



**Karolinska  
Institutet**

Karolinska Institutet

<http://openarchive.ki.se>

---

This is a Peer Reviewed Accepted version of the following article, accepted for publication in *Current Drug Targets*.

2015-12-23

# Targeting receptor tyrosine kinases using monoclonal antibodies : the most specific tools for targeted-based cancer therapy

Shabani, Mahdi; Hojjat-Farsangi, Mohammad

---

*Curr Drug Targets*. 2016;17(14):1687-1703.

<http://doi.org/10.2174/1389450116666151001104133>

<http://hdl.handle.net/10616/44966>

*If not otherwise stated by the Publisher's Terms and conditions, the manuscript is deposited under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives License (<http://creativecommons.org/licenses/by-nc-nd/4.0/>), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited, and is not altered, transformed, or built upon in any way.*

## **Targeting Receptor Tyrosine Kinases using Monoclonal Antibodies: The Most Specific Tools for Targeted-Based Cancer Therapy**

**Mahdi Shabani<sup>1</sup> and Mohammad Hojjat-Farsangi<sup>2,3</sup>**

<sup>1</sup> Monoclonal Antibody Research Center, Avicenna Research Institute, ACECR, Tehran, Iran

<sup>2</sup> Department of Oncology-Pathology, Immune and Gene therapy Lab, Cancer Center Karolinska (CCK), Karolinska University Hospital Solna and Karolinska Institute, Stockholm, Sweden

<sup>3</sup> Department of Immunology, School of Medicine, Bushehr University of Medical Sciences, Bushehr, Iran

### **Corresponding author:**

Mohammad Hojjat-Farsangi, Ph.D, Assistant Professor, Department of Oncology-Pathology, Cancer Center Karolinska (CCK), Karolinska University Hospital, Solna, SE-171 76 Stockholm, Sweden, Tel: +46 8 51 77 43 08, Fax: +46 8 31 83 27, Email: mohammad.hojat-farsangi@Ki.se

**Running title:** Monoclonal antibodies and RTKs targeted therapy

**Abstract:**

Receptor tyrosine kinases (RTKs) family is comprised of different cell surface glycoproteins. These enzymes participate and regulate vital processes such as cell proliferation, polarity, differentiation, cell to cell interactions, signaling, and cell survival. Dysregulation of RTKs contributes to the development of different types of tumors. RTKs deregulation in cancer has been reported for more than 30 RTKs. Due to critical roles of these molecules in cancer, the specific targeting of RTKs in malignancies is a promising approach. Targeted cellular and molecular therapies have been known as a new type of therapeutics, preventing tumor cell proliferation and invasion by interrupting with molecules necessary for tumor growth and survival. Specific targeting of RTKs using monoclonal antibodies (mAbs) in malignancies as well as in autoimmune disorders is of great interest. The growing number of mAbs approved by the authorities implies on the increasing attentions and applications of these therapeutic tools. Due to the high specificity, mAbs are the most promising substances that target RTKs expressed on the tumor cell surface. In this communication, we review the recent progresses in development of mAbs targeting oncogenic RTKs for cancer treatment.

**Keywords:** Cancer, monoclonal antibody, receptor tyrosine kinase, targeted therapy

## INTRODUCTION

Current chemotherapy and radiotherapy modalities and the combinational regimens have improved the life quality of cancer patients. In spite of considerable progresses in the management and treatment of cancer patients; however, their applications have severe side effects on normal cells. Therefore, developing more specific drugs is important to overwhelm the current shortcomings of these modalities. In this context, targeted-based cancer therapy (targeted therapy) agents have significantly progressed and several drugs have got the approval of authorities for application in cancer therapy.

The purpose of targeted cancer therapy is destroying tumor cells by targeting antigens expressed by the cells. Among several targets, receptor tyrosine kinases (RTKs) have distinct properties that make them suitable targets for therapy. RTKs have special biological features and structure for signal transduction [1]. RTKs with oncogenic property have no or minimum activity in normal cells, however their deregulations are seen in cancerous cells. RTK-like orphan receptor 1 (Ror1) is an example that is expressed at a noteworthy level during embryogenesis in neuronal and other fetal tissues [2]. It is also overexpressed on the surfaces of different tumor cells, including chronic lymphocytic leukemia (CLL) [3-5], acute myelogenous leukemia (AML), acute lymphoid leukemia (ALL), mantle cell lymphoma (MCL) [6], hairy cell leukemia (HCL) [6], melanomas [7, 8], and lung cancer [9].

The RTKs roles have been extensively investigated in progression and metastasis of cancer [10, 11]. Aberrant RTKs expression and activation are linked to cancer development, transformation and metastasis [12-14]. Currently, different RTKs are under intensive research for targeting tumor cells [15].

Monoclonal antibodies (mAbs) are the most specific and ideal tools for targeting cell-surface antigens expressed by tumor cells. Due to the high specificity of mAbs, they have fewer side effects compare to other agents such as cytotoxic drugs and small molecule inhibitors (SMIs). Furthermore, mAbs have different mechanism of action to destroy the targeted cells. Some antibodies disrupt the kinase signaling through inhibiting ligands and receptor internalization as well as preventing homo/hetero-dimerization of RTKs, which might result in direct apoptosis. Other mechanisms include effector cells activation [(antibody-dependent cell-mediated cytotoxicity (ADCC)], complement activation [complement dependent cytotoxicity (CDC)] and direct necrosis of malignant cells [4, 16-18].

Currently, several anti-RTK mAbs have shown proper therapeutic effects and thus were approved for cancer therapy. The most famous approved mAbs for targeting RTKs are specific to human epidermal growth factor receptor (EGFR), HER2 (ErbB2) and VEGFRs (Table 1). In this paper: 1) the structure and function of selected RTKs appropriate for mAb therapy, 2) current mAbs applied for cancer therapy, and 3) promising mAbs targeting oncogenic RTKs in preclinical or clinical settings have been described.

Table 1. Oncogenic RTKs for mAb targeted cancer therapy.

RTK	Chromosome location	Mechanism of overexpression	Malignancies (examples)	Development phase	
				Preclinical	Clinical trials
ALK	2p23	Translocation	NSCLC, colorectal carcinoma, breast, oesophageal and renal cell cancers	+	+
AXL	19q13.1	overexpression	AML, CML, NSCLC, lung, colon, breast, esophageal, thyroid and pancreatic cancers, gastrointestinal stromal tumors, astrocytoma-glioblastoma	+	-
CCK4	6p21.1	Mutation	Squamous cell carcinoma, small cell lung, breast, colon and gastric cancers, AML	+	-
DDR1	6p21.33	Mutation, overexpression	NSCLC, breast and ovarian cancers, hepatocellular carcinoma, AML	+	-
DDR2	1q23.3	Mutation	Head and neck squamous cell carcinoma, breast and lung cancers, NSCLC, CML	+	-
EGFR1	7p11.2	Mutation, overexpression	Breast cancer, hepatocellular and head and neck squamous cell carcinomas	+	+
EGFR2	17q12	Mutation, overexpression	Breast cancer, gastric adenocarcinomas	+	+
EGFR3	12q13.2	Mutation, overexpression	Breast cancer	+	-
EGFR4	2q34	Mutation, overexpression	Breast cancer, melanoma	+	-
EPHA1	7q35	Mutation, overexpression	NSCLC, prostate cancer, esophageal squamous cell carcinoma	+	-
EPHA2	1p36.13	Mutation, overexpression	Hepatocellular and colorectal carcinomas, breast cancer, osteosarcoma	+	-
EPHA3	3p11.1	Mutation, overexpression	Glioblastoma, lung cancer, melanoma, ALL, T-cell leukemia, Hodgkins lymphoma	+	-
EPHA4	2q36.1	Mutation	NSCLC, gastric cancer	+	-
EPHA5	4q13.1	Mutation	Breast cancer, hepatocellular carcinoma, ALL	+	-
EPHA6	3q11.2	Mutation	Renal cell carcinoma	+	-
EPHA7	6q16.1	Mutation, overexpression	Hepatocellular carcinoma	-	-
EPHA8	1p36.12	Mutation, overexpression	Colorectal carcinoma, liver tumors	-	-
EPHA10	1p34.3	Mutation, overexpression	Breast cancer, CLL		
EPHB1	3q22.2	Mutation, overexpression	NSCLC, cervical and ovarian cancers	+	-
EPHB2	1p36.12	-	Cervical and breast cancers, melanomas, hepatocellular carcinoma	+	-
EPHB3	3q27.1	-	NSCLC, breast cancer, colorectal carcinoma	+	-
EPHB4	7q22.1	Mutation, overexpression	Breast, melanoma and glioma cancers	+	+
EPHB6	7q33-q35	Mutation, overexpression	CLL, NSCLC, breast cancer		

FGFR1	8p12	Mutation	Lung and breast cancers	+	+
FGFR2	10q26.13	Mutation, overexpression	Lung, breast, thyroid, prostate cancer, cholangiocarcinoma, astrocytoma	+	+
FGFR3	4p16.3	Mutation	Bladder cancer, lung and head carcinomas	+	+
IGF1R	15q26.3	Mutation, overexpression	CLL, breast and pancreatic cancers, hepatocellular and oral squamous cell carcinomas	+	+
IGF2R	6q25.3	Mutations, overexpression	Hepatocellular, squamous cell and colorectal carcinomas, breast, pancreatic and prostate cancers, NSCLC	+	+
FLT3	13q12.2	Mutation, overexpression	AML, acute promyelocytic leukemia	+	+
INSR	19p13.2	Mutation, overexpression	Colorectal carcinoma, prostate cancer	+	+
INSRR	1q23.1	Mutation, overexpression	Neuroblastomas	+	+
KIT	4q12	Mutation	AML, melanoma, ovarian carcinoma, gastrointestinal stromal tumors	+	+
LTK	15q15.1	Mutation	Gastric cancer, lymphomas and leukemias	+	+
MER	2q13	Mutation	Glioblastoma, hepatocellular carcinoma, astrocytoma	+	+
MET	7q31.2	Mutation, overexpression	Hepatocellular carcinoma, CLL, breast, pancreatic and lung cancers, gastric adenocarcinoma	+	+
MUSK	9q31.3	Mutation, overexpression	Ovarian cancer	-	-
NTRK1	1q21-22	Translocation, mutation, overexpression	Thyroid and breast cancers, lung adenocarcinoma, colorectal and oral squamous cell carcinomas	+	+
NTRK2	9q22.1	Translocation	Neuroblastoma, astrocytoma, oral squamous cell carcinoma	+	+
NTRK3	15q25.3	Translocation	Neuroblastoma, breast cancer	+	+
PDGFRA	4q12	Mutation	Lung adenocarcinoma, gastrointestinal stromal tumors	+	+
PDGFRB	5q32	Mutation	Gastrointestinal stromal tumors, glioblastoma	+	+
RET	10q11.2	Mutation	NSCLC, medullary thyroid carcinoma	+	+
RON	3p21.31	Mutation, overexpression	Pancreatic and breast cancers, NSCLC, laryngeal and head and neck squamous cell carcinomas	+	+
ROR1	1p31.3	Overexpression	CLL, ALL, AML, MCL, HCL, melanoma, prostate, lung, breast, pancreatic, colon, ovarian, and uterus cancers	+	+
ROR2	9q22.31	Mutation	Melanoma, medulloblastoma, leiomyosarcoma, gastrointestinal stromal tumor, hepatocellular, renal cell and head and neck carcinomas, osteosarcoma, prostate, testicular, colon cancers	+	-
ROS1	6q22	Deletion, inversion, translocation	NSCLC, cholangiocarcinoma, ovarian and gastric cancers, colorectal carcinoma	+	-
RYK	3q22.2	Translocation, mutation	CML, ovarian cancer	+	-
TEK	9p21.2	Mutation	Bladder cancer, glioblastoma, AML	+	-
TIE	1p34.2	Mutation	Glioblastoma	+	-
TYRO3	15q15.1	Mutation	Melanoma, colon, thyroid, and breast cancers	+	-
VEGFR1	13q12.3	Mutation, overexpression	Ovarian cancer, NSCLC, colorectal carcinoma	+	+
VEGFR2	4q12	Mutation, overexpression	Renal cell and hepatocellular carcinomas	+	+
VEGFR3	5q35.3	Mutation, overexpression	Ovarian and gastric cancers, bladder carcinoma	+	+

