 by association study in Russian type 1 diabetic patients. *Diabetes Res Clin Pract JT - Diabetes research and clinical practice*. 66(1):79-86.
- Chow KM, Wong TY, Li PK. 2005. Genetics of common progressive renal disease. *Kidney Int Suppl JT - Kidney international. Supplement*. (94):S41-5.
- Clausen P, Jacobsen P, Rossing K. et al. 2000. Plasma concentrations of VCAM-1 and ICAM-1 are elevated in patients with Type 1 diabetes mellitus with microalbuminuria and overt nephropathy. *Diabet Med JT - Diabetic medicine : a journal of the British Diabetic Association*. 17(9):644-9.
- Conway BR, Savage DA, Maxwell AP. 2006. Identifying genes for diabetic nephropathy--current difficulties and future directions. *Nephrol Dial Transplant JT - Nephrology, dialysis, transplantation : official publication of the European Dialysis and Transplant Association - European Renal Association*. 21(11):3012-7.
- Di Lorenzo TP, Peakman M, Roep BO. 2007. Translational mini-review series on type 1 diabetes: Systematic analysis of T cell epitopes in autoimmune diabetes. *Clin Exp Immunol JT - Clinical and experimental immunology*. 148(1):1-16.
- Diez JJ, Iglesias P. 2003. The role of the novel adipocyte-derived hormone adiponectin in human disease. *Eur J Endocrinol JT - European journal of endocrinology / European Federation of Endocrine Societies*. 148(3):293-300.
- Ding B. 2003. Distribution of the NPY 1128C allele frequency in different populations. *J Neural Transm JT - Journal of neural transmission (Vienna, Austria : 1996)*. 110(11):1199-204.
- Ding B, Kull B, Liu Z. et al. 2005. Human neuropeptide Y signal peptide gain-of-function polymorphism is associated with increased body mass index: possible mode of function. *Regul Pept JT - Regulatory peptides*. 127(1-3):45-53.
- Echtenkamp SF, Dandridge PF. 1989. Influence of renal sympathetic nerve stimulation on renal function in the primate. *Am J Physiol JT - The American journal of physiology*. 257(2 Pt 2):F204-9.
- Ewens KG, George RA, Sharma K. et al. 2005. Assessment of 115 candidate genes for diabetic nephropathy by transmission/disequilibrium test. *Diabetes JT - Diabetes*. 54(11):3305-18.
- Francke S, Manraj M, Lacquemant C. et al. 2001. A genome-wide scan for coronary heart disease suggests in Indo-Mauritians a susceptibility locus on chromosome 16p13 and replicates linkage with the metabolic syndrome on 3q27. *Hum Mol Genet JT - Human molecular genetics*. 10(24):2751-65.
- Fredman D, White SJ, Potter S. et al. 2004. Complex SNP-related sequence variation in segmental genome duplications. *Nat Genet JT - Nature genetics*. 36(8):861-6.

- Frystyk J, Tarnow L, Hansen TK. et al. 2005. Increased serum adiponectin levels in type 1 diabetic patients with microvascular complications. *Diabetologia JT - Diabetologia*. 48(9):1911-8.
- Fujita H, Morii T, Koshimura J. et al. 2006. Possible relationship between adiponectin and renal tubular injury in diabetic nephropathy. *Endocr J JT - Endocrine journal*. 53(6):745-52.
- Gozali M, Pavia JM, Morris MJ. 2002. Involvement of neuropeptide Y in glucose sensing in the dorsal hypothalamus of streptozotocin diabetic rats - in vitro and in vivo studies of transmitter release. *Diabetologia JT - Diabetologia*. 45(9):1332-9.
- Gu HF, Abulaiti A, Ostenson CG. et al. 2004. Single nucleotide polymorphisms in the proximal promoter region of the adiponectin (APM1) gene are associated with type 2 diabetes in Swedish caucasians. *Diabetes JT - Diabetes*. 53 Suppl 1:S31-5.
