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1 **Sensitivity of Salmonella YG5161 for Detecting PAH-Associated Mutagenicity in Air**

2 **Particulate Matter**

3

4 **Running title: Sensitivity of YG5161 for detection of PAH-mutagenicity in air**

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23

1 **ABSTRACT**

2 The Salmonella/microsome assay is the most used assay for the evaluation of air particulate
3 matter (PM) mutagenicity and a positive correlation between strain TA98 responses and
4 benzo[a]pyrene (B[a]P) levels in PM has been found. However, it seems that the major causes
5 of PM mutagenicity in this assay are the nitro and oxy-PAHs. Salmonella YG5161, a 30-
6 times more responsive strain to B[a]P has been developed. To verify if YG5161 strain was
7 sufficiently sensitive to detect mutagenicity associated with B[a]P mutagenicity, PM samples
8 were collected in Brazil and Sweden, extracted with toluene and tested in the
9 Salmonella/microsome microsuspension assay. PAHs and B[a]P were determined and the
10 extracts were tested with YG5161 and its parental strain TA1538. The extracts were also
11 tested with YG1041 and its parental strain TA98. For sensitivity comparisons, we tested
12 B[a]P and 1-nitropyrene (1-NP) using the same conditions. The minimal effective dose of
13 B[a]P was 155 ng/plate for TA1538 and 7 ng/plate for YG5161. Although the maximum
14 tested dose, 10 m³/plate containing 9 ng of B[a]P in the case of Brazilian sample, was
15 sufficient to elicit a response in YG5161, mutagenicity was detected at a dose as low as 1
16 m³/plate (0.9 ng). This is probably caused by nitro-compounds that have been shown to be
17 even more potent than B[a]P for YG5161. It seems that the mutagenicity of B[a]P present in
18 PM is not detectable even with the use of YG5161 unless more efficient separation to remove
19 the nitro-compounds from the PAH extract is performed.

1 INTRODUCTION

2 The mutagenicity of airborne particulate matter (PM) can be attributed to at least 500
3 identified compounds from different chemical classes [Claxton et al., 2004]. Among these,
4 benzo[a]pyrene (B[a]P), along with other polycyclic aromatic hydrocarbons (PAHs), have
5 received great attention because of their recognized carcinogenic and mutagenic potential
6 [Srogi, 2007].

7 The Salmonella/microsome assay is the most widely used method for the evaluation of
8 the mutagenic activity of pure compounds and environmental samples [Claxton et al., 2010],
9 including atmospheric samples [Claxton and Woodall Jr, 2007]. The assay is sensitive to
10 several PAHs, including B[a]P [DeMarini et al., 2011; Brito et al., 2013]. Some studies have
11 correlated the mutagenic activity detected in the Salmonella/microsome assay with the levels
12 of B[a]P and other non-substituted PAHs present in the samples [Viras et al., 1990; Nielsen et
13 al., 1996; Claxton and Woodall Jr., 2007; Srogi, 2007], although it does not seem to
14 demonstrate a direct relationship. The primary components responsible for the mutagenicity
15 of air particulate matter in the Salmonella assay seem to be nitro and oxy-PAHs [Claxton et
16 al., 2004; Sharma et al., 2007; Umbuzeiro et al., 2008 a, b; Walgraeve et al., 2010]. One
17 explanation for this could be that the typically used strains (TA98 and TA100) are more
18 sensitive to nitro- and oxy- PAHs than to non-substituted PAHs [Claxton et al., 2004; Enya et
19 al., 1997; Enya et al., 1998; Kummrow et al., 2006; Franco et al., 2010]. To enhance the
20 sensitivity of the Salmonella/microsome assay to non-substituted PAHs Matsui et al. [2006]
21 developed the YG5161 strain which is more responsive to B[a]P and other non-substituted
22 PAHs than its parental strain TA1538. The YG5161 strain overexpresses DNA polymerase
23 IV, and has the *dinB* gene of *Escherichia coli* encoded in the pYG768 plasmid, which also
24 confers ampicillin resistance to the strain. DNA polymerase IV facilitates the error-prone
25 bypass of the DNA guanine adducts formed by polycyclic aromatic compounds which, after

