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The skewed balance between regulatory T cells and Th17 in chronic lymphocytic leukemia

Mehdi Yousefi^{1,2}, Ali Akbar Movassaghpour³, Karim Shamsasenjan³, Ghasem ghalamfarsa⁴, Sanam Sadreddini¹, Farhad Jadidi-Niaragh⁵*, and Mohammad Hojjat-Farsangi⁶

- 1. Immunology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran
- 2. Department of Immunology, Faculty of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran
- 3. Hematology Division, Department of Immunology, Faculty of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran
- 4. Cellular and Molecular Research Center, Yasuj University of Medical Sciences, Yasuj, Iran
- 5. Department of Immunology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran
- 6. Department of Oncology-Pathology, Immune and Gene Therapy Lab, Cancer Center Karolinska (CCK), Karolinska University Hospital Solna and Karolinska Institute, Stockholm, Sweden

Department of Immunology, School of Public Health Tehran University of Medical Sciences, Tehran 14155, Iran

Tel: +98-21-88953021; Fax: +98-21-88954913

 $Email: f-jadidin@razi.tums.ac.ir\ and\ farhad_jadidi1986@yahoo.com$

Running title: Imbalanced Treg and Th17 in CLL

^{*}Corresponding author: Farhad Jadidi-Niaragh PhD,

Abstract

While regulatory T (Treg) cells maintain self tolerance and inhibit anti-tumor responses, T helper (Th)17 cells may enhance inflammatory and anti-tumor responses. The balance between these two important T cell subsets has been skewed in many immunopathologic conditions such as autoimmune and cancer diseases. B cell chronic lymphocytic leukemia (CLL) is the most common form of leukemia in Western Word and is characterized with monoclonal expansion of B lymphocytes. There is evidence which implies that the progression of CLL is associated with expansion of Treg and downregulation of Th17 cells. In this review, we will discuss about immunobiology of Treg and Th17 cells and their role in immunopathogenesis of CLL as well as their reciprocal changes during disease progression.

Keywords: chronic lymphocytic leukemia, regulatory T cells, Th17, inerleukin-17, balance

Introduction

Chronic lymphocytic leukemia (CLL) is the most common form of leukemia in the Western world [1], which is characterized by clonal expansion of CD5⁺ B cells in periphery and lymphoid tissues [2]. The unmutated IG heavy chain variable (IGHV) region genes and higher expression of CD38 and ZAP-70 molecules are associated with poor prognosis in CLL patients [3].

CLL progression is mainly affected by the immune status of the CLL patients (Table 1). Several immune cells and mediators contribute to immunopathogenesis of CLL, such as regulatory T (Treg) cells and T helper (Th) 17 cells [4]. Increased absolute number of total T lymphocytes has been shown in CLL patients which was associated with expansion of CD8⁺ T cells, result in a low CD4:CD8 ratio [5]. However, some phenotypic and functional abnormalities have been observed in these cells. While the expression of CD69, CD57 and HLA-DR was increased, the expression of CD28 and CD62L was decreased in these T cells. This phenotype may represent terminally differentiated effector memory T cells [6]. Functionally, it has been shown that both CD4⁺ and CD8⁺ T cells of CLL patients secrete higher amounts of IL-4 compared to normal subjects [7], which led to overexpression of Bcl-2 anti-apoptotic B CLL cells [8]. Interestingly, IL-4-producing CD8⁺ T cells overexpress CD30 molecule which its ligation with CD30L on leukemic and normal B cells leads to proliferation and suppression of these cells, respectively [9]. The main effective anti-tumor CD4⁺ T cell subset involved in the control of B cell malignancies are thought to be Th1 cells [10]. Contrasting results regarding the frequency and balance status between Th1/Th2 cells have been reported in CLL patients. While a shift from Th1 towards Th2 cells has been shown to be associated with CLL progression [11], decreased Th2 [12] or no change in Th1 [13] cell number have also been reported in CLL patients. However, it is suggested that Th cells enhance the expression of anti-apoptotic molecules including MCL1, BCL-XL, BFL1 and Survivin in leukemic B cells in CD40L dependent manner [14]. Moreover, different T cell secreted cytokines such as IL-2, TNF- α , IFN γ , IFN- α and IL-13 were also demonstrated to support leukemic cell proliferation and survival [15]. Importantly, gene expression profiling of purified T cells isolated from CLL patients showed modified expression of genes responsible for differentiation and cytoskeletal organization in CD4⁺ T cells, and cytoskeletal formation, vesicle trafficking, and cytotoxicity in CD8⁺ T cells [16].

Dendritic cells (DC) which are important modulators of T cell responses show some defects in their maturation and antigen presentation process in CLL patients [17]. Interestingly, it has been shown that leukemic cells induce impaired immunologic synapse formation between DCs and T cells [18].

Natural killer (NK) cells are one of the main immune cells in anti-tumor responses. Although it is reported that their frequency is increased in CLL patients [19], however, their cytotoxic function is markedly suppressed [20].

Little is known regarding the serum levels of different cytokines in CLL. Yan and colleagues have performed a comprehensive study in which the wide variety of cytokines divided in 3 cluster (CL) including CL1 (CXCL9, CXCL10, CXCL11, CCL3, CCL4, CCL19, IL-5, IL-12, and IFN- γ), CL2 (TNF- α , IL-6, IL-8, and GM-CSF), and CL3 (IL-1, IL-2, IL-4, IL-15, IL-17, and IFN- α) were investigated in CLL patients. They showed that patients with progressive disease had high CL1 and low CL2 or CL3 levels [21].

However, recent data indicates that the balance status between Treg and Th17 cells has an important role in progression or alleviation of various autoimmune and cancer diseases [22]. Treg and Th17 cells are two important subsets of T cells, which may be considered as main immunomodulators in different immunopathologic conditions, such as tumor. While Treg cells suppress the anti-tumor responses, Th17 Page 3 of 22

cells may stimulate tumor rejection [22]. Recent data indicates that the CLL progression is associated with increased Treg and decreased Th17 numbers [23-25]. So, it may possible that the skewed balance between Treg and Th17 cells may assesses the disease outcome in CLL [22]. In this review, we tried to clarify the immunobiology of Treg and Th17 cells as well as their function in CLL progression. Moreover, we describe the factors responsible for their skewed balance which may be considered as worthy therapeutic targets in future.