- Guler S, Cakir B, Demirbas B. et al. 2002. Plasma soluble intercellular adhesion molecule 1 levels are increased in type 2 diabetic patients with nephropathy. *Horm Res JT - Hormone research*. 58(2):67-70.
- Hadjadj S, Aubert R, Fumeron F. et al. 2005. Increased plasma adiponectin concentrations are associated with microangiopathy in type 1 diabetic subjects. *Diabetologia JT - Diabetologia*. 48(6):1088-92.
- Hadjadj S, Belloum R, Bouhanick B. et al. 2001. Prognostic value of angiotensin-I converting enzyme I/D polymorphism for nephropathy in type 1 diabetes mellitus: a prospective study. *J Am Soc Nephrol JT - Journal of the American Society of Nephrology : JASN*. 12(3):541-9.
- Hakonarson H, Grant SF, Bradfield JP. et al. 2007. A genome-wide association study identifies KIAA0350 as a type 1 diabetes gene. *Nature JT - Nature*. 448(7153):591-4.
- Hanson RL, Craig DW, Millis MP. et al. 2007. Identification of PVT1 as a candidate gene for end-stage renal disease in type 2 diabetes using a pooling-based genome-wide single nucleotide polymorphism association study. *Diabetes JT - Diabetes*. 56(4):975-83.
- Hara K, Boutin P, Mori Y. et al. 2002. Genetic variation in the gene encoding adiponectin is associated with an increased risk of type 2 diabetes in the Japanese population. *Diabetes JT - Diabetes*. 51(2):536-40.
- He Z, King GL. 2004. Microvascular complications of diabetes. *Endocrinol Metab Clin North Am JT - Endocrinology and metabolism clinics of North America*. 33(1):215-38, xi-xii.
- Hirata K, Shikata K, Matsuda M. et al. 1998. Increased expression of selectins in kidneys of patients with diabetic nephropathy. *Diabetologia JT - Diabetologia*. 41(2):185-92.

- Howell WM, Jobs M, Gyllenstein U. et al. 1999. Dynamic allele-specific hybridization. A new method for scoring single nucleotide polymorphisms. *Nat Biotechnol* JT - *Nature biotechnology*. 17(1):87-8.
- Imagawa A, Funahashi T, Nakamura T. et al. 2002. Elevated serum concentration of adipose-derived factor, adiponectin, in patients with type 1 diabetes. *Diabetes Care* JT - *Diabetes care*. 25(9):1665-6.
- Imperatore G, Hanson RL, Pettitt DJ. et al. 1998. Sib-pair linkage analysis for susceptibility genes for microvascular complications among Pima Indians with type 2 diabetes. Pima Diabetes Genes Group. *Diabetes* JT - *Diabetes*. 47(5):821-30.
- Iwao M, Morisaki H, Morisaki T. 2004. Single-nucleotide polymorphism g.1548G > A (E469K) in human ICAM-1 gene affects mRNA splicing pattern and TPA-induced apoptosis. *Biochem Biophys Res Commun* JT - *Biochemical and biophysical research communications*. 317(3):729-35.
- Iyengar SK, Schelling JR, Sedor JR. 2002. Approaches to understanding susceptibility to nephropathy: From genetics to genomics. *Kidney Int* JT - *Kidney international*. 61(1 Suppl):61-7.
- Jamal H, Jones PM, Byrne J. et al. 1991. Peptide contents of neuropeptide Y, vasoactive intestinal polypeptide, and beta-calcitonin gene-related peptide and their messenger ribonucleic acids after dexamethasone treatment in the isolated rat islets of Langerhans. *Endocrinology* JT - *Endocrinology*. 129(6):3372-80.
- Joling P, Boom S, Johnson J. et al. 1994. Domain 5 of the intercellular adhesion molecule-1 (ICAM-1) is involved in adhesion of B-cells and follicular dendritic cells. *Adv Exp Med Biol* JT - *Advances in experimental medicine and biology*. 355:131-5.