1 repair, will lead to the deletion of two base pairs and consequently shifting of the reading
2 frame [Matsui et al., 2006]. These authors suggested the possibility of using YG5161 as a
3 major strain for the detection of the mutagenicity of non-substituted PAHs such as B[a]P.
4 Because this compound needs to be metabolized to react with DNA [Uppstad et al., 2010], the
5 addition of S9 mix is required for its detection in the Salmonella/mutagenicity assay.

6 Some strains have also been developed to be more sensitive to different compounds.
7 For example YG1041 strain, a derivative of TA98, is more sensitive to nitroarenes and
8 aromatic amines because it overproduces nitroreductase and *O*-acetyltransferases, both
9 important enzymes in the activation of such compounds [Hagiwara et al., 1993]. Similarly,
10 the strain YG7108 which is derived from TA1535 is more responsive to alkylating agents
11 [Yamada et al., 1997]. Both strains have been used in the identification of the types of
12 compounds predominantly responsible for the mutagenic activity of a test samples
13 [Umbuzeiro et al., 2011]. Mutlu et al. [2013] demonstrated the applicability of strains with
14 different sensitivities for analyzing environmental samples. In a hierarchical clustering
15 analysis they showed that although PAHs, aromatic amines, and nitro-compounds were
16 present in diesel exhaust extracts, oxy-PAHs were the cause of much of the mutagenicity.

17 When atmospheric particulate air samples are tested they usually demonstrate a clear
18 increase with YG1041 in relation to TA98, indicating that the mutagenicity is related to nitro-
19 and oxy- PAHs and not B[a]P, although PAHs are present in those samples when chemical
20 analyses are performed [DeMarini et al., 2004; Umbuzeiro et al., 2008a,b].

21 The objective of this work was to verify if the YG5161 strain in the
22 Salmonella/microsome microsuspension assay was sufficiently sensitive to B[a]P
23 mutagenicity in air particulate matter collected in two different locations, Limeira, Brazil and
24 Stockholm, Sweden. We determined the total PAH and B[a]P levels and tested the same
25 extracts with YG5161 and its parental strain, TA1538. We also tested the same extracts with

1 YG1041 and its parental strain, TA98. For sensitivity comparisons we tested several
2 concentrations of B[a]P and 1-nitropyrene (1-NP) using the same conditions.

3 4 **MATERIAL AND METHODS**

5 **Sampling sites**

6 Total atmospheric particulate matter (PM) was collected at two sites: the campus of
7 the Faculty Technology at UNICAMP, Limeira, Brazil (LIMEIRA) and the campus of the
8 Stockholm University, Stockholm, Sweden (STHOLM). The LIMEIRA site is impacted by
9 heavy traffic including cars and trucks, industrial emissions, and sugar cane growing and
10 harvesting activities, including biomass burning. The mutagenic potencies of previously
11 evaluated extracts from this site using the Salmonella/microsome assay [Alves, 2011] and
12 surrounding areas [Umbuzeiro et al., 2008a, b] were among the highest potencies found in the
13 literature. The STHOLM site is also impacted by heavy traffic with emissions from cars and
14 trucks as the main source of pollution. PM samples from this site have recently been used to
15 investigate the toxicological impact of PAHs in air PM [Jarvis et al., 2013a,b].

16 Total PM samples from Limeira were collected at street level on a glass-fiber filter
17 (254 × 233 mm; 0.33 mm pore size. Energética Ind. Com. LTDA, Rio de Janeiro, RJ, Brazil)
18 using a high-volume sampler (Energética Ind. Com. LTDA, Rio de Janeiro, RJ, Brazil)
19 operated at an average flow rate of 1,130 L/min for 24 h. The air total PM samples from
20 Stockholm, Sweden were collected at roof-top level (22 m from the ground) on a
21 fluorocarbon-coated glass fiber filter (235 mm of diameter; Fiberfilm Filters, Pallflex, Pall
22 Corporation, Putnam, CT, USA) with an average flow rate of 1,209 L/min for 71 h. The total
23 PM concentrations for Limeira were 95.8 ug/m³ and for Stockholm, 7.16 ug/m³.