Regulatory T cells

About a half century ago, Miller and colleagues were provided thymectomized mice and observed that mice were immunocompromized and developed autoimmunity and lymphoproliferative disease [26]. However, the term of immunosuppression was described in 1970s by Gershon *et al.* for the first time [27]. Subsequently, Sakaguchi *et al.* introduced the CD4⁺ suppressor or regulatory T (Treg) cells with increased CD25 (IL-2 receptor α) expression [28]. Identification of Treg lineage-specific transcription factor, forkhead box transcription factor 3 (FoxP3), led to better discrimination of these cells from other T cell subsets [29]. It has been shown that development of Treg cells depends on T cell receptor (TCR) specificity, similar to conventional CD4⁺ and CD8⁺ T cells [30], but with very limited overlap in TCR repertoire [31]. During thymic development of thymocytes, variations in avidity and duration of TCR signaling are the key points in T cell differentiation [32].

There are different subsets of Treg cells in circulation [33]. Treg cells can derive from the thymus (tTregs), which are also known as natural Treg (nTreg). Some Treg cells can also arise extrathymically in the periphery (pTregs) following induction of Foxp3 in consequence of antigen exposure, which are also known as inducible Treg (iTreg) [34]. The differentiation of tTreg cells is mediated through recognition of self-antigen/MHC molecules expressed on thymic epithelial cells in the presence of costimulatory signals such as CD28 [35], but not transforming growth factor (TGF)-β [36]. However, maximum tTreg differentiation and its peripheral maintenance are depend on continuous TCR signaling in the presence of costimulatory signals, particularly IL-2 [37] and to lesser extent IL-7 and IL-15 [38]. It has been suggested that generation of pTreg occurs in response to foreign antigens such as food, allergens, and particularly commensal bacteria [39]. The suboptimal TCR signals and high concentration of TGF-β are pivotal for induction of pTreg cells [40]. Surprisingly, while CD28 signaling is crucial for tTreg development [35], it prevents pTreg induction [41], implying different costimulatory requirements for tTreg and pTreg cells.

Based on some newly identified markers such as Helios and Neuropilin-1 (Nrp-1), it has been suggested that the majority of Treg cells in circulation are tTreg [42]. However, there is evidence which implies the expression of Helios and Nrp-1 in pTreg cells [43]. *In vivo* investigation of tTreg and pTreg TCR repertoire based on Nrp-1 expression showed that there is limited overlap between these two subsets of Treg cells which implies the different lineage development of tTregs and pTregs. Moreover, limited overlap was observed between Treg and conventional T cells which indicates pTreg cells constitute a very small subset of conventional T cells [43, 44].

Regulatory mechanisms of Treg cells can be divided into four mechanisms, including 1) secretion of inhibitory cytokines such as IL-10, TGF- β and IL-35, 2) direct cytolysis by granzymes, perforin, TRAIL-DR5 (TNF-related apoptosis inducing ligand-death receptor-5) and interaction with galectins, 3)

metabolic disruption through IL-2 deprivation and the generation of adenosine, and 4) DC modulation [45].

Th17 cells

Th17 cell is characterized with the production of cytokines, such as IL-17A, IL-17F, IL-6, IL-9, IL-21, IL-22, IL-23, IL-26 and TNF-α, and enhances the clearance of fungi and extracellular bacteria and tissue inflammation in autoimmune diseases. It seems that there are three distinct subsets of Th17 cells (as shown in figure 1), including conventional Th17 cells which are known as Th17-β cells, Th17-23 and Th17-1 cells [46]. Th17-β cells produce IL-17A, IL-17F, IL-10, CCL20 and express CXCR6 chemokine receptor. The differentiation of these cells is induced through TGF-β in combination with IL-6, IL-1, and IL-21. Th17-23 cells produce higher levels of IL-22 and CCL9 in addition to IFN-γ, IL-17A and IL-17F and express CXCR3 chemokine receptor. The differentiation of Th17-23 cells is promoted by the combination of cytokines such as IL-6, IL-23, IL-1β, and IL-21 (in the absence of TGF-β) [46-49]. The third subset of Th17 cells is Th17-1 cell which is characterized with the production of IFN-y and other Th17 common cytokines such as IL-17A, and expresses CCR6 and CXCR3 chemokine receptors. IL-1β, IL-21, IL-6, IL-12, TNF-α, and IL-23 are the main required cytokine microenvironment for development of Th17-1 cells [50, 51]. Among these cytokines, IL-12 can particularly induce IFNy generation in Th17 which leads to induction of Th17-1 subset, that rapidly lose the IL-17 production ability, and shift toward Th1 cells [51]. These Th17-derived Th1 cells (known as non-classic Th1 cells) differ from conventional Th1 cells, because Th17-1 cells express genes such as RORC, CD161, CCR6, IL-4-induced gene 1 and IL-17 receptor E, which are absent in conventional Th1 cells [52]. Moreover, it has recently been shown that the transcription factors Runx1 or Runx3, in association with T-box transcription factor (T-bet), are essential for the generation of Th17-1 cells [53]. It is unknown whether Th17-1 cells represent a stable subtype of Th17 cells or a transitional phenotype between Th17 and Th1 cells [54]. However, it has been supposed that Th17-1 cells derive from Th17-23 cells in the presence of Th1-polarizing cytokine milieu, particularly IL-12 [55].

The retinoic acid-related orphan receptor (ROR) γ t, ROR α , and STAT3 are three main lineage-specific transcription factors that induce Th17 differentiation [56]. STAT3 induces ROR γ t and ROR α under Th17-favoring conditions [57]. Other transcription factors such as ROR γ [58], interferon regulatory factor (IRF)4 [59], B-cell-activating transcription factor (BATF) [60], RUNX1 [61], c-musculoaponeurotic fibrosarcoma (-Maf) [62], AHR [63], and hypoxia-inducible factor 1 (HIF1) α [64] are also involved in the differentiation and development of Th17 cells.