- Jones CA, Krolewski AS, Rogus J. et al. 2005. Epidemic of end-stage renal disease in people with diabetes in the United States population: do we know the cause?. *Kidney Int* JT - *Kidney international*. 67(5):1684-91.
- Kado S, Wakatsuki T, Yamamoto M. et al. 2001. Expression of intercellular adhesion molecule-1 induced by high glucose concentrations in human aortic endothelial cells. *Life Sci* JT - *Life sciences*. 68(7):727-37.
- Kallio J, Pesonen U, Karvonen MK. et al. 2001. Enhanced exercise-induced GH secretion in subjects with Pro7 substitution in the prepro-NPY. *J Clin Endocrinol Metab* JT - *The Journal of clinical endocrinology and metabolism*. 86(11):5348-52.
- Karvonen MK, Pesonen U, Koulu M. et al. 1998. Association of a leucine(7)-to-proline(7) polymorphism in the signal peptide of neuropeptide Y with high serum cholesterol and LDL cholesterol levels. *Nat Med* JT - *Nature medicine*. 4(12):1434-7.
- Karvonen MK, Ruottinen S, Koulu M. et al. 2006. Nutrient intake, weight, and Leu7Pro polymorphism in prepro-neuropeptide Y in children. *J Clin Endocrinol*

- Metab JT - The Journal of clinical endocrinology and metabolism. 91(11):4664-8.
- Kissebah AH, Sonnenberg GE, Myklebust J. et al. 2000. Quantitative trait loci on chromosomes 3 and 17 influence phenotypes of the metabolic syndrome. Proc Natl Acad Sci U S A JT - Proceedings of the National Academy of Sciences of the United States of America. 97(26):14478-83.
- Koulu M, Movafagh S, Tuohimaa J. et al. 2004. Neuropeptide Y and Y2-receptor are involved in development of diabetic retinopathy and retinal neovascularization. Ann Med JT - Annals of medicine. 36(3):232-40.
- Krikun G, Schatz F, Mackman N. et al. 2000. Regulation of tissue factor gene expression in human endometrium by transcription factors Sp1 and Sp3. Mol Endocrinol JT - Molecular endocrinology (Baltimore, Md.). 14(3):393-400.
- Krolewski M, Eggers PW, Warram JH. 1996. Magnitude of end-stage renal disease in IDDM: a 35 year follow-up study. Kidney Int JT - Kidney international. 50(6):2041-6.
- Lampeter ER, Kishimoto TK, Rothlein R. et al. 1992. Elevated levels of circulating adhesion molecules in IDDM patients and in subjects at risk for IDDM. Diabetes JT - Diabetes. 41(12):1668-71.
- Levine AS, Jewett DC, Cleary JP. et al. 2004. Our journey with neuropeptide Y: effects on ingestive behaviors and energy expenditure. Peptides JT - Peptides. 25(3):505-10.
- Li L, He S, Sun JM. et al. 2004. Gene regulation by Sp1 and Sp3. Biochem Cell Biol JT - Biochemistry and cell biology = Biochimie et biologie cellulaire. 82(4):460-71.
- Lin J, Hu FB, Rimm EB. et al. 2006. The association of serum lipids and inflammatory biomarkers with renal function in men with type II diabetes mellitus. Kidney Int JT - Kidney international. 69(2):336-42.
- Lindstrom T, Frystyk J, Hedman CA. et al. 2006. Elevated circulating adiponectin in type 1 diabetes is associated with long diabetes duration. Clin Endocrinol (Oxf) JT - Clinical endocrinology. 65(6):776-82.
- Liu Y, Freedman BI. 2005. Genetics of progressive renal failure in diabetic kidney disease. Kidney Int Suppl JT - Kidney international. Supplement. (99):S94-7.