24

25 **Organic extraction and PAH analysis**

1 Extraction was performed by pressurized fluid extraction using an ASE 200
2 accelerated solvent extraction system (Dionex Co., Sunnyvale, CA, USA) using toluene for 5
3 x 30 min. This extraction procedure was previously developed and validated for analysis of
4 PAHs in air PM [Bergvall and Westerholm, 2008]. Aliquots of the extracts were evaporated
5 gently under a nitrogen stream at 55°C until dryness and re-dissolved in dimethylsulfoxide
6 (DMSO) for the biological assays. Portions of the extracts were analyzed for 42 PAHs (Table
7 I) using silica solid phase extraction (SPE) cartridges and on-line liquid chromatography-gas
8 chromatography/mass spectrometry as described in detail elsewhere [Sadiktsis et al., 2014]. A
9 blank filter was also extracted and analyzed as an analytical control.

10

11 **Salmonella/microsome microsuspension assay**

12 The extracts were assayed in the Salmonella/microsome microsuspension assay [Kado
13 et al., 1983; DeMarini et al., 1989] in dose-response experiments using the *Salmonella* strains
14 TA1538, TA98, YG5161 and YG1041 with and without metabolic activation (S9). Strains
15 were kindly provided by Dr. Takehiko Nohmi, except for TA98, which was kindly provided
16 by Dr. Larry Claxton. Table II summarizes the genetic characteristics of the strains. B[a]P and
17 1-NP were also tested with the four strains in dose response experiments. Overnight cultures
18 (around 10⁹ cells/mL) were concentrated 5-fold by centrifugation (10,000xg at 4 °C for 10
19 min) and resuspended into 0.015 M sodium phosphate buffer. A volume of 50 µL of cell
20 suspension, 50 µL of 0.015 M sodium phosphate buffer or S9 mix, and 5 µL of the sample
21 were incubated at 37 °C for 90 min without shaking. To the mixture, 2 mL of molten agar was
22 added and poured onto a minimal agar plate. Colonies were counted after 66 h of incubation
23 at 37 °C by hand with the aid of a stereomicroscope. Toxicity was also carefully evaluated by
24 observing the background of the agar plates. Metabolic activation was provided by Aroclor

1 1254-induced Sprague Dawley rat liver S9 mix (MolTox, Boone, NC) prepared at 4 % v/v
2 and supplemented with the required co-factors [Mortelmans and Zeiger, 2000].

3 PM extracts were tested at 0.02, 0.2, 1, 2, 5 and 10 m³ per plate. Positive controls
4 without S9 were 4-nitroquinoline-oxide (4NQO) at 0.125 µg/plate for TA98 and YG5161 and
5 4-nitro-o-phenylenediamine (4NOP) at 2.5 µg/plate for YG1041. With S9, 2-aminoanthracene
6 (2AA) at 0.625 µg/plate was used as a positive control for TA1538, TA98 and YG5161 and at
7 0.03125 µg/plate for YG1041. Duplicates of each concentration were tested, except for the
8 negative control that was tested in triplicate. DMSO was used as negative control and to
9 dilute the extracts and positive controls. The extract of a clean (blank) filter was also tested.

10 Data were analyzed with the Salanal computer program using the Bernstein model
11 [Bernstein et al., 1982]. Samples were considered positive when a significant difference
12 among the tested doses and the negative control (ANOVA) and a significant positive dose
13 response were observed. Results were expressed as revertants per m³ and per mg of PM
14 equivalent. Also, the minimum effective dose (MED) to elicit a positive response was
15 calculated for B[a]P and 1-NP. MED was defined as the dose that provided a doubling of
16 revertants based on the calculated linear regression curve.