ALK: anaplastic lymphoma kinase, ALL: acute lymphoblastic leukemia, AML: acute myeloid leukemia, CLL: chronic lymphocytic leukemia, CML: chronic myeloid leukemia, DDR: discoidin domain receptor, EGFR: epidermal growth factor receptor, EPH: ephrin receptors, FGFR: fibroblast growth factor receptor, IGFR: insulin growth factor receptor, INSR: insulin receptor, LTK: leukocyte tyrosine kinase, MCL: mantle cell lymphoma, HCL: hairy cell leukemia, MuSK: muscle-specific kinase, NSCLC: non-small cells lung carcinoma, NTRK: neurotrophic tyrosine kinase, PDGFR: platelet-derived growth factor receptor, ROR: receptor tyrosine kinase-like orphan receptor, RYK: receptor related to tyrosine kinases, VEGFR: vascular endothelial growth factor receptor [19].

## **RECEPTOR TYROSINE KINASES (RTKs)**

The large family of RTKs consists of cell surface receptors discovered more than 25 years ago [20] which are divided into 20 sub-families with 58 members [21]. These receptors are involved in crucial processes, for instance in cell proliferation, differentiation, cell-cell interaction, migration, signaling, metabolism and survival [22]. RTKs structure consists of 3 distinct regions, including extracellular, transmembrane and cytoplasmic domains [23]. The extracellular part of RTKs contains the ligand binding site that interacts with the surface, soluble and extracellular matrix proteins [24]. This part is also involved in the RTKs dimerization that is critical for the activation of tyrosine kinase (TK) in the cytoplasmic portion of the RTKs [14, 19, 25]. The extracellular regions of RTKs have a combination of several globular domains, including immunoglobulin (Ig)-like, cysteine-rich, fibronectin type III-like, EGF-like, and Sema domains [23]. The cytoplasmic part consists of a lipophilic transmembrane helix and is followed by different regions such as the TK domain and carboxy-terminal region. Several serine and tyrosine residues at the cytoplasmic region are phosphorylated following ligand binding to extracellular part. Indeed, these serine and tyrosine residues serve as docking sites for proteins having Src homology (SH) 2 domain to regulate RTKs catalytic function [14, 19, 26]. Among different regions of RTKs, the TK domain shows the highest conservation level [27]. Adenosine triphosphate (ATP) binding site mutations might change the activity of receptor and consequently inactivate the biological function of the receptor that might change the RTK to pseudo-RTK without or with low enzymatic activity [26, 28]. The carboxy-terminal tail of RTKs is the most distal and non-catalytic part. This region has the highest degree of heterogeneity in length and sequence, even among members of the same RTK subclass. The carboxy-terminal tail contains tyrosine residues that are phosphorylated by intracellular kinases. Some reports have shown the crucial role of C-terminal part in modulating kinase activity that provides proper conformation for TK domain [26].

Aberrant activation of RTKs owing to receptor overexpression, gene amplification, impaired downregulation, chromosomal translocation, and mutations contribute to cancer development and progression [19, 29, 30]. RTKs dysregulation have been reported for more than 30 RTKs (Table 1) [13]. These changes in the structure of RTKs make these receptors become potent oncoproteins, leading to the neoplastic alteration [13, 30].

In the following sections, different RTKs that are proper for mAb targeting have been described.

## RTKs: PROPER TARGETS FOR CANCER THERAPY

### EPIDERMAL GROWTH FACTOR RECEPTOR (EGFR)

EGFR (ErbB-1/HER1), HER2/neu (ErbB-2), HER3 (ErbB-3), and HER4 (ErbB-4) are EGFR (ErbB) family members that play important roles in the normal cell functions regulation. Overexpression and mutation are involved in EGFR abnormal activation that is related to several tumors development. Therefore, specific inhibition of EGFR is one of the key targets for cancer treatment. In this context various mAbs have been approved for targeting solid tumors that overexpress the members of this family. Due to the overexpression of EGFR family members, mAbs targeting these RTKs are appropriate drugs for the treatment of several malignancies (Table 2) [31, 32].

Cetuximab (Erbix) is a chimeric anti-EGFR mAb approved on February 12, 2014 for metastatic colorectal carcinoma (mCRC) patients refractory to irinotecan-based chemotherapy. Cetuximab was the first mAb approved to treat this type of cancer and is indicated in combination therapy with irinotecan, a chemotherapy drug approved for colorectal cancer treatment, or alone if irinotecan is intolerable for patients [33]. Cetuximab is prescribed as second- or third-line of therapy in mCRC [33] and the head and neck squamous cell carcinoma [32]. Moreover, on July 6, 2012, the FDA approved cetuximab in combination with chemotherapy drugs (irinotecan, 5-fluorouracil, and leucovorin) for application as first-line treatment of EGFR-expressing mCRC patients having wild-type *K-ras* (mutation-negative) (<http://www.fda.gov>). Cetuximab inhibited the binding of activating ligand to EGFR and also prevented receptor dimerization, leading to disruption of the signal transduction cascade [34].

Table 2. Current therapeutic monoclonal antibodies for targeted-based cancer cell therapies.

Name	Trade name	Target	Antibody format	Malignancy	Stage for treatment		
					Preclinical	Clinical trial	Approved for treatment
AVE1642	ND	IGF1R	Humanized IgG1	NSCLC, multiple myeloma, Ewing's sarcoma	+	I	NY
Bevacizumab	Avastin	VEGF	Humanized IgG1	Glioblastoma, NSCLC, metastatic colon and kidney cancer	+	I-IV	+
Cetuximab	Erbix	EGFR	Chimeric IgG1	Head and neck squamous cell carcinoma, MCC	+	I-IV	+
Cirtuzumab (UC-961)	ND	ROR1	Humanized IgG1	CLL	+	I	+



<b>Cixutumumab (IMC-A12)</b>	ND	IGF1R	Fully human IgG1	Thymic carcinoma, soft tissue sarcomas, osteosarcoma, breast cancer, Ewing's sarcoma	+	I-II	NY
<b>Dalotuzumab (MK-0646)</b>	ND	IGF1R	Humanized IgG1	Advanced colorectal carcinoma, NSCLC	+	I-II	NY
<b>Figitumumab (CP-751871)</b>	ND	IGF1R	Fully human IgG1	Adrenocortical carcinoma, NSCLC, multiple myeloma	+	I	NY
<b>Ganitumab</b>	ND	IGF1R	Fully human IgG1	Metastatic solid tumors	+	I-II	NY
<b>Narnatumab (IMC-RON8)</b>	ND	RON	Humanized IgG1	Advanced solid tumors	+	I	NY
<b>Onartuzumab (MetMab)</b>	ND	MET	Humanized IgG1	Advanced NSCLC	+	II	NY
<b>Panitumumab</b>	Vectibix	EGFR	Fully human IgG1	Metastatic colon cancer	+	I-IV	+
<b>Pertuzumab</b>	Perjeta	HER2	Humanized IgG1	Metastatic breast cancer	+	I-IV	+
<b>PF-03446962</b>	ND	ALK1	Humanized IgG2	HCC, Advanced malignant pleural Mesothelioma, relapsed or refractory urethelial cancer	+	NY	NY
<b>RG1507</b>	ND	IGF1R	Fully human IgG1	Metastatic NSCLC	+	I-II	NY
<b>Robatumumab (SCH717454)</b>	ND	IGF1R	Humanized IgG1	Advanced colorectal carcinoma, NSCLC	+	I-II	NY
<b>Trastuzumab</b>	Herceptin (Herclon)	HER2	Humanized IgG1	Breast cancer, gastric adenocarcinoma, gastroesophageal junction adenocarcinoma	+	I-IV	+
<b>Trastuzumab emtansine</b>	Kadcyla	HER2	Humanized IgG1	Advanced breast cancer	+	I-IV	+

CLL: chronic lymphocytic leukemia, EGFR: epidermal growth factor receptor, HCC: hepatocellular carcinoma, HER: human epidermal growth factor receptor, IGF1R: insulin growth factor receptor, MCC: metastatic colorectal carcinoma, ND: not defined, NSCLC: non-small cells lung carcinoma, NY: not yet, ROR: receptor tyrosine kinase-like orphan receptor, VEGFR: vascular endothelial growth factor receptor

Panitumumab (Vectibix) is a fully human mAb specific for EGFR used as second- or third-line of the treatment for mCRC patients akin to cetuximab [33, 35]. This mAb was approved on September 27, 2006, for EGFR-expressing mCRC cases with cancer progression or following chemotherapy regimens containing fluoropyrimidine, oxaliplatin, and irinotecan (<http://www.fda.gov/>). Panitumumab in combination with chemotherapy might also be useful for NSCLC patients treatment [36]. The mechanism of action is similar to cetuximab, but it does not promote ADCC [37]. Panitumumab in combination with folinic acid, fluorouracil, oxaliplatin has also been approved as first-line therapy in mCRC patients [38].

The ErbB-2/HER2, another EGFR family member, has intensively been investigated as an important RTK which is overexpressed and/or hyperactivated in various malignancies. The extracellular region of the receptor is divided into four domains. No ligand has been recognized for HER2. HER2 overexpression can transform cells in a ligand-independent manner which

makes it unusual member in ErbB family [39]. The HER2 importance in targeted therapy is highlighted by several molecular and pathological outcomes. HER2 amplification is related to the processes of tumorigenesis and pathologic features such as tumor size, invasion, and metastatic spread. HER2 has higher expression and activity in metastatic tumors than in non-aggressive tumors [40]. HER2 overexpression is found in 10-34% of invasive breast cancers with a poor prognosis [41]. Furthermore, overexpression of HER2 has been detected in several tumors such as lung, ovary, salivary gland, prostate, colon, and pancreatic cancers [40] as well as in hematologic malignancies such as ALL and AML [42].