Contrary to above mentioned factors which promote Th17 development, there are several factors that inhibit Th17 differentiation. It has been shown that different hallmark cytokines or lineage-specific transcription factors for Th1, Th2 and Treg cells prevent commitment to Th17 lineage or inhibit the generation of Th17-derived cytokines [65]. IL-27 and interferon (IFN) β , are also the negative regulator of Th17 cells [66, 67]. The suppressor of cytokine signalling 3 (SOCS3) and Ets-1 transcription factor are other inhibitors of Th17 cells [68, 69].

Regulatory T cells in CLL

Beyer and colleagues showed the increased frequency of Treg cells in CLL patients, which was correlated with disease progression, for the first time in 2005 [23]. The latter reports were confirmed the data published by Beyer *et al.* [24, 70]. Moreover, Jack and coworkers showed that the increased frequency of Treg cells in CLL patients is due to increased formation through CD27-CD70 interaction and increased resistance to apoptosis via increased levels of Bc1-2 anti-apoptotic protein [71]. Following these findings, the increased frequency and absolute number of Treg cells in CLL patients and its association with disease progression were demonstrated, repeatedly [72-76] (as shown in table 2).

Although it seems that CD4⁺ Treg cells are increased in CLL patients, little is known regarding the immunobiology of CD8⁺ Treg cells in these patients. We have recently been shown that the frequencies of both CD8⁺FoxP3⁺ and CD8⁺CD25⁺FoxP3⁺ Treg cells were increased in CLL patients and correlated with disease progression [4]. Other study has also been confirmed our experience [77]. Nunes and colleagues have also shown emergence of CD8⁺PD-1⁺ replicative senescent phenotype in early stage CLL and its correlation with disease progression [78].

The increased frequency of Treg cells not only was correlated with disease progression but also associated with different prognostic markers of CLL patients such as IGHV mutation status [72], CD38 expression [72], and the stage of disease [23, 24, 70, 73].

The functional capacity of Treg cells is another important issue in the immunobiology of Treg cells in CLL patients which is investigated in our [4] and some other studies [23, 77]. The first report regarding the Treg cells function in CLL was reported by Giannopoulos and coworkers which showed that the higher frequencies of Treg cells were correlated with decreased T cell responses against viral and tumor antigens [24]. Following this observation, Beyer et al. have been showed that the treatment of CLL patients with fludarabine led to decreased frequency and function of Treg cells as assessed by 5-bromo-2deoxyuridine (BrdU) incorporation test [23]. Another group showed that the cytotoxicity of Treg cells was increased in CLL patients as investigated through the expression of CD107a marker on Treg cells. However, they observed any change in Fas-ligand expression between CLL patients and normal subjects. They have also been demonstrated that Treg cells kill isolated B cells in part through granzyme A. The suppressive function of Treg cells on the proliferation of allogeneic peripheral blood mononuclear cells (PBMCs) was also intact as analyzed by Alamar Blue proliferation assay [79]. The intact suppressive function of Treg cells in CLL patients has also been reported by other studies as detected via CFSE (carboxyfluorescein diacetate succinimidyl ester) proliferation assay [72, 77]. It is suggested that Treg cells in CLL patients deprive effector T cells from IL-2 through secretion of soluble CD25, so which prevent anti-tumor responses exerted by effector T cells [80]. Recently, we have also been showed that the soluble factors in the supernatant of in vitro-stimulated Treg cells could inhibit the proliferation of both effector T cells and B cells from CLL patients similar to normal subjects [4]. Most recently, Rissiek et al. have been studied the composition and function of circulating T cells during the stages of monoclonal B cell lymphocytosis (MBL), early, and more advanced CLL. They showed that suppressive Treg cells arise early, during the pre-CLL stage of MBL. Moreover, T cell functional assays demonstrated an increasingly suppressive regulatory function initiating at the MBL stage. An increasingly suppressive phenotype has been observed during disease progression, which can be in part reversed by chemoimmunotherapy [81].

It should be clarified that the overall outcome resulted following the inhibition of Treg cells in CLL disease will be useful or not. Recently, it has been demonstrated that the treatment of CLL patients with thalidomide leads to better prognosis and reduced Treg cells [82]. Moreover, these reduced Treg cells were mainly Nrp-1⁺ which are tTreg [83]. Furthermore, vaccination of CLL patients with tumor lysate loaded-dendritic cells was associated with reduced Treg cells [84]. In addition, our recent data showed that expansion of Treg cells in association with disease progression in CLL patients could reduce NKT-like cells which have anti-tumor potential [85, 86]. Thus, it seems that downregulation of Treg cells will associated with better prognosis in CLL patients, however, these assumptions cannot be justified without performing comprehensive studies for clarifying of precise details regarding Treg cells in immunopathogenesis of CLL.

Th17 cells in CLL

Little is known regarding the immunobiology of IL-17 producing T cells in pathogenesis of cancer. There are some studies about Th17 cells in various solid tumors and a few haematological malignancies [87]. The protective role of Th17 cells has been implied in different solid tumors [88]. In contrast to solid tumors, little is known about the immunobiology of Th17 cells in hematologic malignancies, particularly CLL [89]. Initial studies about the IL-17 producing T cells in CLL were recently performed in a limited sample size and showed highly variable results [90-92]. Following these observations, we showed that the frequencies of both Th17 and Tc17 cells were markedly decreased in CLL patients and correlated with disease progression. Moreover, we found that reduced frequency of Th17 cells was associated with unmutated IGHV, which implied the protective role of Th17 cells in CLL. However we observed any association between the number of IL-17 producing T cells and other prognosis markers such as CD38 or ZAP-70 expression [93]. It has recently been demonstrated that the frequency of IL-17 producing follicular Th cells was increased in CLL patients and correlated with advanced CLL stages [94]. It should be noted that this study cannot imply the deleterious role of Th17 or Tc17 cells, because fTh cells can secrete different mediators in comparison with Th17 or Tc17 (such as IL-21) which may describe fThmediated tumor promotion [95]. Consistent with our results, Idler and colleagues have reported that treatment of CLL patients with lenalidomide led to induction of Th17 cells, which implies the protective role of Th17 cells in CLL [92]. Increased frequency of IL-17 producing T cells has also been reported in CLL/SLL patients compared to other B cell malignancies [90]. Another group has also been reported that increased frequency of Th17 cells was correlated with better prognostic markers and longer survival. In addition, non-Th17 IL-17A-expressing cells were present in CLL spleens as maturing granulocytes and mature mast cells, suggesting the microenvironmental milieu in leukemic spleens enhances the recruitment and expansion of Th17 and other IL-17-expressing cells [96]. Hus and coworkers have also suggested the protective role for Th17 cells in CLL patients. They showed that the frequency of Th17 cells and IL-17A levels were significantly decreased in advanced disease stages. Furthermore, the frequency of Th17 cells was lower in patients who died or responded to first-line therapy with fludarabine compared to surviving patients and non-responders, respectively. IL-17A inversely correlated with the time from CLL diagnosis to the start of therapy and was lower in patients who required treatment during follow-up. Moreover, Th-17 and IL-17A values were adversely correlated with bad prognostic factors such as 17p and 11q deletion, CD38 and ZAP-70 expression. The results of this study suggest that Th17 may play a beneficial role in CLL [97]. Altogether, it seems that the frequency of Th17 cells trends to be decreased with CLL progression implying the protective role of these cells in CLL patients (Table 3).