17

18 **RESULTS AND DISCUSSION**

19

20 In this study, we evaluated the mutagenicity of PM organic extracts from two cities:
21 Limera, Brazil and Stockholm, Sweden. The concentration of PM collected in LIMEIRA
22 was 95.8 µg/m³ and in STHOLM was 7.16 µg/m³. The total PAH content in the LIMEIRA
23 sample was 10.66 ng/m³ with 0.9 ng/m³ of B[a]P, and the STHOLM sample contained 4.43
24 ng/m³ with 0.29 ng/m³ of B[a]P (Table I). The mean of the number of revertants per plate
25 obtained for the four strains including negative, blank, positive controls and calculated
26 potencies are presented in Table III for LIMEIRA and Table IV for STHOLM. In LIMEIRA,

1 the potency for TA98 without S9 was 100 revertants/m³, similar what had been reported for
2 other cities in Brazil [Sato et al., 1995; Ducatti and Vargas, 2003; Umbuzeiro et al., 2008 a,
3 b]. The potency of the STHOLM sample was 4 times lower (25 revertants/m³). The potency
4 values decreased for both locations when S9 was used (Tables III and IV). According to
5 Claxton et al. [2004] this is a first indication of a response of nitroaromatics. In general the
6 samples from LIMEIRA were more mutagenic than the STHOLM in all strains/conditions
7 tested, especially with TA98 with S9 (13-fold difference). However, because only one sample
8 from each site was analyzed it was not possible to compare the mutagenicity of the two sites
9 with a high level of confidence. A comprehensive study, which includes additional sampling
10 at the same season of the year, is being conducted to provide additional information on the
11 potencies at both sites.

12 To establish how sensitive each strain was to B[a]P, the prototypical non-substituted
13 PAH, and 1-NP, a typical nitroaromatic, their mutagenicities were assessed using the same
14 protocol used to evaluate the PM samples.

15 YG5161 was the most sensitive strain for B[a]P showing a MED of 7 ng/plate in
16 contrast to 77.5 ng/plate for TA1538, 265 ng/plate for YG1041, and 1,400 ng/plate for TA98
17 (Table V). Although several non-substituted PAHs were detected in both PM samples (Table
18 II), the potency of the YG5161 response did not provide a typical B[a]P response, which
19 would be an increase of 30-fold in relation to TA1538 (Table V). This is most likely due to
20 the fact that the amount of B[a]P present in the 10 m³ of PM (the maximum dose tested) was
21 9 ng/plate and the MED of B[a]P for YG5161 is 7 ng/plate (Table V). All the comparisons
22 were made with S9 because non-substituted PAHs require S9 to be activated.

23 YG1041, as expected, demonstrated much higher sensitivity to 1-NP than any other
24 strain, with a MED of 0.03 ng/plate. MEDs for TA98, TA1538 and YG5161 were 0.45, 1.8
25 and 1.8 ng/plate, respectively (Table VI). 1-NP was chosen as an example for testing purpose,

1 but other nitrocompounds (e.g. dinitropyrenes, nitrobenzanthrones) are more mutagenic and
2 are likely to be present in PM samples. For both PM samples we observed a typical response
3 of nitroaromatics because the potency of YG1041 was 35 times higher than TA98. This
4 comparison was made without S9 because all the potencies decreased with S9. This behavior
5 is consistent with the hypothesis that nitroaromatics are the major cause of the mutagenic
6 effect.

7 The data suggest that B[a]P is not the compound predominantly causing the mutagenic
8 responses of the PM analyzed in this study, but that nitroaromatics such as 1-NP seems to be
9 causing the observed effect. Chemical analysis of nitroaromatics could be performed in the
10 same extracts in order to determine their contribution to the mutagenicity.