Trastuzumab (Herceptin) was the first anti-HER2 mAb approved for application in HER2<sup>+</sup> breast cancer patients [43]. Trastuzumab is a humanized IgG1 mAb and has shown a 35% response rate in metastatic breast cancer (MBC) patients who received no earlier chemotherapy [44]. A phase III clinical trial on MBC patients with HER2 amplification has showed that trastuzumab with chemotherapy combinational regimen was associated with a few months delay in disease progression (median, 7.4 vs. 4.6 months), a higher rate of objective response (50% vs. 32%) and survival (median, 25.1 vs. 20.3 months) [45]. Trastuzumab was approved on September 25, 1998 by the FDA for MBC patients with HER2 protein overexpression and who have received chemotherapy drugs for metastatic disease treatment (<http://www.fda.gov/>). In January 2010, the European Medicines Agency (EMA) approved trastuzumab in combination with chemotherapy for the treatment of metastatic stomach adenocarcinoma or gastroesophageal junction with HER2 overexpression [46]. Trastuzumab acts via inhibition of receptor homo/hetero-dimerization, internalization and endocytic destruction [47, 48]. Although trastuzumab is well-accepted as the standard drug in the breast cancer therapy, up to 40% of MBC patients do not respond to trastuzumab and in those who respond, the median progression time is less than one year [49, 50]. Moreover, acquired trastuzumab resistance is a serious concern ending in disease progression [50, 51]. In general, these limitations call for design of new and superior mAbs for MBC therapy. Pertuzumab (Perjeta) is a newly approved anti-HER2 mAb that prevents ligand-dependent HER2:HER3 dimerization and reduces intracellular signaling pathways [52]. Pertuzumab received approval for the treatment of HER2<sup>+</sup> MBC patients in June 8, 2012. The combination of pertuzumab, trastuzumab and docetaxel has been found to have an overall survival benefit in HER2<sup>+</sup> MBC patients used in the first-line setting [53]. Recently, the FDA approved the combination of trastuzumab, pertuzumab, and docetaxel as first-line treatment for MBC patients

(<http://www.fda.gov/>). At present, pertuzumab in combination with trastuzumab and platinum-fluoropyrimidine is under study in a phase II clinical trial as a first-line therapy in gastric cancer [54].

Fourth HER2-targeted agent, trastuzumab emtansine (T-DM1, Kadcyła) was approved on February 22, 2013, as a single agent for HER2<sup>+</sup> MBC patients, who have received trastuzumab and/or a taxane [55].

## **VASCULAR ENDOTHELIAL GROWTH FACTOR (VEGF)**

VEGF-A (usually referred to VEGF), VEGF-B, VEGF-C, VEGF-D, and placental growth factor are members of VEGF family that are expressed by various solid tumors [56]. Members of this family induce angiogenesis through binding to VEGFR1-3 expressed on the vascular endothelium. Angiogenesis is defined as a controlled process accountable for new blood vessels formation and is correlated to advanced-stage disease as well as clinically aggressive tumor subtypes [56]. Angiogenesis regulation is achieved by a variety of endogenous activators and inhibitors during the normal physiological processes such as the menstrual cycle and wound healing [57]. VEGF is predominate stimulator of angiogenesis during physiological condition such as embryogenesis, skeletal growth and reproductive functions as well as pathological angiogenesis associated with solid tumors [56]. Inhibition of angiogenesis prevents tumor expansion, survival and metastasis. In this perspective, blocking tumor cells angiogenesis is the main focus of several drug discoveries for solid tumors.

Bevacizumab (Avastin), an angiogenesis inhibitor, is an anti-VEGF humanized mAb that prevents binding of VEGF to VEGFRs and was approved for the treatment of MBC [58], mCRC [59] and NSCLC [60] patients in combination with cytotoxic chemotherapy [37, 61]. Bevacizumab is also used in combination with IFN- $\alpha$  in metastatic renal cell carcinoma therapy [62, 63]. Combination of bevacizumab and chemotherapy drugs has improved the survival of patients with mCRC and NSCLC compared to chemotherapy alone [64, 65]. Bevacizumab increased overall survival of mCRC patients 4.7 and 2.1 months following first-line and second-line therapies, respectively as described in a phase III clinical trial [66]. The increasing use of bevacizumab has resulted in the appearance of bevacizumab-resistant tumor cells as a result of

the upregulation of other angiogenic factors [67]. These findings proposed requisite for targeting cancer cells via multi-targets to inhibit compensatory mechanisms in tumor escape.

### **INSULIN GROWTH FACTOR RECEPTOR (IGFR)**

The insulin-like growth factor 1 receptor (IGF1R) is a member of RTK family ubiquitously expressed in most human cells with the exception of hepatocytes and mature B lymphocytes [68]. The IGF1R is activated by its ligands insulin-like growth factor (IGF)-1 and -2 and plays important roles in cell proliferation control in mammalian cells *in vitro* and *in vivo*, regulation of lipids, proteins, and carbohydrates metabolism and the maintenance of glucose homeostasis [69, 70]. IGF-1 has a critical function in hypertrophy of skeletal muscle and other target tissues mainly through IGF1R. Mice lacking IGF1R die in late stage of development showing the strong growth-promoting effect of this RTK [69].

The IGF1R is dysregulated in several cancers such as lung, prostate and breast cancers [71]. The anti-apoptotic effects of IGF1R raised tolerance to SMIs and cytotoxic drugs. In breast cancer, IGF1R expression induced tumor cell resistance to EGFR inhibitors like erlotinib. It is described that crosstalk between EGFR and IGF1R and the common signaling pathway are the main reason for tumor cell resistance [72]. Importantly, in prostate cancer IGFIR signaling is crucial for cell survival and growth when cancer cells progress to androgen independence [73].

Currently, several anti-IGF1R mAbs are under study for application in solid tumors treatment. Ganitumab (AMG 479) is one of the successful mAbs for targeting IGF1R<sup>+</sup> tumors. The safety, maximum-tolerated dose (MTD), pharmacokinetics, and evidence of anti-tumor activity have been studied in several trials [74]. Fatigue, fever, thrombocytopenia, chills, rash, and anorexia were the most common adverse effects. A maximum dose of 20 mg/kg was shown to be safe and anti-tumor effects were promising [74]. A phase II study evaluated the ganitumab monotherapy safety and efficacy in metastatic Ewing family or desmoplastic small round cell tumors. Of 35 subjects examined for response, partial response and disease stability were observed at 5% and 48% of patients, respectively [75]. In metastatic pancreatic cancer, the ganitumab efficacy and safety were evaluated in a phase II clinical trial. Ganitumab was well-tolerated and improved 6-month overall survival rate in 57% of patients [76].

Cixutumumab (IMC-A12) is a fully human IgG1 mAb specific to the IGF1R with potential anti-tumor effects. This mAb prevented the binding of the IGF-1 to IGF1R and inhibited the activation of phosphoinositol 3-kinase (PI3K)/Akt signaling pathway and reduced the survival of cancer cell followed by induction of apoptosis [77]. The effects were tested in several cancers, including thymic, head and neck and hepatocellular carcinomas, soft tissue sarcomas, osteosarcoma, breast, prostate, islet cell, and pancreatic cancers (Table 2) [77].

RG1507 is another fully human IgG1 mAb specific for IGF1R which was assessed in a phase I trial in children with relapsed or refractory solid tumors. Although, dose-limiting toxicities were not observed, no objective responses were seen [78]. Dalotuzumab (MK-0646) [79], figitumumab (CP-751871) [80], robatumumab (SCH717454) [81], and AVE1642 [82] are other anti-IGF1R mAbs that have been evaluated in several malignancies in preclinical stages with promising results.

## **Axl RECEPTOR TYROSINE KINASE**

Axl is a member of the TAM (Tyro3, Axl and Mer) family [83]. The TAM members are distinguished by having a conserved sequence in the TK domain and adhesion molecule-like domains in the extracellular part [83]. Almost the entire ectodomain of TAM members comprise of Ig-like and fibronectin type III domains. These structures are vital in cell to cell contacts and are similar to the structure of neural cell adhesion molecule which consists of five Ig domains and two fibronectin type III domains [83]. TAM members are involved in the clearance of apoptotic cells. Protein S and growth arrest-specific factor 6 (GAS6), bind to the apoptotic cells surface and directly bind to TAM members expressed on phagocytes and finally engulf and clear apoptotic cells [84]. Axl is overexpressed in several cancers, including Burkitt's lymphoma [85], CLL [86], hepatocellular carcinoma [87], multiple myeloma [88], breast [89], and pancreatic cancers [90] and is correlated with cancer poor prognosis [91]. Axl is involved in proliferation and invasion of tumor cells, mainly in pancreatic carcinoma [90].

Currently, anti-Axl mAbs are under investigation in preclinical stages. Two anti-Axl mAbs (D9 and E8), have been shown to hamper Axl activity and stopped pancreatic cancer cells proliferation and migration. These mAbs inhibited Axl phosphorylation and Akt pathway

downstream with no effect on GAS6 binding, reduced the expression of Axl due to internalization in pancreatic cells. Anti-Axl mAbs therapy in xenografted mice declined subcutaneous and orthotopic pancreatic tumor xenografts growth [92].

## **DISCOIDIN RECEPTOR (DDR)**

Discoidin receptors (DDR1 and DDR2) are two closely related RTKs that contain a discoidin homology domain in their extracellular regions [93, 94]. The DDRs were firstly identified by homology cloning based on their catalytic kinase domains, and then different types of collagen were recognized as DDR functional ligands [93, 95].

Human DDR1 coding gene is located to chromosome 6 between the HLA-E and HLA-C genes [96]. Structurally, DDR1 has a discoidin homology domain which contains the collagen binding site, a discoidin-like domain which involves in collagen-induced receptor activation. DDR1 transmembrane domain mediates collagen-independent receptor dimerization and a large intracellular part with phosphorylated tyrosines that may serve as docking sites for DDR binding proteins and a TK domain [97].

DDRs play roles in the regulation of critical cellular processes like cell differentiation, proliferation, migration and survival [98]. They control remodeling of extracellular matrixes by modulating the expression and activation of matrix metalloproteinase (MMP) [98]. Indeed DDRs are exclusively located on the cells to be sensors for extracellular matrix and also to be a regulator of a wide range of cell functions as mentioned above [97].

The DDR1 transcript is expressed in several normal tissues, restricted to epithelial cells with the highest level in lung, brain, spleen, kidney, and placenta [98-100]. Mutations and altered expression of DDRs are observed in many cancer types implies that they might be involved in the development and progression of cancers [97]. DDR1 upregulation has been reported in several types of cancers for instance in NSCLC [101], primary and MBC [99], brain tumors [102], ovarian [103, 104], and prostate cancers [105]. It is worthy to consider that findings related to the DDR1 expression encounters conflicting results in breast cancer [106]. DDR1 can be inhibited by imatinib, ponatinib and DDR1-IN-1 (selective type II inhibitor) inhibitors may have application in malignancies such as lung cancer [107-109]. However, currently no specific therapeutic mAb to this molecule has been developed [110].

## **EPHA and EPHB**

Eph family receptor interacting proteins (Eph receptors) represent the largest family of RTKs [111]. Eph receptors are segregated into the two subclasses A (EphA1–A8, EphA10) and B (EphB1–B4, EphB6) according to their extracellular sequence homology, binding affinity and structure [112, 113]. The Eph ligands, ephrins, are divided into ephrin A (GPI anchored proteins) and ephrin B (transmembrane proteins) groups, based on their structure and sequence similarities [112, 114]. Cell to cell interactions are necessary in Eph activation through ephrin ligands binding because all Eph receptors and their ligands are membrane-bound proteins [111]. Eph and ephrins interactions lead to a bidirectional signaling in both the receptor and the ligand bearing cell [115].

Eph receptors are involved in a broad range of biological functions, including angiogenesis, cell attachment and cell segregation [116]. For instance, the Eph receptors have significant roles in the neuronal and vascular networks establishment during embryonic development and regulation of excitatory synapses function [117]. Eph receptors play functions in a variety of aspects of the tumor formation, including tumorigenesis, proliferation and metastasis [113, 118-120].