Although some data regarding the protective mechanisms exerted by Tc17 cells have already been reported [98], little is known regarding the protective mechanisms of IL-17 producing cells for the control of tumor cells. However, considering pro-inflammatory function of these cells, it seems that they provide an inflammatory microenvironment in which tumoral cells could be killed. Some subsets of Th17 cells have been shown to secret both IFN γ and IL-17 cytokines which are effective factors in anti-tumor responses [99]. It is suggested that Th17- β cells suppress anti-tumor responses in part through secretion of IL-10 and generation of adenosine via expression of CD73 and CD39. Controversially, it is supposed that Th17-23 cells enhance anti-tumor responses in part through secretion of IFN γ and degeneration of adenosine to inosine via adenosine deaminase expressed in these cells [55]. It doesn't seem that Th17 cells use contact-dependent mechanisms in their anti-tumor responses. However, there is no sufficient data regarding the functionality and precise mechanisms by which Th17 cells exert their protective role in CLL immunopathogenesis. Investigation of IFN γ -producing Th17 cells in CLL patients might be critical for better understanding the role of these cells in immunopathogenesis of CLL.

The skewed balance between Treg and Th17 in CLL

Generally, it is accepted that the progression of autoimmune diseases is associated with downregulation of Treg and expansion of Th17 cells [22, 100]. On the other hand, cancer progression is supposed to be linked to Treg expansion and Th17 reduction [93]. The skewed balance between these cells may result in the burden of tissue inflammation, autoimmune diseases or cancer [46]. Although several groups have been investigated this issue in autoimmune diseases, little is known regarding their imbalance in cancer [101]. In order to study this balance in CLL, we have recently analyzed different subsets of Treg and IL-17 producing T cells in these patients. Our results showed that the increased frequencies of different Treg subsets are associated with a downregulation of Th17 and Tc17 cells. Moreover, the imbalance between mRNA levels of lineage specific transcription factors FoxP3 and RORyt were confirmed the imbalance between Treg and IL-17 producing T cells [93]. Consistently, another group was recently demonstrated that the frequency of Th17 cells positively correlated with iNKT and adversely with Treg cells which implied the beneficial role of Th17 cells in CLL [97]. It has also been demonstrated that some therapeutic drugs such as lenalidomide trends to induce Th17 and inhibits Treg cells which was associated with the better CLL prognosis [92]. Imbalanced frequency of Treg/Th17 in CLL patients is also reported by other investigators [75, 102]. Recently, it has been suggested that CD39⁺ Treg cells can suppress Th17 cells [103]. Moreover, it has recently been showed that CD39⁺ Treg cells inhibited development of Th17 cells in human [104] and animal cancer models [105]. So which, we have enumerated CD39⁺ Treg cells in CLL patients. Our data showed that expansion of CD39⁺ Treg cells was associated with downregulation of IL-17 producing T cells (Th17 and Tc17). However, there was no significant correlation between the expansion of CD39⁺ Treg and downregulation of IL-17 producing T cells. Thus, it may be possible that these two subsets of T cells are modulated differently in CLL [93]. Consistently, Pulte and coworkers found that the frequencies of both CD4⁺CD39⁺ and CD8⁺CD39⁺ T cells were increased in CLL patients and correlated with disease progression [106]. It should be noted that their investigated populations were not Treg and solely represented CD39 molecule expressing T cells. In addition to Treg cells, there is evidence which implies malignant B cells as potent modulator of balance status between Treg and Th17 cells. It has been shown that malignant B cells inhibit IL-17 producing T cells and stimulate Treg development in non-Hodgkin's lymphoma by mechanisms in which CD27-CD70 or CD28-B7.1,2

interactions are involved [90]. Although the precise mechanism(s) of this modulation is(are) not clarified, it may be possible that malignant B cells secrete high amounts of TGF-β [107] and to lesser extent IL-2 [79] which their combinatorial effect is critical stimulator of Treg induction and Th17 inhibition [22]. Furthermore, since Th17 cells express IL-10R [108] and leukemic B cells are one of the main sources of IL-10, it may possible that leukemic B cells suppress Th17 cells in IL-10-dependent manner [109]. On the other hand, IL-10 produced by leukemic B cells not only can suppress Th17 cells, but also may induce Treg cells. Different modulators of Th17 differentiation including cytokines (such as IL-25 and IL-27) [46] may be involved in observed imbalance during disease progression which are not studied in CLL until now. The nurse-like cells and hepatocyte growth factor were recently identified as Treg inducer factors in CLL patients. It is demonstrated that hepatocyte growth factor is in large amounts in serum of CLL patients and interacts with its receptor on leukemic B cells, nurse-like cells and monocytes, known as c-MET, which leads to increased leukemic B cells survival. The mechanism by which nurse-like cells and monocytes induce Treg and inhibit other T cell subsets is in part through production of TGF-β, IL-10 and indoleamine 2,3-dioxygenase enzyme [110]. Thus, it may possible that during disease progression, nurse-like cells induces Treg and inhibits Th17 cells. A vascular growth factor (VEGF) receptor, Nrp-1, on Treg cells is another inducer of Treg cells in CLL patients. Recently, Piechnik and colleagues reported that the expression of Nrp-1 was significantly higher on leukemic lymphocytes, Treg and plasmacytoid DCs of CLL patients compared to normal individuals. The positive correlation was also detected between expression of VEGF receptors (FLT1, NRP1) and FoxP3 expression. Moreover, treatment with thalidomide which was previously showed decreased Treg cells, reduced the expression of Nrp-1 on Treg cells in vitro [83]. Thus, it seems that tTreg cells that express Nrp-1 may potently stimulated by VEGF during CLL progression. Beside the above mentioned factors that have identified as modulator of Treg/Th17 balance in CLL (Table 4), several other factors can also control their balance that are elusive in CLL (as shown in figure 2).