11 Other authors also used YG5161 to test different samples. Sharma et al. [2007] used
12 TA98, YG1041, and YG5161 strains, in the Salmonella/microsome microsuspension assay to
13 evaluate the genotoxicity of organic fractions of PM collected from an incineration energy
14 plant and urban air. The fraction containing moderately polar PAHs, alkyl-PAHs and *O*- and
15 *S*-heterocyclics was mutagenic only in the presence of S9 mix, indicating that compounds
16 such as B[a]P could be responsible for the observed effect. Because the mutagenic activity
17 with the YG5161 strain was lower in comparison to TA98 it is plausible to suspect that non-
18 substituted PAHs are not predominantly contributing to the observed mutagenic activity.
19 Conversely, the fraction containing N-heterocyclics, nitro-, amino- and oxy-PAHs was more
20 mutagenic without S9 and higher in YG1041 than TA98, suggesting that nitroarenes and
21 aromatic amines are likely to be the major cause of the mutagenic activity.

22 Maertens et al. [2009] used the Salmonella/microsome assay with several
23 strains/conditions, including TA98, YG1041 and YG5161 to, evaluate the mutagenicity of
24 tobacco and marijuana smoke condensates. All of the tested smoke condensates presented
25 positive responses. The mutagenic potencies obtained with YG1041 and with YG5161 were

1 higher than the ones with TA98 indicating that aromatic amines and non-substituted PAHs
2 were contributing to the mutagenicity. Although the authors identified B[a]P and
3 benzo[a]anthracene in the samples, they contributed to less than 0.1% of the total
4 mutagenicity indicating that there are other major mutagens in the mixture.

5 Yauk et al. [2012] observed mutagenic activity in cigarette smoke condensate extracts
6 using the Salmonella/microsome assay with the TA98, YG1041, and YG5161 strains in the
7 presence of S9 mix. The authors observed that the potencies with YG1041 were greater than
8 those for TA98 but the increase of YG5161 potencies in relation to TA98 was not significant,
9 indicating that non-substituted PAHs could not fully explain the mutagenicity.

10 Therefore we conclude that even the highly sensitive strain to B[a]P, YG5161, does
11 not seem to be able to distinguish the response of B[a]P in samples containing nitroaromatics.
12 We observe that only 1 m³ is sufficient to elicit a positive response in the
13 Salmonella/microsuspension assay for the majority of the strains/conditions. Both for the
14 LIMEIRA and STHOLM air samples.

15 Therefore we can suggest that although associations were observed between B[a]P
16 content and mutagenicity, this compound does not totally explain the mutagenicity of PM air
17 samples tested in the Salmonella/microsome assay unless amounts of B[a]P are higher than 7
18 ng/plate and nitrocompounds are eliminated by the fractionation procedure.

19

20 **CONFLICT OF INTEREST**

21 The authors declare no conflict of interest.

22

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3

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Contribution of the Authors

Gisela A. Umbuzeiro: designed experiments, performed the Salmonella/microsome assay,
analyzed data and drafted the manuscript

Fábio Kummrow: designed experiments, performed the Salmonella/microsome assay,
analyzed data and drafted the manuscript

Daniel Alexandre Morales: designed experiments, performed the Salmonella/microsome
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Ian W. H. Jarvis: designed experiments, analyzed data and drafted the manuscript

Christoffer Bergvall: designed experiments, analyzed data and drafted the manuscript

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Kristian Dreij: designed experiments, analyzed data and drafted the manuscript

TABLE I. PAHs content (pg/m³) in extracts of the PM samples collected in Limeira, Brazil and Stockholm, Sweden.