Regarding the expression profile, the Eph receptors/ephrins family has wide expression in adult tissues with organ-site-specific patterns. For instance EphA6, EphA8 and EphB1 transcripts have unique pattern in normal brain and testis [121]. The Eph receptors protein expression is downregulated in normal adult tissues [122, 123]. Eph and ephrin proteins are expressed at lower levels in adult comparing with embryonic tissues. This low-level expression may play a function in architecture of the kidney, the vasculature and the adult gut [124, 125]. On the contrary, Eph receptors and ephrins are relatively expressed at high level in the adult's brain [124, 126].

Eph receptors and ephrins are differentially expressed in a variety of human cancers [111]. For instance, EphA2 overexpression has been reported in breast, colorectal, prostate, lung, hepatocellular, and gastric and brain tumors [124, 125, 127]. Moreover, Eph receptors expression has been found in several leukemias such as AML [128].

Base on Eph receptors characteristics, there is a focus on Eph targeted cancer therapy. A fully human mAb (1C1) targeting EphA2 has been generated by Jackson et al [129]. 1C1 selectively bond to EphA2 and stimulated tyrosine phosphorylation, internalization and degradation of the

EphA2 receptor. Considering this internalize, an antibody-drug conjugate (ADC) has been prepared as a vehicle for delivery of cytotoxic drug to the cancer cells. EphA2 ADC selectively targeted and inhibited the growth of endometrial and ovarian malignant cells expressing EphA2 *in vitro* and *in vivo* [129-131]. After success in preclinical studies, this anti-EphA2 immunoconjugate has been studied in clinical trial phase I. Clinical trial data showed that the immunoconjugate did not have safety profile and the trial was terminated due to treatment-related bleeding and coagulation events [132]. Although findings were disappointing; however, targeting other Eph family members by mAbs is still ongoing. In this context, recently anti-EphA10 mAb has been introduced that significantly inhibited breast cancer cells proliferation and proposed EphA10 as a promising target for breast cancer therapy [123].

### **Met oncogene**

c-Met, also named Met is a proto-oncogene that encodes hepatocyte growth factor receptor (HGFR). The coding gene is located on chromosome 7q31. Met is the hepatocyte growth factor (HGF) high affinity receptor which is structurally composed of  $\alpha$  and  $\beta$  chains disulfide-linked heterodimer with 45 and 145 kDa mass, respectively. The  $\alpha$  chain and the N-terminal of the  $\beta$  chain form the extracellular region of Met [133]. Met is a master regulator RTK of cell survival, growth, differentiation, mobility, and cell division and have an essential role in normal development [134]. During embryogenesis, HGF and Met expression are vital for cell growth, development of hepatocytes, placental trophoblasts, and myoblasts [135, 136]. After birth, activation of the HGF-Met pathway appears to be involved in hepatic, renal, and other organ regeneration after injury, epithelial-mesenchymal transition (EMT) and wound healing [137].

Met and HGF interaction results in autophosphorylation of several key tyrosine residues, which recruit different downstream molecules, including phosphorylation of tyrosine residues 1234 and 1235 in the kinase domain that is crucial for kinases activation [137]. Activated Met phosphorylates and binds to growth factor receptor-bound protein 2 (Grb2) and Grb2-associated binding protein 1 (Gab1), acting as a scaffold protein and stimulating Met interactions with molecules involved in the PI3K/Akt, the signal transducer and activator of transcription factor (Stat), the mitogen-activated protein kinase (MAPK) and the NF- $\kappa$ B pathways [137].

Altered Met levels, and activity have been noted in several cancers, such as renal, colon, melanoma, and breast cancers [137]. Met is dysregulated in tumors by gene amplification,



translocation, mutation, and overexpression of Met and HGF proteins. Met dysregulation is correlated with an increased tendency for metastatic disease and poor overall survival. Blocking Met or HGF leads to cell growth inhibition and apoptosis induction and tumor cells necrosis. Therefore, targeting HGF and Met is an area of active research [138]. Several mAbs targeting Met have been developed with promising results in cancer therapy.

Onartuzumab (MetMAb) is an affinity-matured and humanized monovalent mAb that binds to the Sema domain of Met. It has been demonstrated that onartuzumab blocked HGF binding and prevented Met phosphorylation and signaling. Biochemical data and crystallography of onartuzumab antigen-binding site has shown that it binds to extracellular domain of Met. Therefore, onartuzumab specifically blocks binding of HGF  $\alpha$ -chain to Met [139]. These findings propose that  $\alpha$ -chain dimerization on Met leads to Met signaling activation [139]. In a KP4 pancreatic tumor cell mouse xenograft model onartuzumab suppressed tumor cell growth, decreased the Met phosphorylation and increased the mice survival [140]. Observing the effectiveness of onartuzumab in the animal model led to start clinical trials.

A dose-escalation phase I clinical trial checked the effects of onartuzumab and its combination with bevacizumab in 43 advanced solid tumors patients [140]. Onartuzumab half-life was 11 days, and no adverse effects were observed in combination with bevacizumab. Mild adverse reactions were hypo albuminuria, fatigue and peripheral edema [140]. In a phase II study, onartuzumab and erlotinib combination was evaluated in advanced NSCLC patients after initial therapy [141]. Patients randomly received 15 mg/kg onartuzumab every three weeks in combination with 150 mg erlotinib daily. Combination of onartuzumab with erlotinib showed a considerable improvement in progression free survival (1.5 to 2.9 months) and median overall survival from 3.8 to 12.6 months [141]. Interestingly, onartuzumab has been shown to be effective in EGFR-driven tumors acquire resistance to erlotinib [142]. In another phase II clinical trial, onartuzumab in combination with erlotinib was used in advanced and previously treated NSCLC patients [142]. This combination improved progression free survival and overall survival of patients with Met-overexpressed tumors [142]. Currently, a phase III clinical trial is testing the onartuzumab and erlotinib combination in NSCLC (<http://clinicaltrials.gov>).

In a recent study, p21-activated kinase 1 (PAK1) which is a central protein in pancreatic adenocarcinoma cells survival and downstream signaling pathways has been shown to be responsible for resistance to anti-Met mAbs and SMIs (such as onartuzumab and crizotinib,

respectively). In human, PAK1 expression has been shown to be highly associated with Met expression and is linked to tumor metastasis [143]. PAK1 inhibition blocked signaling to cytoskeletal effectors and tumor cell motility driven by HGF/Met. Combination of anti-Met mAbs with PAK1 inhibitors has been shown to overcome resistance mediated by PAK1. Inhibition of PAK1 attenuated *in vivo* tumor growth and metastasis in a model of pancreatic adenocarcinoma [143].

CE-355621 mAb is another anti-Met mAb that inhibited ligand-dependent activation of Met in A549 cell line. This mAb antagonized Met function by inhibiting the receptor activation and downregulating of downstream signaling pathways. In multiple xenograft tumor models, significant inhibition of Met activity and consequently tumor growth has been demonstrated. For instance, CE-355621 has been shown to inhibit up to 98% of U87MG glioblastoma cell and GTL-16 gastric cancer cells growth in xenograft models [144].

### **RECEPTEUR D'ORIGINE NANTAIS (Ron)**

The Ron (macrophage-stimulating protein receptor, MST1R) RTK belongs to Met family kinase. Ron stimulates proliferation, survival, cell migration, and invasion in the presence of ligand [145]. Ron also plays a role in the innate immunity by regulating macrophages migration and phagocytic activity [145]. It is expressed on epithelial cells and macrophages and controls the inflammatory response to various insults. All functional responses mediated by Ron and Met are initiated by their respective ligand [145]. Ron is produced as a single premature 180 kDa chain and then is cleaved into a 160 kDa  $\beta$ -chain and a 40 kDa  $\alpha$ -chain. The  $\alpha$ -chain is bound to the extracellular region of the  $\beta$ -chain by a disulfide bridge [146]. The Sema domain on the N-terminal of the Ron binds to HGF and macrophage stimulating protein (MSP) as ligands [147]. Subsequently, ligand binding induces Ron autophosphorylation that generates docking sites for downstream signaling molecules. Activated Ron interacts with the PI3K subunit PIK3R1, phospholipase C (PLC)  $\gamma$ 1 or the adapter Gab1. Then, these downstream effector molecules recruit and activate the Ras/Erk, PI3K/Akt axis or PLC $\gamma$ /PKC signaling pathways [147]. Ron signaling promotes epithelial cell proliferation, migration and survival at the wound site which stimulates wound healing process [147].

Ron expression profile in malignancies showed that it is overexpressed in several cancers, including pancreatic cancer cells and pancreatic cancer stem cells [148, 149], rectal cancer [150], and DLBCL [151]. Lack of both Met and Ron expression was shown to be associated with inferior overall survival in DLBCL patients [151]. Moreover, abnormal overexpression of Ron induced the generation of oncogenic variants such as Ron160 and Ron165. Currently, seven Ron oncogene variants have been recognized which are produced by alternative splicing or protein truncation [152-154]. Ron160 is expressed in primary invasive ductal, lobular and lymph node-involved breast cancer cells [154]. This variant is mostly observed in invasive ductal and lymph node-involved cases. Blocking of Ron160 signaling by PHA665752 inhibitor blocked Du4475 cell anchorage-independent growth and prompted apoptotic cell death. In addition, PHA665752 inhibitor prevented 3T3-Ron160 and Du4475 cell-mediated tumor cell growth implanted in mouse mammary fat pad [154].

Ron has been considered as a RTK for mAb targeting. In this milieu, anti-Ron mAbs inhibited 60% of subcutaneous and orthotopic tumor growth in nude mice model [155]. Likewise, anti-Ron antibody has also been demonstrated to be suitable for delivery of chemotherapeutic agents to the cancer cells. Anti-Ron mAb Zt/c9 doxorubicin immunoliposomes (Zt/c9-Dox-IL) has been shown to direct the toxin into the pancreatic CSCs (CD24<sup>+</sup>CD44<sup>+</sup>ESA<sup>+</sup>). Zt/c9-Dox-IL binding to Ron expressed on CSCs rapidly stimulated Ron internalization that led to doxorubicin uptake and subsequently reduced the viability of CSCs [149].

The inhibitory effects of anti-Ron mAbs have been examined in colon cancer. Zt/g4 mAb downregulated Ron expression in SW620 colon cancer cell line both in concentration and time dependent manner. Antibody-induced receptor internalization is the mechanism of Ron downregulation. Ron downregulation led to intracellular signaling inhibition by decreasing Erk1/2 and Akt phosphorylation. Moreover, Zt/g4 mAb changed SW620 cell morphology and affected its colony formation and increased its sensitivity in response to gemcitabine-induced cytotoxicity.

Anti-Ron mAb Zt/f2 targets an epitope located in a 49 amino acid residues of the  $\beta$ -chain extracellular domain of Ron [156]. This sequence is vital in Ron maturation and activation regulation. In mice models, Ron targeting by anti-Ron mAb Zt/f2 inhibited tumor growth of oncogenic Ron160 expressing NIH-3T3 cells. Moreover, this mAb attenuated cell-mediated tumor growth of colon cancer HT-29 in athymic nude mice [156].

In addition to solid tumors, Ron overexpression has been demonstrated in Burkitt's lymphoma (BL) cell lines and human lymphoma samples [157]. Anti-Ron mAb Zt/f2 also downregulated cell proliferation and colony formation and induced tumor cell apoptosis and cell cycle arrest in Raji BL cells [157].