The mTOR is an important factor that can regulate Treg/Th17 balance in many immunopathologic conditions [111]. While the suboptimal activation of mTOR induces Treg, potent activation of mTOR, particularly mTORC1, induces Th1 and Th17 phenotypes [112]. However, treatment of CLL patients with different mTOR inhibitors in order to arrest the cell cycle was not successful, which might be in part due to its stimulatory function on Treg cells which was not addressed in these trials [113].

The transcription factor hypoxia inducible factor-1 (HIF-1) which is expressed in nearly all mammalian cells is another important factor that can regulate Treg/Th17 balance [114]. It has been demonstrated that HIF-1 can stimulate differentiation, expansion, function and survival of Th17 cells both *in vitro* and *in vivo* through direct and indirect manner. On the other hand, HIF-1 downregulates the development of FoxP3⁺ Treg cells [64]. However, as HIF-1 is overexpressed in leukemic CLL B cells and is associated with disease progression [115], it seems that HIF-1 cannot be considered as worthy tool for downregulation of Treg and expansion of Th17 cells.

Lipid oxidation can skew the Treg/Th17 balance toward Treg cells. Treg cells are more dependent to energy resulted from lipid oxidation (and not glycolytic process) than other T cell subsets [116]. On the other hand, lipid oxidation inhibits the development of Th17 cells, *in vitro* [116]. It has been reported that lipid oxidation is a mechanism of resistance to glucocorticoids -mediated cytotoxicity in CLL, and peroxisome proliferator-activated receptor (PPAR)α inhibition is an approach to improve the

therapeutic efficacy of glucocorticoids [117]. Thus, inhibition of lipid oxidation not only facilitates glucocorticoids-mediated cytotoxicity, but also prevents Treg expansion.

AMP-activated protein kinase (AMPK), which stimulates lipid metabolism and inhibits mTOR, is another Th17/Treg axis modulator [118]. It has been demonstrated that both tTreg and pTreg showed increased AMPK activity [116]. Moreover, AMPK is a potent negative regulator of mTORC1 [111]. Thus, AMPK activity drives Treg/Th17 balance toward Treg cells, however, as AMPK activation is involved in apoptosis of leukemic B cell [119], its inhibition may lead to tumor expansion in CLL.

PPARs are recently known as Treg/Th17 balance modulators. PPARs are transcription factors which are activated by fatty acids and seen as different isoforms including α , β/δ , and γ , each with different ligand [120]. It has been shown that PPAR γ activation inhibits the induction of ROR γ t, STAT3 and IL-17, so which it inhibits the differentiation of Th17 cells [121]. In addition, activation of PPAR β/δ suppresses both Th17 and Th1 cells and increases IL-10 production [122]. Moreover, ligation of PPAR α shifts Th1 response toward Th2 and declines the generation of pro-inflammatory cytokines [120]. On the other hand, ligation of PPAR γ and to lesser extent PPAR α not only decreased Th17 responses, but also induced Treg development [123, 124]. Interestingly, while mTOR activates PPAR γ , it inhibits PPAR α [111], which implies very complex network in regulating Treg/Th17 balance. Current data shows that while inhibition of PPAR α attenuates CLL progression, PPAR γ ligation leads to apoptosis of leukemic B cell [117, 125]. So which it seems that PPAR α may be better target for reversing Treg/Th17 imbalance in CLL patients compared to PPAR γ .

The cytosolic fatty acid-binding proteins (FABPs) are other players in skewed balance between Treg/Th17. It has been shown that FABPs could promote Th17 development in part through downregulation of PPARy [126].

Liver X receptors (LXR) which have two different isoforms including LXRα and LXRβ, express in Th cells and regulates gene products involved in metabolism of cholesterol and fatty acid [127]. It has been shown that the ligation of LXR inhibited expression of RORγt, IL-17 and Th17-related molecules [128]. This suppression was in part through induction of sterol regulatory element binding protein (SREBP-1), which binds to and suppresses AHR [129]. Although there is no data regarding the effect of LXR on Treg cells in CLL, however, its ligation has been led to apoptosis in leukemic B cells from CLL patients [130].

The hormone nuclear receptor, estrogen-related receptor (ERR) α is another candidate in regulating Treg/Th17 balance. It is suggested that ERR α ligation activates Th17 development, while inhibits Treg induction [131]. Moreover, in pre-clinical study, it has been shown that ligation of estrogen-related receptor was associated with ameliorative effects [132].

The relative amino acid abundance leads to activation of mTORC1 which in turn stimulates Th17 induction and inhibits Treg development [133]. On the other hand, amino acid starvation leads to downregulation of IL-17 producing T cells [134].

It has been shown that the activation of PI3K/Akt pathway leads to development of Th17 cells in part through stimulation of mTORC1, accumulation of RORγt and inhibition of Th17 inhibitor, Gfi-1 [135]. Since PI3K/Akt is regulator of both mTOR and HIF-1, it can be as an important modulator of Treg/Th17

axis [136]. Moreover, factors that interfere with PI3K/Akt pathway such as programmed death-1 (PD-1) inhibit Th17 differentiation and induce Treg phenotype [137]. However, as the stimulation of PI3K/Akt pathway stimulates the proliferative capacity of leukemic cells [138], it seems that blocking this pathway in order to expansion of Th17 cells may be rational. However, inhibition of this pathway may induce apoptosis in B-CLL cells [139].