PAH	LIMEIRA (pg/m ³)		STHOLM (pg/m ³)	
	Mean	SD	Mean	SD
Phenanthrene	798	27	250	2
Anthracene	180	14	33.6	0.7
3-Methylphenanthrene	47.0	2.5	23.8	0.7
2-Methylphenanthrene	65.4	2.1	32.0	1.3
2-Methylantracene	14.8	0.7	5.75	0.52
9-Methylphenanthrene	34.5	1.5	18.0	1.4
1-Methylphenanthrene	43.3	4.2	37.2	0.8
4H-Cyclopenta[def]phenanthrene	58.9	2.0	47.8	0.1
2-Phenylanthracene	35.0	2.9	26.9	2.6
3,6-Dimethylphenanthrene	5.03	0.20	1.70	0.11
3,9-Dimethylphenanthrene	22.9	1.5	8.31	0.75
Fluoranthene	671	6	368	6
Pyrene	895	25	439	8
1-Methylfluoranthene	40.9	3.5	46.1	3.7
Benzo[a]fluorene	28.7	1.2	27.0	2.4
Benzo[b]fluorene	16.2	1.5	13.0	0.8
2-Methylpyrene	30.8	2.9	18.4	1.4
4-Methylpyrene	52.3	4.7	30.7	2.9
1-Methylpyrene	47.7	3.9	32.1	3.2
Benzo[ghi]fluoranthene	346	15	207	8
Benzo[c]phenanthrene	48.5	3.4	77.6	2.1
Benzo[b]naphto(1,2-d)thiophene	9.95	0.31	2.16	0.71
Benz[a]anthracene	332	3	306	10
3-Methylchrysene	39.1	2.3	21.2	2.0
2-Methylchrysene	61.0	3.3	47.9	3.5
6-Methylchrysene	40.2	2.5	32.6	2.5
1-Methylchrysene	47.1	2.6	51.5	3.3
Benzo[b]fluoranthene	974	19	465	8
Benzo[k]fluoranthene	440	5	180	4
Benzo[e]pyrene	1050	31	352	13
Benzo[a]pyrene	899	17	289	4
Perylene	164	6	45.1	0.2
Indeno[1,2,3-cd]fluoranthene	65.4	1.5	28.2	0.3
Indeno[1,2,3-cd]pyrene	908	28	230	12
Dibenz[a,h]anthracene	55.9	2.7	35.6	3.3
Picene	76.2	2.2	30.9	1.7
Benzo[ghi]perylene	1620	35	351	7
Dibenzo[a,l]pyrene	4.79	0.37	5.14	0.28
Dibenzo[a,e]pyrene	15.7	0.6	32.4	2.0
Coronene	366	8	171	10
Dibenzo[a,i]pyrene	9.23	0.28	3.66	0.17
Dibenzo[a,h]pyrene	3.03	0.11	2.53	0.19
Total	10,663		4,427	

TABLE II. Summary of the main genetic characteristics of *Salmonella typhimurium* specific strains.

Strain	Description	Reversion event	Plasmids	Reference
TA1538	<i>hisD3052, Δ(uvrB, bio), rfa</i>	Frameshift	No plasmids	Maron and Ames, [1983]
TA98	<i>hisD3052, Δ(uvrB, bio), rfa, Ap^r</i>	Frameshift	pKM101	Maron and Ames, [1983]
YG1041	<i>hisD3052, Δ(uvrB, bio), rfa, Ap^r and Km^r, NR and O-AT overproducing strain</i>	Frameshift	pKM101, pYG233	Hagiwara et al. [1993]
YG5161	<i>hisD3052, Δ(uvrB, bio), rfa, Ap^r, DNA Pol IV overproducing</i>	Frameshift	pYG768	Matsui et al. [2006]

his – mutation in the histidine operon
 $\Delta uvrB$ – deletion of *uvrB* gene
 Δbio – deletion of biotin gene
rfa – mutation cause partial loss of the lipopolysaccharide barrier
 Ap^r – resistant to ampicillin
 NR – nitroreductase
 O-AT – O-acetyltransferase
 DNA Pol. IV – DNA Polymerase IV
 Km^r – resistant to Kanamycin

TABLE III. Mutagenicity for LIMEIRA PM extracts in the Salmonella/microsome microsuspension assay with TA98, YG1041, TA1538 and YG5161 without and with S9.