IMC-Ron8, the first anti-Ron mAb entered into the clinical trial, downregulated the expression of Ron in pancreatic cancer cells and inhibited MSP-induced Ron activation, survivin expression and Akt and Erk activation. The co-treatment of pancreatic cells with panobinostat (PS) and anti-Ron mAb decreased Ron expression and Akt activation, and increased cleavage of PARP compared to both treatments alone [148].

Overall, reported data shows that anti-Ron mAbs reduced Ron expression, impaired signaling events and enhanced sensitivity towards cytotoxic drugs in cancer cells [158].

### **TROPOMYOSIN-RELATED KINASE (TrkA/NTrk1)**

The first member of tropomyosin-related kinase (Trk) family was primarily discovered as a new oncogene in colon carcinoma at 1986 [159]. TrkA/Ntrk1 is a normal cellular counterpart of the oncogenic Trk and is highly expressed in the developing nervous system [160]. TrkB/NTrk2 and TrkC/NTrk3 are other members of Trk family with highly sequence homology to TrkA [160]. TrkA is a functional and high-affinity nerve growth factor (NGF) receptor that mediates some of the signal transduction processes induced by this neurotrophic growth factor [161].

Trk receptors were introduced as important prognostic factors in neuroblastoma, in a way that TrkA and TrkB are correlated with favorable and unfavorable disease, respectively [162, 163]. Truncated forms of Trk receptors have been identified lacking the kinase domain. For example alternative TrkA splice variant has only functional extracellular domains and is expressed in human neuroblastoma [164]. This oncogenic NGF-unresponsive isoform antagonizes NGF/TrkA signaling that is accountable for arresting of neuroblastoma cell growth. During tumor progression, the truncated form provides a mechanism for switching anti-oncogenic signals of NGF/TrkA/Ras/MAPK to tumor-promoting signals of truncated-TrkA/IP3K/Akt/NF- $\kappa$ B [164].

Due to the importance of TrkA-NGF pathway in anti-oncogenic signals, targeting of TrkA with mAbs specific for docking site of TrkA would be an appropriate strategy to inhibit neuroblastoma progression and growth.

Tanezumab, a humanized NGF blocking mAb, has recently shown hopeful results in clinical trials for osteoarthritic pain because NGF/TrkA signaling is important for normal and pathological feeling of pain [165]. However, TrkA could not pass the criteria as an appropriate target for cancer therapy by mAbs. Moreover, there are reports that proposed TrkA for cancer targeting by other modalities like SMIs [166-169].

### **RECEPTOR TYROSINE KINASE-LIKE ORPHAN RECEPTOR 1 (Ror1)**

Ror family, consists of Ror1 and Ror2, is another member of the RTK families that is evolutionarily conserved [170]. Ror1 consists of three domains, immunoglobulin (Ig)-like, cysteine rich (CRD) and kringle (KNG), in the extracellular part [171]. Of the three extracellular regions, CRD region has been described to have the antigen binding site [172, 173]. The cytoplasmic part contains a TK domain with protein kinase activity, and further downstream serine, threonine- and proline-rich motifs. Ror1 is located on chromosome 1p31.3 and consists of 937 amino acids with an estimated molecular weight of 97-105 kDa. Human and mice Ror1 also have 97% amino acid identity [174, 175]. Ror1 and Ror2 have important roles in embryonic skeletal and cardiac as well as nervous system development in mouse [2, 176]. In this regard, Ror1 and Ror2 adjust hippocampal neurons neurite growth and branching pattern that are important in brain development [177]. In adults, Ror1 has been indicated to be expressed at mRNA level in several tissues, including colon, adipocytes, kidney, liver, lung, Ovary, lymph nodes, prostate, and testis [178]. However, Ror1 protein has not been detected in adult normal tissues [178].

Interestingly, Ror1 overexpression has been found in several malignancies, including CLL [179], ALL [180], AML [6, 181], HCL [6], MCL [6], melanoma [7], breast, pancreatic, lung [182], and ovarian cancers [14]. Recently it is shown that Ror1 can contribute in CLL leukemogenesis by interacting with T-cell leukemia 1 (TCL1) and causing elevated Akt activation and leukemia cell proliferation [183].

Overexpression of Ror1 on cancer cells and lacking expression in normal tissues introduced Ror1 as an appropriate target for cancer therapy. During the recent years that Ror1 has been recognized as an important tumor antigen, a few mAbs have been produced for therapeutic approaches [4, 178, 184]. Up to date, only an anti-Ror1 mAb, cirmtuzumab (humanized) (UC-961) has passed the preclinical stages. This mAb has been developed from the D10 anti-Ror1 mAb. This mAb showed a high specificity and affinity ( $KD=4nM$ ) for Ror1. Intravenous injection of cirmtuzumab followed by infusion of human  $Ror1^{+}CD5^{+}B220^{low}$  murine leukemia cells from Ror1xTCL1 transgenic mice, as well as of human  $Ror1^{+}$  CLL cells into the peritoneal cavity of Rag-2-/-/γc-/- immunodeficient mice, induced clearance of leukemic cells in the spleen and peritoneal cavity. Cirmtuzumab had not only a direct killing effect on tumor cells but was also internalized by malignant B cells. An ADC using this mAb showed promising results with enhanced cytotoxic activity against Ror1 expressing cells. Cirmtuzumab-ADC cleared Ror1 expressing CLL cells *in vivo* in xenografted mice and *in vitro* using adenocarcinoma cell lines of the breast and pancreas [185].

Overall, among 30 known oncogenic RTKs, several have not been well-studied for targeted therapy or several obstacles are in front of researchers to target them by mAbs. Proper and suitable RTKs for mAb targeting such as Musk, Mer, Eph receptors, NTrk, Ror2, Ros, Ryk, Tek, Tie, and Tyro3 are overexpressed in several tumors. A few mAb against these RTKs are in early stages of preclinical studies. More studies are necessary to investigate these RTKs for targeted therapies.

## **CONCLUSION**

Oncogenic RTKs are one of the appropriate classes of surface molecules for mAb targeted therapy. However, in spite of large efforts in targeting RTKs by specific mAbs, the evidences show the unsuccessfulness of this field and only a few antibodies have been entered clinical trials or have been approved by FDA for clinical application. Several obstacles, including differential post-translational modification of RTKs within a tumor population in different cancers as well as within different individual patients, paradigm of CSCs and the lack of enough information about their characteristics are in front of researchers. A better understanding of molecular, genetic and epigenetic factors involved in the pathogenesis of cancer, especially in a patient-based manner might be a suitable way to cure cancer in each patient. Tumor heterogeneity and the presence of several types of cancer cells within a special tumor suggest that the future of cancer targeted therapy may apply to target a particular cellular or molecular pathway within a specific cell type (cell-based therapy). On the other hand, current evidences show a better response in combinational therapy that targets different critical and pivotal molecules involved in cancer growth and progression. Overall, cancer therapy by mAbs may need to define new critical and vital targets on cancer cells and new approaches are warranted, and current methods might be necessary to be revised.

### **Competing interest**

The authors have no relevant affiliation or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript.

### **Acknowledgements**

This study was supported by grants from Felix Mindus foundation, the CLL Global Research Foundation, Stockholm, Sweden, and Monoclonal Antibody Research Center, Avicenna Research Institute, ACECR, Tehran, Iran.



## REFERENCES

1. Hubbard SR, Miller WT. Receptor tyrosine kinases: mechanisms of activation and signaling. *Curr Opin Cell Biol* 2007; 19: 117-123.
2. Nomi M, Oishi I, Kani S, et al. Loss of mRor1 enhances the heart and skeletal abnormalities in mRor2-deficient mice: redundant and pleiotropic functions of mRor1 and mRor2 receptor tyrosine kinases. *Mol Cell Biol* 2001; 21: 8329-8335.
3. Daneshmanesh AH, Mikaelsson E, Jeddi-Tehrani M, et al. Ror1, a cell surface receptor tyrosine kinase is expressed in chronic lymphocytic leukemia and may serve as a putative target for therapy. *Int J Cancer* 2008; 123: 1190-1195.
4. Daneshmanesh AH, Hojjat-Farsangi M, Khan AS, et al. Monoclonal antibodies against ROR1 induce apoptosis of chronic lymphocytic leukemia (CLL) cells. *Leukemia* 2012; 26: 1348-1355.
5. Hojjat-Farsangi M, Khan AS, Daneshmanesh AH, et al. The Tyrosine Kinase Receptor ROR1 Is Constitutively Phosphorylated in Chronic Lymphocytic Leukemia (CLL) Cells. *PLoS One* 2013; 8: e78339.
6. Daneshmanesh AH, Porwit A, Hojjat-Farsangi M, et al. Orphan receptor tyrosine kinases ROR1 and ROR2 in hematological malignancies. *Leuk Lymphoma* 2013; 54: 843-850.
7. Hojjat-Farsangi M, Ghaemimanesh F, Daneshmanesh AH, et al. Inhibition of the receptor tyrosine kinase ROR1 by anti-ROR1 monoclonal antibodies and siRNA induced apoptosis of melanoma cells. *PLoS One* 2013; 8: e61167.
8. O'Connell MP, Marchbank K, Webster MR, et al. Hypoxia induces phenotypic plasticity and therapy resistance in melanoma via the tyrosine kinase receptors ROR1 and ROR2. *Cancer Discov* 2013.
9. Yamaguchi T, Yanagisawa K, Sugiyama R, et al. NKX2-1/TTF1/TTF-1-Induced ROR1 is required to sustain EGFR survival signaling in lung adenocarcinoma. *Cancer Cell* 2012; 21: 348-361.
10. Xu AM, Huang PH. Receptor tyrosine kinase coactivation networks in cancer. *Cancer Res* 2010; 70: 3857-3860.
11. Robertson SC, Tynan J, Donoghue DJ. RTK mutations and human syndromes: when good receptors turn bad. *Trends Genet* 2000; 16: 368.
12. Porter AC, Vaillancourt RR. Tyrosine kinase receptor-activated signal transduction pathways which lead to oncogenesis. *Oncogene* 1998; 17: 1343-1352.
13. Blume-Jensen P, Hunter T. Oncogenic kinase signalling. *Nature* 2001; 411: 355-365.
14. Hojjat-Farsangi M. Small-molecule inhibitors of the receptor tyrosine kinases: promising tools for targeted cancer therapies. *Int J Mol Sci* 2014; 15: 13768-13801.
15. Connolly DT, Heuvelman DM, Nelson R, et al. Tumor vascular permeability factor stimulates endothelial cell growth and angiogenesis. *J Clin Invest* 1989; 84: 1470-1478.
16. Bauer K, Rancea M, Roloff V, et al. Rituximab, ofatumumab and other monoclonal anti-CD20 antibodies for chronic lymphocytic leukaemia. *Cochrane Database Syst Rev* 2012; 11: CD008079.
17. Schaal AD. Alemtuzumab (Campath 1-H). *Clin J Oncol Nurs* 2005; 9: 630-632.
18. Honeychurch J, Alduaij W, Azizyan M, et al. Antibody-induced nonapoptotic cell death in human lymphoma and leukemia cells is mediated through a novel reactive oxygen species-dependent pathway. *Blood* 2012; 119: 3523-3533.
19. Hojjat-Farsangi M, Moshfegh A, Daneshmanesh AH, et al. The receptor tyrosine kinase ROR1 - An oncofetal antigen for targeted cancer therapy. *Semin Cancer Biol* 2014.
20. Gschwind A, Fischer OM, Ullrich A. The discovery of receptor tyrosine kinases: targets for cancer therapy. *Nat Rev Cancer* 2004; 4: 361-370.
21. Robinson DR, Wu YM, Lin SF. The protein tyrosine kinase family of the human genome. *Oncogene* 2000; 19: 5548-5557.