- Maeda S, Osawa N, Hayashi T. et al. 2007. Genetic variations associated with diabetic nephropathy and type II diabetes in a Japanese population. Kidney Int Suppl JT - Kidney international. Supplement. (106):S43-8.
- Makino K, Kataoka Y, Hirakawa Y. et al. 2001. A leucine(7)-to-proline(7) polymorphism in the signal peptide of neuropeptide Y was not identified in the Japanese population. J Clin Pharm Ther JT - Journal of clinical pharmacy and therapeutics. 26(3):201-3.

- Marshall SM. 2003. Clinical features and management of diabetic nephropathy. In: Pickup JC, Williams G, editors. Textbook of diabetes. Boston: Blackwell Publishing.
- Matsui H, Suzuki M, Tsukuda R. et al. 1996. Expression of ICAM-1 on glomeruli is associated with progression of diabetic nephropathy in a genetically obese diabetic rat, Wistar fatty. *Diabetes Res Clin Pract JT - Diabetes research and clinical practice.* 32(1-2):1-9.
- Mattevi VS, Zembrzusi VM, Hutz MH. 2002. Association analysis of genes involved in the leptin-signaling pathway with obesity in Brazil. *Int J Obes Relat Metab Disord JT - International journal of obesity and related metabolic disorders : journal of the International Association for the Study of Obesity.* 26(9):1179-85.
- McKnight AJ, Maxwell AP, Sawcer S. et al. 2006. A genome-wide DNA microsatellite association screen to identify chromosomal regions harboring candidate genes in diabetic nephropathy. *J Am Soc Nephrol JT - Journal of the American Society of Nephrology : JASN.* 17(3):831-6.
- Mein CA, Esposito L, Dunn MG. et al. 1998. A search for type 1 diabetes susceptibility genes in families from the United Kingdom. *Nat Genet JT - Nature genetics.* 19(3):297-300.
- Michel MC, Gaida W, Beck-Sickinger AG. et al. 1992. Further characterization of neuropeptide Y receptor subtypes using centrally truncated analogs of neuropeptide Y: evidence for subtype-differentiating effects on affinity and intrinsic efficacy. *Mol Pharmacol JT - Molecular pharmacology.* 42(4):642-8.
- Miller JA, Thai K, Scholey JW. 2000. Angiotensin II type 1 receptor gene polymorphism and the response to hyperglycemia in early type 1 diabetes. *Diabetes JT - Diabetes.* 49(9):1585-9.
- Moczulski DK, Rogus JJ, Antonellis A. et al. 1998. Major susceptibility locus for nephropathy in type 1 diabetes on chromosome 3q: results of novel discordant sib-pair analysis. *Diabetes JT - Diabetes.* 47(7):1164-9.
- Moczulski DK, Scott L, Antonellis A. et al. 2000. Aldose reductase gene polymorphisms and susceptibility to diabetic nephropathy in Type 1 diabetes mellitus. *Diabet Med JT - Diabetic medicine : a journal of the British Diabetic Association.* 17(2):111-8.
- Mohamed-Ali V, Armstrong L, Clarke D. et al. 2001. Evidence for the regulation of levels of plasma adhesion molecules by proinflammatory cytokines and their soluble receptors in type 1 diabetes. *J Intern Med JT - Journal of internal medicine.* 250(5):415-21.
- Moltz JH, McDonald JK. 1985. Neuropeptide Y: direct and indirect action on insulin secretion in the rat. *Peptides JT - Peptides.* 6(6):1155-9.

- Morris MJ, Pavia JM. 2004. Increased endogenous noradrenaline and neuropeptide Y release from the hypothalamus of streptozotocin diabetic rats. *Brain Res JT - Brain research*. 1006(1):100-6.
- Mueller PW, Rogus JJ, Cleary PA. et al. 2006. Genetics of Kidneys in Diabetes (GoKinD) study: a genetics collection available for identifying genetic susceptibility factors for diabetic nephropathy in type 1 diabetes. *J Am Soc Nephrol JT - Journal of the American Society of Nephrology : JASN*. 17(7):1782-90.