Doses	Mean of Number of Revertants/plate and <i>Standard Deviation (SD)</i>							
m ³ /plate	TA98				YG1041			
	-S9		+S9		-S9		+S9	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Negative control	52.7	4.51	53.3	14.4	157.7	17.1	145.7	9.1
0.02	53.5	2.12	58.0	7.1	262.0	24.1	151.5	17.7
0.2	68.5	6.4	59.5	0.7	849.5	7.8	329.5	16.3
1	144.0*	22.6	69.0	8.5	1,922.5	160.5	1,344.0	73.5
2	207.5	45.9	59.0	2.8	3,583.5	231.2	3,312.5	50.2
5	595.5	37.5	76.0	9.9	4,145.5	381.1	5,308.0	151.3
10	1,110.0	70.7	133.0	18.4	3,861.5	362.7	6,355.5	0.7
Blank Filter	60.0	1.4	47.0	2.8	146.0	30.4	137.0	11.3
Positive control	927.0	101.1	1,518.0	352.8	908.0	173.9	2,344.0	7
Potency revertants/m³	100		6.3		3,500		1,100	

Doses	Mean of Number of Revertants/plate and <i>Standard Deviation (SD)</i>							
m ³ /plate	TA1538				YG5161			
	-S9		+S9		-S9		+S9	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Negative control	15.0	2.0	12.7	2.5	26.3	4.0	28.0	2.65
0.02	16.5	0.7	14.5	2.1	23.5	0.7	29.5	7.8
0.2	28.0	2.8	18.5	2.1	43.0	7.1	34.0	7.1
1	89.5	0.7	52.0	9.9	99.0	2.8	57.5	2.1
2	146.0	18.3	69.0	14.1	136.0	4.2	85.5	3.5
5	306.5	24.7	147.5	13.4	272.0	4.2	162.0	7.1
10	429.0	38.2	209.0	1.4	395.0	8.5	179.0	28.3
Blank Filter	20.0	1.4	21.0	9.2	18.0	0.7	29.0	6.4
Positive control	392.0	50.2	2,451.0	746.0	677.0	111.0	3,114.0	514.1
Potency revertants/m³	62		28		61		27	

*Shaded values represent the number of revertants that equal or more than twice the negative controls

TABLE IV. Mutagenicity data for STHOLM PM extracts of the Salmonella/microsome microsuspension assay with TA98, YG1041, TA1538 and YG5161 without and with S9.

Doses		Mean of Number of Revertants/plate and <i>Standard Deviation (SD)</i>							
m ³ /plate		TA98				YG1041			
		-S9		+S9		-S9		+S9	
		Mean	SD	Mean	SD	Mean	SD	Mean	SD
Negative control		52.7	4.5	53.3	14.4	157.7	17.1	145.7	9.1
0.02		58.5	6.4	51.5	3.5	173.5	6.4	157.0	5.7
0.2		61.0	12.7	43.0	1.4	364.5	2.1	173.5	16.3
1		73.0	1.4	50.0	5.7	936.0	135.7	418.5	4.9
2		93.0	14.1	58.0	8.5	2,108.0	159.8	683.5	37.5
5		195.0*	59.4	70.5	3.5	3,790.0	181.0	1,059.5	33.2
10		728.0	31.1	98.5	0.7	4,695.0	29.7	2,885.5	191.6
Blank filter		50.0	11.3	57.0	14.8	178.0	24.0	153.0	3.5
Positive control		927.0	101.1	1,518.0	352.8	908.0	173.9	2,344.0	171.1
Potency revertants/m³		25		4.8		890		270	

Doses		Mean of Number of Revertants/plate and <i>Standard Deviation (SD)</i>							
m ³ /plate		TA1538				YG5161			
		-S9		+S9		-S9		+S9	
		Mean	SD	Mean	SD	Mean	SD	Mean	SD
Negative control		15.0	2.0	12.7	2.5	26.3	4.0	28.0	2.6
0.02		15.0	1.4	11.5	3.5	26.5	2.1	20.0	0.0
0.2		24.0	2.8	20.5	3.5	27.5	9.2	28.5	2.1
1		38.0	0.0	30.0	4.2	40.0	0.0	36.0	2.8
2		66.0	8.5	47.0	1.4	55.5	0.7	58.0	2.8
5		162.5	9.2	93.5	19.1	129.0	5.6	101.5	16.3
10		264.5	41.7	117.0	1.4	213.5	9.2	143.0	5.7
Blank filter		14.0	4.2	18.0	7.1	26.0	2.8	25.0	7.8
Positive control		392.0	50.2	2,451.0	746.0	677.0	111.0	3,114.0	514.1
Potency revertants/m³		26		12		18		15	