The vitamin A metabolite, RA [140] and vitamin D are other factors, which can skew the Treg/Th17 balance toward anti-inflammatory status via suppression of Th17 development and enhancing Treg differentiation [141]. Although there is controversial results regarding the apoptotic effects of IL-21 on leukemic B cells [95], it seems that vitamin D insufficiency is associated with CLL poor prognosis [142].

The inflammatory lipid mediators may also play an important role in modulation of Treg/Th17 balance. It is reported that leukotriene (LT) B4 and prostaglandin (PG) E2 reduced the Treg frequency and Foxp3 expression in dose-dependent manner. On the other hand, LTB4 increases and PG2 decreases the IL-17 production and RORγt expression [143]. Available data indicates that increased levels of LTB4 [144] and PGE2 [145] is associated with CLL progression, so which targeting lipooxygenase or cyclooxygenase pathways may be useful in correction of Treg/Th17 imbalance in CLL patients.

The Treg/Th17 interaction can also regulate their balance. Fletcher and colleagues demonstrated that the CD39 expressing Treg cells could attenuate IL-17 generation in multiple sclerosis [103]. CD39 in cooperation with CD73 can cleave adenosine triphosphate (ATP) to immunosuppressive adenosine [103]. Thus, a decreased frequency of CD39⁺ Treg cells can lead to development of Th17 cells as we showed in CLL patients [93]. However, this hypothesis needs to investigation of adenosine receptor expression in Th17 cells.

The phenotype conversion or T cell plasticity may be another factor responsible for Treg/Th17 imbalance. It has been observed that Treg cells can express Th17-related transcription factors ROR γ t and ROR α and produce IL-17 [146]. Moreover, FoxP3 binds to and inhibits both ROR γ t and ROR α in dose dependent manner [147]. It has been demonstrated that STAT3 induces HIF-1 which binds to and targets Foxp3 for proteosomal degradation [64]. Furthermore, FoxP3 binds to and inhibits Runx transcription factor which is positive regulator of Th17 differentiation [61]. These results imply the role of pro-inflammatory or immunosuppressive microenvironment in phenotype conversion of Treg or Th17 cells. Moreover, it is proposed that epigenetic modification can also enhance the Treg conversion into Th17 cell [148].

Since TGF-β can induce both FoxP3 and RORγt in different cytokine milieu in a Smad2-/Smad3-dependent manner [147], it seems that TGF-β plays a central role in T cell fate decision.

Conclusion and future perspective

As discussed above, the progression of CLL was associated with upregulation of Treg and downregulation of IL-17 producing T cells [93]. This finding implied that Treg cells could be as tumor promoting cells whereas Th17 might be protective during CLL progression. However, the relevance of increased frequency of Treg and decreased Th17 cells to the expansion of leukemic B cells and disease progression are not fully understood. As Treg cells could suppress effector T cell subsets [4, 23], so which they might negatively influence the control of leukemic B cells expansion. Moreover, the mechanism(s)

by which Th17 cells can exert their anti-tumor effects in CLL is elusive. It may be possible that they suppress tumor progression through production of IFN- γ [55].

On the other hand, treatment of CLL patients with some therapeutic agents such as lenalidomide was associated with promotion of Th17 and inhibition of Treg cells and the better CLL prognosis [92]. The immunomodulatory effects of lenalidomide have also been investigated in other immune cells. It has been shown that lenalidomide can enhance anti-tumor function of NK, NKT-like and CD4⁺ T cells [149]. However, little is known regarding the immunomodulatory effects of several novel therapeutic agents such as ibrutinib, idelalisib, or ABT-199 [150]. In spite of these data, there are many unanswered questions regarding the immunobiology of different subsets of Treg and Th17 cells in CLL which require further investigations. However, it seems that the control of CLL needs to inhibition of Treg expansion and induction of Th17, concomitantly. Consistently, there are several factors (as discussed in previous section) which could affect the Treg/Th17 balance. So, combinatorial regulation of such Treg/Th17 modulators might be as a promising approach in treatment of CLL. In addition, identifying the new factors which can regulate Treg/Th17 balance might be as interesting field for investigators.

Conflict of interest statement

None of the authors has any conflict of interest to declare.

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None.

Executive summary

Background

- CLL is the most common leukemia in the Western Word which is incurable until now.
- The imbalanced Treg/Th17 frequency has been observed in several autoimmune and cancer disease.
- The balance status between Treg and Th17 cells may influences immunopathogenesis of CLL.

Regulatory T cells

- Treg cells generally divides into two groups, known as thymus-derived tTreg and peripherally induced pTreg cells.
- FoxP3 is the lineage-specific transcription factor and is indispensable for Treg development.
- Treg cells maintain self tolerance and suppress anti-tumor responses.

Th17 cells

- Th17 cells typically divided into three groups, known as Th17-β, Th17-23 and Th17-1 cells.
- RORyt and STAT3 transcription factors are the main inducers of Th17 differentiation.
- It seems that Th17 cells inhibit tumor progression.

Treg cells in CLL

- The frequencies of different subsets of Treg cells are increased in CLL patients and correlated with disease progression.
- The function of Treg cells is intact in CLL patients.

Th17 cells in CLL

- Th17 cells are decreased in CLL patients.
- It seems that Th17 cells are protective during disease progression.

The skewed Treg/Th17 in CLL

• Downregulation of Th17 cells is associated with Treg expansion during CLL progression.

Conclusion & future perspective

- CLL progression is associated with Treg expansion and Th17 reduction.
- Several factors can affect Treg/Th17 balance which are elusive in CLL.
- There is no comprehensive study regarding the different Th17 subsets in CLL.
- The precise anti-tumor mechanisms exerted by Th17 cells are unknown.

Figure legends

Figure 1: The differentiation of different subsets of Th17 cells. There are three subsets of Th17 cells including Th17- β , Th17-23 and Th17-1. While differentiation of Th17- β cells depends on TGF- β , differentiation of Th17-23 and Th17-1 is TGF- β independent. Non-classic Th1 cells derive from Th17-1 cells and differ from conventional Th1 cells.