- Myrsen-Axcrona U, Karlsson S, Sundler F. et al. 1997. Dexamethasone induces neuropeptide Y (NPY) expression and impairs insulin release in the insulin-producing cell line RINm5F. Release of NPY and insulin through different pathways. *J Biol Chem JT - The Journal of biological chemistry*. 272(16):10790-6.
- Neamat-Allah M, Feeney SA, Savage DA. et al. 2001. Analysis of the association between diabetic nephropathy and polymorphisms in the aldose reductase gene in Type 1 and Type 2 diabetes mellitus. *Diabet Med JT - Diabetic medicine : a journal of the British Diabetic Association*. 18(11):906-14.
- Nelson CL, Karschimkus CS, Dragicevic G. et al. 2005. Systemic and vascular inflammation is elevated in early IgA and type 1 diabetic nephropathies and relates to vascular disease risk factors and renal function. *Nephrol Dial Transplant JT - Nephrology, dialysis, transplantation : official publication of the European Dialysis and Transplant Association - European Renal Association*. 20(11):2420-6.
- Nerup J, Platz P, Andersen OO. et al. 1974. HL-A antigens and diabetes mellitus. *Lancet JT - Lancet*. 2(7885):864-6.
- Ng DP, Krolewski AS. 2005. Molecular genetic approaches for studying the etiology of diabetic nephropathy. *Curr Mol Med JT - Current molecular medicine*. 5(5):509-25.
- Nordman S, Ding B, Ostenson CG. et al. 2005. Leu7Pro polymorphism in the neuropeptide Y (NPY) gene is associated with impaired glucose tolerance and type 2 diabetes in Swedish men. *Exp Clin Endocrinol Diabetes JT - Experimental and clinical endocrinology & diabetes : official journal, German Society of Endocrinology [and] German Diabetes Association*. 113(5):282-7.
- Okada S, Shikata K, Matsuda M. et al. 2003. Intercellular adhesion molecule-1-deficient mice are resistant against renal injury after induction of diabetes. *Diabetes JT - Diabetes*. 52(10):2586-93.
- Omi H, Okayama N, Shimizu M. et al. 2002. Participation of high glucose concentrations in neutrophil adhesion and surface expression of adhesion molecules on cultured human endothelial cells: effect of antidiabetic medicines. *J Diabetes Complications JT - Journal of diabetes and its complications*. 16(3):201-8.

- Osterholm AM, He B, Pitkaniemi J. et al. 2007. Genome-wide scan for type 1 diabetic nephropathy in the Finnish population reveals suggestive linkage to a single locus on chromosome 3q. *Kidney Int JT - Kidney international.* 71(2):140-5.
- Palmer JP, Asplin CM, Clemons P. et al. 1983. Insulin antibodies in insulin-dependent diabetics before insulin treatment. *Science JT - Science (New York, N.Y.).* 222(4630):1337-9.
- Parving HH. 2001. Diabetic nephropathy: prevention and treatment. *Kidney Int JT - Kidney international.* 60(5):2041-55.
- Payton MA, Hawkes CJ, Christie MR. 1995. Relationship of the 37,000- and 40,000-M(r) tryptic fragments of islet antigens in insulin-dependent diabetes to the protein tyrosine phosphatase-like molecule IA-2 (ICA512). *J Clin Invest JT - The Journal of clinical investigation.* 96(3):1506-11.
- Pettersson-Fernholm K, Karvonen MK, Kallio J. et al. 2004. Leucine 7 to proline 7 polymorphism in the preproneuropeptide Y is associated with proteinuria, coronary heart disease, and glycemic control in type 1 diabetic patients. *Diabetes Care JT - Diabetes care.* 27(2):503-9.