22. Ullrich A, Schlessinger J. Signal transduction by receptors with tyrosine kinase activity. *Cell* 1990; 61: 203-212.
23. Hubbard SR, Till JH. Protein tyrosine kinase structure and function. *Annu Rev Biochem* 2000; 69: 373-398.
24. Watson ME. Compilation of published signal sequences. *Nucleic Acids Res* 1984; 12: 5145-5164.
25. Ostman A, Bohmer FD. Regulation of receptor tyrosine kinase signaling by protein tyrosine phosphatases. *Trends Cell Biol* 2001; 11: 258-266.
26. Hubbard SR. Structural analysis of receptor tyrosine kinases. *Prog Biophys Mol Biol* 1999; 71: 343-358.
27. Hunter T, Cooper JA. Protein-tyrosine kinases. *Annu Rev Biochem* 1985; 54: 897-930.
28. Bennisroune A, Gardin A, Aunis D, et al. Tyrosine kinase receptors as attractive targets of cancer therapy. *Crit Rev Oncol Hematol* 2004; 50: 23-38.
29. Haglund K, Rusten TE, Stenmark H. Aberrant receptor signaling and trafficking as mechanisms in oncogenesis. *Crit Rev Oncog* 2007; 13: 39-74.
30. Abella JV, Park M. Breakdown of endocytosis in the oncogenic activation of receptor tyrosine kinases. *Am J Physiol Endocrinol Metab* 2009; 296: E973-984.
31. Garrett CR, Eng C. Cetuximab in the treatment of patients with colorectal cancer. *Expert Opin Biol Ther* 2011; 11: 937-949.
32. Specenier P, Vermorken JB. Cetuximab in the treatment of squamous cell carcinoma of the head and neck. *Expert Rev Anticancer Ther* 2011; 11: 511-524.
33. Saif MW, Kim R. Incidence and management of cutaneous toxicities associated with cetuximab. *Expert Opin Drug Saf* 2007; 6: 175-182.
34. Luedke E, Jaime-Ramirez AC, Bhave N, et al. Monoclonal antibody therapy of pancreatic cancer with cetuximab: potential for immune modulation. *J Immunother* 2012; 35: 367-373.
35. Kim R. Cetuximab and panitumumab: are they interchangeable? *Lancet Oncol* 2009; 10: 1140-1141.
36. Zaorsky NG, Sun Y, Wang Z, et al. Identification of a KRAS mutation in a patient with non-small cell lung cancer treated with chemoradiotherapy and panitumumab. *Cancer Biol Ther* 2013; 14.
37. Weiner LM, Surana R, Wang S. Monoclonal antibodies: versatile platforms for cancer immunotherapy. *Nat Rev Immunol* 2010; 10: 317-327.
38. Cheng AL, Li J, Vaid AK, et al. Adaptation of international guidelines for metastatic colorectal cancer: an asian consensus. *Clin Colorectal Cancer* 2014; 13: 145-155.
39. Cho HS, Mason K, Ramyar KX, et al. Structure of the extracellular region of HER2 alone and in complex with the Herceptin Fab. *Nature* 2003; 421: 756-760.
40. Luis M, Tavares A, Carvalho LS, et al. Personalizing therapies for gastric cancer: Molecular mechanisms and novel targeted therapies. *World J Gastroenterol* 2013; 19: 6383-6397.
41. Perez EA, Spano JP. Current and emerging targeted therapies for metastatic breast cancer. *Cancer* 2012; 118: 3014-3025.
42. de Mello RA, Marques AM, Araujo A. HER2 therapies and gastric cancer: A step forward. *World J Gastroenterol* 2013; 19: 6165-6169.
43. Graziano C. HER-2 breast assay, linked to Herceptin, wins FDA's okay. *CAP Today* 1998; 12: 1, 14-16.
44. Gradishar WJ. Emerging approaches for treating HER2-positive metastatic breast cancer beyond trastuzumab. *Ann Oncol* 2013; 24: 2492-2500.
45. Slamon DJ, Leyland-Jones B, Shak S, et al. Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. *The New England journal of medicine* 2001; 344: 783-792.

46. Norman G, Rice S, Spackman E, et al. Trastuzumab for the treatment of HER2-positive metastatic adenocarcinoma of the stomach or gastro-oesophageal junction. *Health Technol Assess* 2011; 15 Suppl 1: 33-42.
47. Chen JS, Lan K, Hung MC. Strategies to target HER2/neu overexpression for cancer therapy. *Drug Resist Updat* 2003; 6: 129-136.
48. Hudis CA. Trastuzumab--mechanism of action and use in clinical practice. *N Engl J Med* 2007; 357: 39-51.
49. Awada A, Bozovic-Spasojevic I, Chow L. New therapies in HER2-positive breast cancer: a major step towards a cure of the disease? *Cancer treatment reviews* 2012; 38: 494-504.
50. Nahta R, Esteva FJ. Trastuzumab: triumphs and tribulations. *Oncogene* 2007; 26: 3637-3643.
51. Huang Y, Fu P, Fan W. Novel targeted therapies to overcome trastuzumab resistance in HER2-overexpressing metastatic breast cancer. *Current drug targets* 2013; 14: 889-898.
52. Capelan M, Pugliano L, De Azambuja E, et al. Pertuzumab: new hope for patients with HER2-positive breast cancer. *Annals of oncology : official journal of the European Society for Medical Oncology / ESMO* 2013; 24: 273-282.
53. Miles D, Baselga J, Amadori D, et al. Treatment of older patients with HER2-positive metastatic breast cancer with pertuzumab, trastuzumab, and docetaxel: subgroup analyses from a randomized, double-blind, placebo-controlled phase III trial (CLEOPATRA). *Breast Cancer Res Treat* 2013; 142: 89-99.
54. Roche. H-L. A Study of Pertuzumab in Combination With Trastuzumab and Chemotherapy in Patients With HER2-Positive. *Advanced Gastric Cancer* 2011. Available from: URL: <http://clinicaltrials.gov/ct2012/show/NCT01461057>.
55. Jelovac D, Emens LA. HER2-directed therapy for metastatic breast cancer. *Oncology (Williston Park)* 2013; 27: 166-175.
56. Ferrara N, Gerber HP, LeCouter J. The biology of VEGF and its receptors. *Nature medicine* 2003; 9: 669-676.
57. Sullivan LA, Brekken RA. The VEGF family in cancer and antibody-based strategies for their inhibition. *MAbs* 2010; 2: 165-175.
58. Miller K, Wang M, Gralow J, et al. Paclitaxel plus bevacizumab versus paclitaxel alone for metastatic breast cancer. *N Engl J Med* 2007; 357: 2666-2676.
59. Hurwitz H, Fehrenbacher L, Novotny W, et al. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *N Engl J Med* 2004; 350: 2335-2342.
60. Sandler A, Gray R, Perry MC, et al. Paclitaxel-carboplatin alone or with bevacizumab for non-small-cell lung cancer. *N Engl J Med* 2006; 355: 2542-2550.
61. Ellis LM, Hicklin DJ. VEGF-targeted therapy: mechanisms of anti-tumour activity. *Nat Rev Cancer* 2008; 8: 579-591.
62. Melichar B, Prochazkova-Studentova H, Vitaskova D. Bevacizumab in combination with IFN-alpha in metastatic renal cell carcinoma: the AVOREN trial. *Expert Rev Anticancer Ther* 2012; 12: 1253-1261.
63. Rini BI, Halabi S, Rosenberg JE, et al. Bevacizumab plus interferon alfa compared with interferon alfa monotherapy in patients with metastatic renal cell carcinoma: CALGB 90206. *J Clin Oncol* 2008; 26: 5422-5428.
64. Cartwright TH. Adverse events associated with antiangiogenic agents in combination with cytotoxic chemotherapy in metastatic colorectal cancer and their management. *Clinical colorectal cancer* 2013; 12: 86-94.
65. Sandomenico C, Costanzo R, Carillio G, et al. Bevacizumab in non small cell lung cancer: development, current status and issues. *Current medicinal chemistry* 2012; 19: 961-971.

66. Tappenden P, Jones R, Paisley S, et al. Systematic review and economic evaluation of bevacizumab and cetuximab for the treatment of metastatic colorectal cancer. *Health Technol Assess* 2007; 11: 1-128, iii-iv.
67. Hirschi KK, Rohovsky SA, D'Amore PA. PDGF, TGF-beta, and heterotypic cell-cell interactions mediate endothelial cell-induced recruitment of 10T1/2 cells and their differentiation to a smooth muscle fate. *The Journal of cell biology* 1998; 141: 805-814.
68. Keehn CA, Saeed S, Bickle K, et al. Expression of insulin-like growth factor-I receptor in primary cutaneous carcinomas. *Journal of cutaneous pathology* 2004; 31: 368-372.
69. LeRoith D, Roberts CT, Jr. The insulin-like growth factor system and cancer. *Cancer Lett* 2003; 195: 127-137.
70. Baserga R, Hongo A, Rubini M, et al. The IGF-I receptor in cell growth, transformation and apoptosis. *Biochimica et biophysica acta* 1997; 1332: F105-126.
71. Warshamana-Greene GS, Litz J, Buchdunger E, et al. The insulin-like growth factor-I receptor kinase inhibitor, NVP-ADW742, sensitizes small cell lung cancer cell lines to the effects of chemotherapy. *Clin Cancer Res* 2005; 11: 1563-1571.
72. Jones HE, Goddard L, Gee JM, et al. Insulin-like growth factor-I receptor signalling and acquired resistance to gefitinib (ZD1839; Iressa) in human breast and prostate cancer cells. *Endocr Relat Cancer* 2004; 11: 793-814.
73. Krueckl SL, Sikes RA, Edlund NM, et al. Increased insulin-like growth factor I receptor expression and signaling are components of androgen-independent progression in a lineage-derived prostate cancer progression model. *Cancer Res* 2004; 64: 8620-8629.
74. Tolcher AW, Sarantopoulos J, Patnaik A, et al. Phase I, pharmacokinetic, and pharmacodynamic study of AMG 479, a fully human monoclonal antibody to insulin-like growth factor receptor 1. *J Clin Oncol* 2009; 27: 5800-5807.
75. Tap WD, Demetri G, Barnette P, et al. Phase II study of ganitumab, a fully human anti-type-1 insulin-like growth factor receptor antibody, in patients with metastatic Ewing family tumors or desmoplastic small round cell tumors. *J Clin Oncol* 2012; 30: 1849-1856.
76. Kindler HL, Richards DA, Garbo LE, et al. A randomized, placebo-controlled phase 2 study of ganitumab (AMG 479) or conatumumab (AMG 655) in combination with gemcitabine in patients with metastatic pancreatic cancer. *Ann Oncol* 2012; 23: 2834-2842.
77. McKian KP, Haluska P. Cixutumumab. *Expert Opin Investig Drugs* 2009; 18: 1025-1033.
78. Bagatell R, Herzog CE, Trippett TM, et al. Pharmacokinetically guided phase 1 trial of the IGF-1 receptor antagonist RG1507 in children with recurrent or refractory solid tumors. *Clin Cancer Res* 2011; 17: 611-619.
79. Doi T, Muro K, Yoshino T, et al. Phase 1 pharmacokinetic study of MK-0646 (dalotuzumab), an anti-insulin-like growth factor-1 receptor monoclonal antibody, in combination with cetuximab and irinotecan in Japanese patients with advanced colorectal cancer. *Cancer Chemother Pharmacol* 2013; 72: 643-652.
80. Becerra CR, Salazar R, Garcia-Carbonero R, et al. Figitumumab in patients with refractory metastatic colorectal cancer previously treated with standard therapies: a nonrandomized, open-label, phase II trial. *Cancer Chemother Pharmacol* 2014; 73: 695-702.
81. Lin EH, Lenz HJ, Saleh MN, et al. A randomized, phase II study of the anti-insulin-like growth factor receptor type 1 (IGF-1R) monoclonal antibody robatumumab (SCH 717454) in patients with advanced colorectal cancer. *Cancer Med* 2014; 3: 988-997.
82. Macaulay VM, Middleton MR, Protheroe AS, et al. Phase I study of humanized monoclonal antibody AVE1642 directed against the type 1 insulin-like growth factor receptor (IGF-1R), administered in combination with anticancer therapies to patients with advanced solid tumors. *Ann Oncol* 2013; 24: 784-791.