Figure 2: Balance status between Treg and Th17 cells in cancer. The factors that influence the development of Treg and Th17 cells are shown. While the development of Th17 cells inhibits tumor progression, Treg expansion can promote tumor burden. Treg: regulatory T cell, IL: interleukin, Th: T helper, FoxP3: forkhead box protein P3, TGF-β: transforming growth factor-β, RA: retinoic acid, LTB4: leukotriene B4, PGE2: prostaglandin E2, oxLDL: oxidized-low density lipoprotein, mTOR: mammalian target of rapamycin, HIF-1: hypoxia inducible factor-1, AMPK: AMP-activated protein kinase, PPAR: peroxisome proliferator-activated receptor, FABPs: fatty acid-binding proteins, LXR: Liver X receptor, ERR: estrogen-related receptor, Nrp-1: neuropilin-1, HGF: hepatocyte growth factor

References

- 1. Morton LM, Wang SS, Devesa SS, Hartge P, Weisenburger DD, Linet MS: Lymphoma incidence patterns by WHO subtype in the United States, 1992-2001. *Blood* 107(1), 265-276 (2006).
- 2. Chiorazzi N, Rai KR, Ferrarini M: Chronic lymphocytic leukemia. *N. Engl. J. Med.* 352(8), 804-815 (2005).
- 3. Mellstedt H, Choudhury A: T and B cells in B-chronic lymphocytic leukaemia: Faust, Mephistopheles and the pact with the Devil. *Cancer Immunol. Immun.* 55(2), 210-220 (2006).
- 4. Jadidi-Niaragh F, Yousefi M, Memarian A *et al.*: Increased Frequency of CD8+ and CD4+ Regulatory T Cells in Chronic Lymphocytic Leukemia: Association with Disease Progression. *Cancer Invest.* 31(2), 121-131 (2013).
 - ** It demonstrates that the frequency of different Treg subsets is increased in CLL patioent which correlated with disease progression.
- 5. Zaknoen S, Kay N: Immunoregulatory cell dysfunction in chronic B-cell leukemias. *Blood reviews* 4(3), 165-174 (1990).
- 6. Riches JC, Ramsay AG, Gribben JG: Immune reconstitution in chronic lymphocytic leukemia. *Curr. Hematol. Malig. Rep.* 7(1), 13-20 (2012).
- 7. Mu X, Kay NE, Gosland MP, Jennings CD: Analysis of blood T-cell cytokine expression in B-chronic lymphocytic leukaemia: evidence for increased levels of cytoplasmic IL-4 in resting and activated CD8 T cells. *Br. J. Haematol.* 96(4), 733-735 (1997).
- 8. Kay NE, Han L, Bone N, Williams G: Interleukin 4 content in chronic lymphocytic leukaemia (CLL) B cells and blood CD8+ T cells from B-CLL patients: impact on clonal B-cell apoptosis. *Br. J. Haematol.* 112(3), 760-767 (2001).
- 9. Cerutti A, Kim EC, Shah S *et al.*: Dysregulation of CD30+ T cells by leukemia impairs isotype switching in normal B cells. *Nat. Immunol.* 2(2), 150-156 (2001).
- 10. Haabeth OaW, Lorvik KB, Hammarström C *et al.*: Inflammation driven by tumour-specific Th1 cells protects against B-cell cancer. *Nat. Commun.* 2, 240 (2011).
- 11. Podhorecka M, Dmoszynska A, Rolinski J, Wasik E: T type 1/type 2 subsets balance in B-cell chronic lymphocytic leukemia—the three-color flow cytometry analysis. *Leuk. Res.* 26(7), 657-660 (2002).
- 12. Hill S, Peters S, Ayliffe M, Merceica J, Bansal A: Reduced IL-4 and interferon-gamma (IFN-) expression by CD4 T cells in patients with chronic lymphocytic leukaemia. *Clin. Exp. Immunol.* 117, 8-11 (1999).
- 13. Gallego A, Vargas J, Castejon R *et al.*: Production of intracellular IL-2, TNF-α, and IFN-γ by T cells in B-CLL. *Clin. Cytom.* 56(1), 23-29 (2003).
- 14. Granziero L, Ghia P, Circosta P *et al.*: Survivin is expressed on CD40 stimulation and interfaces proliferation and apoptosis in B-cell chronic lymphocytic leukemia. *Blood* 97(9), 2777-2783 (2001).
- 15. Herishanu Y, Katz B-Z, Lipsky A, Wiestner A: Biology of chronic lymphocytic leukemia in different microenvironments: clinical and therapeutic implications. *Hematol. Oncol. Clin. North. Am.* 27(2), 173-206 (2013).
- 16. Görgün G, Holderried TA, Zahrieh D, Neuberg D, Gribben JG: Chronic lymphocytic leukemia cells induce changes in gene expression of CD4 and CD8 T cells. *J. Clin. Invest.* 115(7), 1797-1805 (2005).
- 17. Orsini E, Pasquale A, Maggio R *et al.*: Phenotypic and functional characterization of monocyte-derived dendritic cells in chronic lymphocytic leukaemia patients: influence of neoplastic CD19+cells in vivo and in vitro. *Br. J. Haematol.* 125(6), 720-728 (2004).
- 18. Ramsay AG, Johnson AJ, Lee AM *et al.*: Chronic lymphocytic leukemia T cells show impaired immunological synapse formation that can be reversed with an immunomodulating drug. *J. Clin. Invest.* 118(7), 2427-2437 (2008).