- Pihlajamaki J, Karhapaa P, Vauhkonen I. et al. 2003. The Leu7Pro polymorphism of the neuropeptide Y gene regulates free fatty acid metabolism. *Metabolism JT - Metabolism: clinical and experimental.* 52(5):643-6.
- Playford RJ, Mehta S, Upton P. et al. 1995. Effect of peptide YY on human renal function. *Am J Physiol JT - The American journal of physiology.* 268(4 Pt 2):F754-9.
- Qi XM, Wu GZ, Wu YG. et al. 2006. Renoprotective effect of breviscapine through suppression of renal macrophage recruitment in streptozotocin-induced diabetic rats. *Nephron Exp Nephrol JT - Nephron. Experimental nephrology.* 104(4):e147-57.
- Quinn M, Angelico MC, Warram JH. et al. 1996. Familial factors determine the development of diabetic nephropathy in patients with IDDM. *Diabetologia JT - Diabetologia.* 39(8):940-5.
- Ray D, Mishra M, Ralph S. et al. 2004. Association of the VEGF gene with proliferative diabetic retinopathy but not proteinuria in diabetes. *Diabetes JT - Diabetes.* 53(3):861-4.
- Ritz E. 2005. Managing anaemia and diabetes: a future challenge for nephrologists. *Nephrol Dial Transplant JT - Nephrology, dialysis, transplantation : official publication of the European Dialysis and Transplant Association - European Renal Association.* 20 Suppl 6:vi21-5.
- Ronaghi M, Uhlen M, Nyren P. 1998. A sequencing method based on real-time pyrophosphate. *Science JT - Science (New York, N.Y.).* 281(5375):363, 365.
- Rudberg S, Rasmussen LM, Bangstad HJ. et al. 2000. Influence of insertion/deletion polymorphism in the ACE-I gene on the progression of diabetic glomerulopathy

- in type 1 diabetic patients with microalbuminuria. *Diabetes Care* JT - *Diabetes care*. 23(4):544-8.
- Saraheimo M, Forsblom C, Fagerudd J. et al. 2005. Serum adiponectin is increased in type 1 diabetic patients with nephropathy. *Diabetes Care* JT - *Diabetes care*. 28(6):1410-4.
- Schutta MH. 2007. Diabetes and hypertension: epidemiology of the relationship and pathophysiology of factors associated with these comorbid conditions. *J Cardiometab Syndr* JT - *Journal of the cardiometabolic syndrome*. 2(2):124-30.
- Schwarz PE, Govindarajulu S, Towers W. et al. 2006. Haplotypes in the promoter region of the ADIPOQ gene are associated with increased diabetes risk in a German Caucasian population. *Horm Metab Res* JT - *Hormone and metabolic research. Hormon- und Stoffwechselforschung. Hormones et métabolisme*. 38(7):447-51.
- Seaquist ER, Goetz FC, Rich S. et al. 1989. Familial clustering of diabetic kidney disease. Evidence for genetic susceptibility to diabetic nephropathy. *N Engl J Med* JT - *The New England journal of medicine*. 320(18):1161-5.
- Shaw CJ, Lupski JR. 2004. Implications of human genome architecture for rearrangement-based disorders: the genomic basis of disease. *Hum Mol Genet* JT - *Human molecular genetics*. 13 Spec No 1:R57-64.
- Shimazaki A, Kawamura Y, Kanazawa A. et al. 2005. Genetic variations in the gene encoding ELMO1 are associated with susceptibility to diabetic nephropathy. *Diabetes* JT - *Diabetes*. 54(4):1171-8.
- Spielman RS, McGinnis RE, Ewens WJ. 1993. Transmission test for linkage disequilibrium: the insulin gene region and insulin-dependent diabetes mellitus (IDDM). *Am J Hum Genet* JT - *American journal of human genetics*. 52(3):506-16.