*Shaded values represent the number of revertants that equal or more than twice the negative controls

TABLE V. Mutagenic responses in the *Salmonella*/microsome microsuspension assay for benzo[a]pyrene with the strains, TA98, YG1041, TA1538 and YG5161 in the presence of S9.

Doses ng/plate	Mean of Number of Revertants/plate and <i>Standard Deviation (SD)</i>							
	TA98		YG1041		TA1538		YG5161	
	Mean	<i>SD</i>	Mean	<i>SD</i>	Mean	<i>SD</i>	Mean	<i>SD</i>
Negative control	28.0	7.1	159.0	21.2	15.5	0.7	21.0	5.7
0.5							32.0	7.1
1.0							28.5	0.7
2.0							34.0	2.8
4.0							48.5	3.5
7.8	32.5	3.5	164.5	29.0	14.5	0.7	47.5	2.1
15.6	31.0	1.7	183.0	35.4	25.0	5.7	71.5	6.4
31.2	37.5	3.5	210.5	10.6	30.5	4.9	114.0	1.4
62.5	30.5	7.8	214.0	21.2	44.0	17.0	110.5	12.0
125	37.5	9.2	306.0	31.1	49.5	12.0	155.0	8.5
250	42.5	6.4	339.0	38.2	68.0	4.2	203.5	19.1
500	39.0	5.7	462.5	17.7	98.5	10.6	230.0	5.7
1,000	60.0*	14.1	471.0	9.9	125.5	27.6	285.0	1.4
2,000	63.0	11.3	470.5	0.7	106.0	5.7	271.5	2.1
Potency revertants/ng	0.02		0.6		0.2		3.0	
MED**	1,400		265		77.5		7	

*Shaded values represent the number of revertants that equal or more than twice the negative controls

**MED = Minimal Effective Dose in ng/plate

TABELA VI. Mutagenic responses in the *Salmonella*/microsome microsuspension assay for 1-nitropyrene with the strains, TA98, YG1041, TA1538 and YG5161 in the absence of S9.

Doses ng/plate	Mean of Number of Revertants/plate and <i>Standard Deviation (SD)</i>							
	TA98		YG1041		TA1538		YG5161	
	<i>Mean</i>	<i>SD</i>	<i>Mean</i>	<i>SD</i>	<i>Mean</i>	<i>SD</i>	<i>Mean</i>	<i>SD</i>
Negative control	42.5	4.9	201.0	21.2	18.0	0	24.0	4.2
0.01			257.5	10.6				
0.02			310.5	23.3				
0.03			383.0	35.4				
0.04			410.5	2.1				
0.05			506.5	24.7				
0.1	48.0	2.8	791.5	3.5	18.5	0.7	33.5	10.6
0.3	58.0	15.6	3,063.5	101.1	17.5	7.8	29.0	5.7
0.7	101.0*	7.1	4,353.0	110.3	28.5	6.4	45.5	0.7
1.0	129.5	0.7			19.0	5.7	36.0	5.7
2.5	266.0	41.0			36.5	3.5	68.0	11.3
5.0	536.5	140.7			63.5	0.7	76.5	13.4
10	1,001.0	75.0			139.5	46.0	135.0	33.9
20	1,883.5	82.7			595.0	110.3	334.0	19.8
Potency revertants/ng	94		5,900		10		13	
MED**	0.45		0.03		1.8		1.8	

*Shaded values represent the number of revertants that equal or more than twice the negative controls

**MED - Minimal Effective Dose in ng/plate