83. Linger RM, Keating AK, Earp HS, et al. TAM receptor tyrosine kinases: biologic functions, signaling, and potential therapeutic targeting in human cancer. *Adv Cancer Res* 2008; 100: 35-83.
84. Nguyen KQ, Tsou WI, Kotenko S, et al. TAM receptors in apoptotic cell clearance, autoimmunity, and cancer. *Autoimmunity* 2013; 46: 294-297.
85. Hojjat-Farsangi M, Daneshmanesh A, Khan A, et al. Targeting the tyrosine kinase receptors ROR1 and AXL (RTKs) in Burkitt's lymphoma. 2014, 19th European Hematology association (EHA), June 12-15; Abstract No. P424.
86. Hojjat-Farsangi M, Daneshmanesh A, Salam Khan A, et al. Targeting the tyrosine kinase receptors ROR1 and AXL (RTKs) in Burkitt's lymphoma. . 2014, 19th Congress of European Hematology Association (EHA) – June 12-15, Milan, Italy, Abstract No: 3589.
87. Reichl P, Dengler M, van Zijl F, et al. Signaling of Axl via 14-3-3zeta activates autocrine transforming growth factor-beta signaling in hepatocellular carcinoma. *Hepatology* 2014.
88. Waizenegger JS, Ben-Batalla I, Weinhold N, et al. Role of Growth arrest-specific gene 6-Mer axis in multiple myeloma. *Leukemia* 2014.
89. D'Alfonso TM, Hannah J, Chen Z, et al. Axl receptor tyrosine kinase expression in breast cancer. *J Clin Pathol* 2014; 67: 690-696.
90. Tsou AP, Wu KM, Tsen TY, et al. Parallel hybridization analysis of multiple protein kinase genes: identification of gene expression patterns characteristic of human hepatocellular carcinoma. *Genomics* 1998; 50: 331-340.
91. Linger RM, Keating AK, Earp HS, et al. Taking aim at Mer and Axl receptor tyrosine kinases as novel therapeutic targets in solid tumors. *Expert Opin Ther Targets* 2010; 14: 1073-1090.
92. Leconet W, Larbouret C, Chardes T, et al. Preclinical validation of AXL receptor as a target for antibody-based pancreatic cancer immunotherapy. *Oncogene* 2013.
93. Vogel W, Gish GD, Alves F, et al. The discoidin domain receptor tyrosine kinases are activated by collagen. *Molecular cell* 1997; 1: 13-23.
94. Vogel W. Discoidin domain receptors: structural relations and functional implications. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology* 1999; 13 Suppl: S77-82.
95. Shrivastava A, Radziejewski C, Campbell E, et al. An orphan receptor tyrosine kinase family whose members serve as nonintegrin collagen receptors. *Molecular cell* 1997; 1: 25-34.
96. Edelhoff S, Sweetser DA, Distèche CM. Mapping of the NEP receptor tyrosine kinase gene to human chromosome 6p21.3 and mouse chromosome 17C. *Genomics* 1995; 25: 309-311.
97. Borza CM, Pozzi A. Discoidin domain receptors in disease. *Matrix biology : journal of the International Society for Matrix Biology* 2014; 34: 185-192.
98. Leitinger B. Discoidin domain receptor functions in physiological and pathological conditions. *International review of cell and molecular biology* 2014; 310: 39-87.
99. Barker KT, Martindale JE, Mitchell PJ, et al. Expression patterns of the novel receptor-like tyrosine kinase, DDR, in human breast tumours. *Oncogene* 1995; 10: 569-575.
100. Alves F, Vogel W, Mossie K, et al. Distinct structural characteristics of discoidin I subfamily receptor tyrosine kinases and complementary expression in human cancer. *Oncogene* 1995; 10: 609-618.
101. Ford CE, Lau SK, Zhu CQ, et al. Expression and mutation analysis of the discoidin domain receptors 1 and 2 in non-small cell lung carcinoma. *British journal of cancer* 2007; 96: 808-814.
102. Weiner HL, Huang H, Zagzag D, et al. Consistent and selective expression of the discoidin domain receptor-1 tyrosine kinase in human brain tumors. *Neurosurgery* 2000; 47: 1400-1409.
103. Heinzelmann-Schwarz VA, Gardiner-Garden M, Henshall SM, et al. Overexpression of the cell adhesion molecules DDR1, Claudin 3, and Ep-CAM in metaplastic ovarian epithelium and ovarian

- cancer. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2004; 10: 4427-4436.
104. Quan J, Yahata T, Adachi S, et al. Identification of receptor tyrosine kinase, discoidin domain receptor 1 (DDR1), as a potential biomarker for serous ovarian cancer. *International journal of molecular sciences* 2011; 12: 971-982.
  105. Shimada K, Nakamura M, Ishida E, et al. Prostate cancer antigen-1 contributes to cell survival and invasion through discoidin receptor 1 in human prostate cancer. *Cancer science* 2008; 99: 39-45.
  106. Valiathan RR, Marco M, Leitinger B, et al. Discoidin domain receptor tyrosine kinases: new players in cancer progression. *Cancer metastasis reviews* 2012; 31: 295-321.
  107. Canning P, Tan L, Chu K, et al. Structural mechanisms determining inhibition of the collagen receptor DDR1 by selective and multi-targeted type II kinase inhibitors. *Journal of molecular biology* 2014; 426: 2457-2470.
  108. Day E, Waters B, Spiegel K, et al. Inhibition of collagen-induced discoidin domain receptor 1 and 2 activation by imatinib, nilotinib and dasatinib. *European journal of pharmacology* 2008; 599: 44-53.
  109. Kim HG, Tan L, Weisberg EL, et al. Discovery of a potent and selective DDR1 receptor tyrosine kinase inhibitor. *ACS chemical biology* 2013; 8: 2145-2150.
  110. Fu HL, Sohail A, Valiathan RR, et al. Shedding of discoidin domain receptor 1 by membrane-type matrix metalloproteinases. *The Journal of biological chemistry* 2013; 288: 12114-12129.
  111. Hafner C, Becker B, Landthaler M, et al. Expression profile of Eph receptors and ephrin ligands in human skin and downregulation of EphA1 in nonmelanoma skin cancer. *Modern pathology : an official journal of the United States and Canadian Academy of Pathology, Inc* 2006; 19: 1369-1377.
  112. Committee EN. Unified nomenclature for Eph family receptors and their ligands, the ephrins. Eph Nomenclature Committee. *Cell* 1997; 90: 403-404.
  113. Nagano K, Kanasaki S, Yamashita T, et al. Expression of Eph receptor A10 is correlated with lymph node metastasis and stage progression in breast cancer patients. *Cancer medicine* 2013; 2: 972-977.
  114. Jensen PL. Eph receptors and ephrins. *Stem Cells* 2000; 18: 63-64.
  115. Bruckner K, Klein R. Signaling by Eph receptors and their ephrin ligands. *Current opinion in neurobiology* 1998; 8: 375-382.
  116. Surawska H, Ma PC, Salgia R. The role of ephrins and Eph receptors in cancer. *Cytokine & growth factor reviews* 2004; 15: 419-433.
  117. Yamaguchi Y, Pasquale EB. Eph receptors in the adult brain. *Current opinion in neurobiology* 2004; 14: 288-296.
  118. Brantley-Sieders DM, Zhuang G, Hicks D, et al. The receptor tyrosine kinase EphA2 promotes mammary adenocarcinoma tumorigenesis and metastatic progression in mice by amplifying ErbB2 signaling. *The Journal of clinical investigation* 2008; 118: 64-78.
  119. Mohammed KA, Wang X, Goldberg EP, et al. Silencing receptor EphA2 induces apoptosis and attenuates tumor growth in malignant mesothelioma. *American journal of cancer research* 2011; 1: 419-431.
  120. Sawamiphak S, Seidel S, Essmann CL, et al. Ephrin-B2 regulates VEGFR2 function in developmental and tumour angiogenesis. *Nature* 2010; 465: 487-491.
  121. Hafner C, Schmitz G, Meyer S, et al. Differential gene expression of Eph receptors and ephrins in benign human tissues and cancers. *Clinical chemistry* 2004; 50: 490-499.
  122. Day BW, Stringer BW, Boyd AW. Eph receptors as therapeutic targets in glioblastoma. *British journal of cancer* 2014.