- Statnick MA, Beavers LS, Conner LJ. et al. 2000. Decreased expression of apM1 in omental and subcutaneous adipose tissue of humans with type 2 diabetes. *Int J Exp Diabetes Res* JT - *International journal of experimental diabetes research*. 1(2):81-8.
- Staunton DE, Marlin SD, Stratowa C. et al. 1988. Primary structure of ICAM-1 demonstrates interaction between members of the immunoglobulin and integrin supergene families. *Cell* JT - *Cell*. 52(6):925-33.
- Sugimoto H, Shikata K, Hirata K. et al. 1997. Increased expression of intercellular adhesion molecule-1 (ICAM-1) in diabetic rat glomeruli: glomerular hyperfiltration is a potential mechanism of ICAM-1 upregulation. *Diabetes* JT - *Diabetes*. 46(12):2075-81.
- The Wellcome Trust Case Control Consortium. 2007. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature* JT - *Nature*. 447(7145):661-78.

- Todd JA, Walker NM, Cooper JD. et al. 2007. Robust associations of four new chromosome regions from genome-wide analyses of type 1 diabetes. *Nat Genet JT - Nature genetics*. 39(7):857-64.
- Toivonen A, Kulmala P, Rahko J. et al. 2004. Soluble adhesion molecules in Finnish schoolchildren with signs of preclinical type 1 diabetes. *Diabetes Metab Res Rev JT - Diabetes/metabolism research and reviews*. 20(1):48-54.
- van de Stolpe A, van der Saag PT. 1996. Intercellular adhesion molecule-1. *J Mol Med JT - Journal of molecular medicine (Berlin, Germany)*. 74(1):13-33.
- Vasseur F, Helbecque N, Dina C. et al. 2002. Single-nucleotide polymorphism haplotypes in the both proximal promoter and exon 3 of the APM1 gene modulate adipocyte-secreted adiponectin hormone levels and contribute to the genetic risk for type 2 diabetes in French Caucasians. *Hum Mol Genet JT - Human molecular genetics*. 11(21):2607-14.
- Vasseur F, Helbecque N, Lobbens S. et al. 2005. Hypoadiponectinaemia and high risk of type 2 diabetes are associated with adiponectin-encoding (ACDC) gene promoter variants in morbid obesity: evidence for a role of ACDC in diabetes. *Diabetologia JT - Diabetologia*. 48(5):892-9.
- Venter JC, Adams MD, Myers EW. et al. 2001. The sequence of the human genome. *Science JT - Science (New York, N.Y.)*. 291(5507):1304-51.
- Vionnet N, Hani EH, Dupont S. et al. 2000. Genomewide search for type 2 diabetes-susceptibility genes in French whites: evidence for a novel susceptibility locus for early-onset diabetes on chromosome 3q27-qter and independent replication of a type 2-diabetes locus on chromosome 1q21-q24. *Am J Hum Genet JT - American journal of human genetics*. 67(6):1470-80.
- Vionnet N, Tregouet D, Kazeem G. et al. 2006. Analysis of 14 candidate genes for diabetic nephropathy on chromosome 3q in European populations: strongest evidence for association with a variant in the promoter region of the adiponectin gene. *Diabetes JT - Diabetes*. 55(11):3166-74.
- Wang ZL, Bennet WM, Wang RM. et al. 1994. Evidence of a paracrine role of neuropeptide-Y in the regulation of insulin release from pancreatic islets of normal and dexamethasone-treated rats. *Endocrinology JT - Endocrinology*. 135(1):200-6.
- Wild S, Roglic G, Green A. et al. 2004. Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. *Diabetes Care JT - Diabetes care*. 27(5):1047-53.
- Zacharova J, Chiasson JL, Laakso M. 2005. The common polymorphisms (single nucleotide polymorphism [SNP] +45 and SNP +276) of the adiponectin gene predict the conversion from impaired glucose tolerance to type 2 diabetes: the STOP-NIDDM trial. *Diabetes JT - Diabetes*. 54(3):893-9.