123. Nagano K, Maeda Y, Kanasaki SI, et al. Ephrin receptor A10 is a promising drug target potentially useful for breast cancers including triple negative breast cancers. *Journal of controlled release : official journal of the Controlled Release Society* 2014.
124. Herath NI, Boyd AW. The role of Eph receptors and ephrin ligands in colorectal cancer. *International journal of cancer Journal international du cancer* 2010; 126: 2003-2011.
125. Xi HQ, Wu XS, Wei B, et al. Eph receptors and ephrins as targets for cancer therapy. *Journal of cellular and molecular medicine* 2012; 16: 2894-2909.
126. Gerlai R. Eph receptors and neural plasticity. *Nature reviews Neuroscience* 2001; 2: 205-209.
127. Wykosky J, Debinski W. The EphA2 receptor and ephrinA1 ligand in solid tumors: function and therapeutic targeting. *Molecular cancer research : MCR* 2008; 6: 1795-1806.
128. Wrobel T, Pogrzeba J, Stefanko E, et al. Expression of Eph A4, Eph B2 and Eph B4 Receptors in AML. *Pathology oncology research : POR* 2014.
129. Jackson D, Gooya J, Mao S, et al. A human antibody-drug conjugate targeting EphA2 inhibits tumor growth in vivo. *Cancer Res* 2008; 68: 9367-9374.
130. Lee JW, Han HD, Shahzad MM, et al. EphA2 immunoconjugate as molecularly targeted chemotherapy for ovarian carcinoma. *Journal of the National Cancer Institute* 2009; 101: 1193-1205.
131. Lee JW, Stone RL, Lee SJ, et al. EphA2 targeted chemotherapy using an antibody drug conjugate in endometrial carcinoma. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2010; 16: 2562-2570.
132. Annunziata CM, Kohn EC, LoRusso P, et al. Phase 1, open-label study of MEDI-547 in patients with relapsed or refractory solid tumors. *Investigational new drugs* 2013; 31: 77-84.
133. Naka T, Boltze C, Samii A, et al. Expression of c-MET, low-molecular-weight cytokeratin, matrix metalloproteinases-1 and -2 in spinal chordoma. *Histopathology* 2009; 54: 607-613.
134. Puri N, Ahmed S, Janamanchi V, et al. c-Met is a potentially new therapeutic target for treatment of human melanoma. *Clin Cancer Res* 2007; 13: 2246-2253.
135. Uehara Y, Minowa O, Mori C, et al. Placental defect and embryonic lethality in mice lacking hepatocyte growth factor/scatter factor. *Nature* 1995; 373: 702-705.
136. Schmidt C, Bladt F, Goedecke S, et al. Scatter factor/hepatocyte growth factor is essential for liver development. *Nature* 1995; 373: 699-702.
137. Parikh RA, Wang P, Beumer JH, et al. The potential roles of hepatocyte growth factor (HGF)-MET pathway inhibitors in cancer treatment. *Onco Targets Ther* 2014; 7: 969-983.
138. Kentsis A, Reed C, Rice KL, et al. Autocrine activation of the MET receptor tyrosine kinase in acute myeloid leukemia. *Nat Med* 2012; 18: 1118-1122.
139. Merchant M, Ma X, Maun HR, et al. Monovalent antibody design and mechanism of action of onartuzumab, a MET antagonist with anti-tumor activity as a therapeutic agent. *Proc Natl Acad Sci U S A* 2013; 110: E2987-2996.
140. Jin H, Yang R, Zheng Z, et al. MetMAb, the one-armed 5D5 anti-c-Met antibody, inhibits orthotopic pancreatic tumor growth and improves survival. *Cancer Res* 2008; 68: 4360-4368.
141. Spigel DR, Ervin TJ, Ramlau RA, et al. Randomized phase II trial of Onartuzumab in combination with erlotinib in patients with advanced non-small-cell lung cancer. *J Clin Oncol* 2013; 31: 4105-4114.
142. Belalcazar A, Azana D, Perez CA, et al. Targeting the Met pathway in lung cancer. *Expert review of anticancer therapy* 2012; 12: 519-528.
143. Zhou W, Jubb AM, Lyle K, et al. PAK1 mediates pancreatic cancer cell migration and resistance to MET inhibition. *J Pathol* 2014.
144. Michaud NR, Jani JP, Hillerman S, et al. Biochemical and pharmacological characterization of human c-Met neutralizing monoclonal antibody CE-355621. *MAbs* 2012; 4: 710-723.

145. Dussault I, Bellon SF. From concept to reality: the long road to c-Met and RON receptor tyrosine kinase inhibitors for the treatment of cancer. *Anticancer Agents Med Chem* 2009; 9: 221-229.
146. Correll PH, Paulson RF, Wei X. Molecular regulation of receptor tyrosine kinases in hematopoietic malignancies. *Gene* 2006; 374: 26-38.
147. Leonard EJ, Danilkovitch A. Macrophage stimulating protein. *Adv Cancer Res* 2000; 77: 139-167.
148. Zou Y, Howell GM, Humphrey LE, et al. Ron knockdown and Ron monoclonal antibody IMC-RON8 sensitize pancreatic cancer to histone deacetylase inhibitors (HDACi). *PLoS One* 2013; 8: e69992.
149. Padhye SS, Guin S, Yao HP, et al. Sustained expression of the RON receptor tyrosine kinase by pancreatic cancer stem cells as a potential targeting moiety for antibody-directed chemotherapeutics. *Mol Pharm* 2011; 8: 2310-2319.
150. Saigusa S, Toiyama Y, Tanaka K, et al. Inhibition of HGF/cMET expression prevents distant recurrence of rectal cancer after preoperative chemoradiotherapy. *Int J Oncol* 2012; 40: 583-591.
151. Koh YW, Hwang HS, Jung SJ, et al. Receptor tyrosine kinases MET and RON as prognostic factors in diffuse large B-cell lymphoma patients receiving R-CHOP. *Cancer Sci* 2013; 104: 1245-1251.
152. Zhou YQ, He C, Chen YQ, et al. Altered expression of the RON receptor tyrosine kinase in primary human colorectal adenocarcinomas: generation of different splicing RON variants and their oncogenic potential. *Oncogene* 2003; 22: 186-197.
153. Lu Y, Yao HP, Wang MH. Multiple variants of the RON receptor tyrosine kinase: biochemical properties, tumorigenic activities, and potential drug targets. *Cancer Lett* 2007; 257: 157-164.
154. Yao HP, Zhuang CM, Zhou YQ, et al. Oncogenic variant RON160 expression in breast cancer and its potential as a therapeutic target by small molecule tyrosine kinase inhibitor. *Curr Cancer Drug Targets* 2013; 13: 686-697.
155. Camp ER, Yang A, Gray MJ, et al. Tyrosine kinase receptor RON in human pancreatic cancer: expression, function, and validation as a target. *Cancer* 2007; 109: 1030-1039.
156. Yao HP, Zhou YQ, Ma Q, et al. The monoclonal antibody Zt/f2 targeting RON receptor tyrosine kinase as potential therapeutics against tumor growth-mediated by colon cancer cells. *Mol Cancer* 2011; 10: 82.
157. Tong X, Zhang X, Fan J, et al. The RON receptor tyrosine kinase is a potential therapeutic target in Burkitt lymphoma. *Cancer Biol Ther* 2013; 14: 370-377.
158. Li Z, Yao H, Guin S, et al. Monoclonal antibody (mAb)-induced down-regulation of RON receptor tyrosine kinase diminishes tumorigenic activities of colon cancer cells. *Int J Oncol* 2010; 37: 473-482.
159. Martin-Zanca D, Hughes SH, Barbacid M. A human oncogene formed by the fusion of truncated tropomyosin and protein tyrosine kinase sequences. *Nature* 1986; 319: 743-748.
160. Schramm A, Schulte JH, Astrahantseff K, et al. Biological effects of TrkA and TrkB receptor signaling in neuroblastoma. *Cancer letters* 2005; 228: 143-153.
161. Klein R, Jing SQ, Nanduri V, et al. The trk proto-oncogene encodes a receptor for nerve growth factor. *Cell* 1991; 65: 189-197.
162. Nakagawara A, Arima-Nakagawara M, Scavarda NJ, et al. Association between high levels of expression of the TRK gene and favorable outcome in human neuroblastoma. *The New England journal of medicine* 1993; 328: 847-854.
163. Aoyama M, Asai K, Shishikura T, et al. Human neuroblastomas with unfavorable biologies express high levels of brain-derived neurotrophic factor mRNA and a variety of its variants. *Cancer letters* 2001; 164: 51-60.
164. Tacconelli A, Farina AR, Cappabianca L, et al. TrkA alternative splicing: a regulated tumor-promoting switch in human neuroblastoma. *Cancer cell* 2004; 6: 347-360.



165. Rosenthal A, Lin JC. Modulation of neurotrophin signaling by monoclonal antibodies. *Handbook of experimental pharmacology* 2014; 220: 497-512.
166. Weeraratna AT, Arnold JT, George DJ, et al. Rational basis for Trk inhibition therapy for prostate cancer. *The Prostate* 2000; 45: 140-148.
167. McCarthy C, Walker E. Tropomyosin receptor kinase inhibitors: a patent update 2009 - 2013. *Expert opinion on therapeutic patents* 2014; 24: 731-744.
168. Liao W, Zhang H, Feng C, et al. Downregulation of TrkA protein expression by miRNA92a promotes the proliferation and migration of human neuroblastoma cells. *Molecular medicine reports* 2014; 10: 778-784.
169. Gudasheva TA, Antipova TA, Konstantinopolsky MA, et al. Nerve growth factor novel dipeptide mimetic GK-2 selectively activates TrkA postreceptor signaling pathways and does not cause adverse effects of native neurotrophin. *Doklady Biochemistry and biophysics* 2014; 456: 88-91.
170. Krause DS, Van Etten RA. Tyrosine kinases as targets for cancer therapy. *The New England journal of medicine* 2005; 353: 172-187.
171. Masiakowski P, Carroll RD. A novel family of cell surface receptors with tyrosine kinase-like domain. *J Biol Chem* 1992; 267: 26181-26190.
172. Yoda A, Oishi I, Minami Y. Expression and function of the Ror-family receptor tyrosine kinases during development: lessons from genetic analyses of nematodes, mice, and humans. *J Recept Signal Transduct Res* 2003; 23: 1-15.
173. Minami Y, Oishi I, Endo M, et al. Ror-family receptor tyrosine kinases in noncanonical Wnt signaling: their implications in developmental morphogenesis and human diseases. *Dev Dyn* 2010; 239: 1-15.
174. Reddy UR, Phatak S, Pleasure D. Human neural tissues express a truncated Ror1 receptor tyrosine kinase, lacking both extracellular and transmembrane domains. *Oncogene* 1996; 13: 1555-1559.
175. Forrester WC. The Ror receptor tyrosine kinase family. *Cell Mol Life Sci* 2002; 59: 83-96.
176. Oishi I, Takeuchi S, Hashimoto R, et al. Spatio-temporally regulated expression of receptor tyrosine kinases, mRor1, mRor2, during mouse development: implications in development and function of the nervous system. *Genes Cells* 1999; 4: 41-56.
177. Paganoni S, Ferreira A. Neurite extension in central neurons: a novel role for the receptor tyrosine kinases Ror1 and Ror2. *Journal of cell science* 2005; 118: 433-446.
178. Baskar S, Kwong KY, Hofer T, et al. Unique cell surface expression of receptor tyrosine kinase ROR1 in human B-cell chronic lymphocytic leukemia. *Clin Cancer Res* 2008; 14: 396-404.
179. Shabani M, Bayat AA, Jeddi-Tehrani M, et al. Ligation of human Fc receptor like-2 by monoclonal antibodies down-regulates B-cell receptor-mediated signalling. *Immunology* 2014; 143: 341-353.
180. Bicocca VT, Chang BH, Masouleh BK, et al. Crosstalk between ROR1 and the Pre-B cell receptor promotes survival of t(1;19) acute lymphoblastic leukemia. *Cancer Cell* 2012; 22: 656-667.
181. Shabani M, Asgarian Omran H, Farsangi MH, et al. Comparative expression profile of orphan receptor tyrosine kinase ROR1 in Iranian patients with lymphoid and myeloid leukemias. *Avicenna J Med Biotechnol* 2011; 3: 119-125.
182. Zhang S, Chen L, Wang-Rodriguez J, et al. The onco-embryonic antigen ROR1 is expressed by a variety of human cancers. *Am J Pathol* 2012; 181: 1903-1910.
183. Widhopf GF, 2nd, Cui B, Ghia EM, et al. ROR1 can interact with TCL1 and enhance leukemogenesis in Emu-TCL1 transgenic mice. *Proceedings of the National Academy of Sciences of the United States of America*; 111: 793-798.
184. Yang J, Baskar S, Kwong KY, et al. Therapeutic potential and challenges of targeting receptor tyrosine kinase ROR1 with monoclonal antibodies in B-cell malignancies. *PLoS One* 2011; 6: e21018.

185. Cui B, Widhopf II G, Prussak CE, et al. Cirtuzumab Vedotin (UC-961ADC3), An Anti-ROR1-Monomethyl Auristatin E Antibody-Drug Conjugate, Is a Potential Treatment For ROR1-Positive Leukemia and Solid Tumors. Blood (ASH Annual Meeting Abstracts) 2013; 122: Abstract No: 1637.