- 19. Ravandi F, O'brien S: Immune defects in patients with chronic lymphocytic leukemia. *Cancer Immunol. Immun.* 55(2), 197-209 (2006).
- 20. Foa R, Fierro M, Lusso P *et al.*: Reduced natural killer T-cells in B-cell chronic lymphocytic leukaemia identified by three monoclonal antibodies: Leu-11, A10, AB8. 28. *Br. J. Haematol.* 62(1), 151-154 (1986).
- 21. Yan X-J, Dozmorov I, Li W *et al.*: Identification of outcome-correlated cytokine clusters in chronic lymphocytic leukemia. *Blood* 118(19), 5201-5210 (2011).
- 22. Jadidi-Niaragh F, Mirshafiey A: The deviated balance between regulatory T cell and Th17 in autoimmunity. *Immunopharmacol. Immunotoxicol.* 34(5), 727-739 (2012).
- 23. Beyer M, Kochanek M, Darabi K *et al.*: Reduced frequencies and suppressive function of CD4+ CD25hi regulatory T cells in patients with chronic lymphocytic leukemia after therapy with fludarabine. *Blood* 106(6), 2018-2025 (2005).
 - * It analyzed the frequency and function of Treg cells in CLL patients for the first time.
- 24. Giannopoulos K, Schmitt M, Kowal M *et al.*: Characterization of regulatory T cells in patients with B-cell chronic lymphocytic leukemia. *Oncol. Rep.* 20(3), 677-682 (2008).
- 25. Jadidi-Niaragh F, Ghalamfarsa G, Yousefi M, Tabrizi MH, Shokri F: Regulatory T cells in chronic lymphocytic leukemia: implication for immunotherapeutic interventions. *Tumour Biol.* 34(4), 2031-2039 (2013).
- 26. Miller JF: Effect of neonatal thymectomy on the immunological responsiveness of the mouse. *Proc. R. Soc. Lond., B, Biol. Sci.* 156(964), 415-428 (1962).
- 27. Gershon RK, Kondo K: Cell interactions in the induction of tolerance: the role of thymic lymphocytes. *Immunology* 18(5), 723 (1970).
- 28. Sadlack B, Löhler J, Schorle H *et al.*: Generalized autoimmune disease in interleukin-2-deficient mice is triggered by an uncontrolled activation and proliferation of CD4+ T cells. *Eur. J. Immunol.* 25(11), 3053-3059 (1995).
- 29. Fontenot JD, Gavin MA, Rudensky AY: Foxp3 programs the development and function of CD4+ CD25+ regulatory T cells. *Nat. Immunol.* 4(4), 330-336 (2003).
 - * It introduced FoxP3 as lineage specific transcription factor for Treg cells.
- 30. Larkin J, Rankin AL, Picca CC *et al.*: CD4+ CD25+ regulatory T cell repertoire formation shaped by differential presentation of peptides from a self-antigen. *J. Immunol.* 180(4), 2149-2157 (2008).
- 31. Liu X, Nguyen P, Liu W *et al.*: T Cell Receptor CDR3 Sequence but Not Recognition Characteristics Distinguish Autoreactive Effector and Foxp3+ Regulatory T Cells. *Immunity* 31(6), 909-920 (2009).
- 32. Singer A, Adoro S, Park J-H: Lineage fate and intense debate: myths, models and mechanisms of CD4-versus CD8-lineage choice. *Nat. Rev. Immunol.* 8(10), 788-801 (2008).
- 33. Jadidi-Niaragh F, Mirshafiey A: Regulatory T-cell as orchestra leader in immunosuppression process of multiple sclerosis. *Immunopharmacol. Immunotoxicol.* 33(3), 545-567 (2011).
- 34. Abbas AK, Benoist C, Bluestone JA *et al.*: Regulatory T cells: recommendations to simplify the nomenclature. *Nat. Immunol.* 14(4), 307-308 (2013).
- 35. Tai X, Cowan M, Feigenbaum L, Singer A: CD28 costimulation of developing thymocytes induces Foxp3 expression and regulatory T cell differentiation independently of interleukin 2. *Nat. Immunol.* 6(2), 152-162 (2005).
- 36. Marie JC, Letterio JJ, Gavin M, Rudensky AY: TGF-β1 maintains suppressor function and Foxp3 expression in CD4+ CD25+ regulatory T cells. *J. Exp. Med.* 201(7), 1061-1067 (2005).
- 37. Knoechel B, Lohr J, Kahn E, Bluestone JA, Abbas AK: Sequential development of interleukin 2–dependent effector and regulatory T cells in response to endogenous systemic antigen. *J. Exp. Med.* 202(10), 1375-1386 (2005).

- 38. Vang KB, Yang J, Mahmud SA, Burchill MA, Vegoe AL, Farrar MA: IL-2,-7, and-15, but not thymic stromal lymphopoeitin, redundantly govern CD4+ Foxp3+ regulatory T cell development. *J. Immunol.* 181(5), 3285-3290 (2008).
- 39. Josefowicz SZ, Lu L-F, Rudensky AY: Regulatory T cells: mechanisms of differentiation and function. *Annu. Rev. Immunol.* 30, 531-564 (2012).
- 40. Kretschmer K, Apostolou I, Hawiger D, Khazaie K, Nussenzweig MC, Von Boehmer H: Inducing and expanding regulatory T cell populations by foreign antigen. *Nat. Immunol.* 6(12), 1219-1227 (2005).
- 41. Semple K, Nguyen A, Yu Y, Wang H, Anasetti C, Yu X-Z: Strong CD28 costimulation suppresses induction of regulatory T cells from naive precursors through Lck signaling. *Blood* 117(11), 3096-3103 (2011).
- 42. Lathrop SK, Santacruz NA, Pham D, Luo J, Hsieh C-S: Antigen-specific peripheral shaping of the natural regulatory T cell population. *J. Exp. Med.* 205(13), 3105-3117 (2008).
- 43. Yadav M, Stephan S, Bluestone JA: Peripherally induced tregs–role in immune homeostasis and autoimmunity. *Front. Immunol.* 7;4:232, (2013).
- 44. Yadav M, Louvet C, Davini D *et al.*: Neuropilin-1 distinguishes natural and inducible regulatory T cells among regulatory T cell subsets in vivo. *J. Exp. Med.* 209(10), 1713-1722 (2012).
- 45. Caridade M, Graca L, Ribeiro RM: Mechanisms underlying CD4+ Treg immune regulation in the adult: from experiments to models. *Front. Immunol.* 18;4:378, (2013).
- 46. Jadidi-Niaragh F, Mirshafiey A: Th17 cell, the new player of neuroinflammatory process in multiple sclerosis. *Scand. J. Immunol.* 74(1), 1-13 (2011).
- 47. Ghoreschi K, Laurence A, Yang X-P *et al.*: Generation of pathogenic TH17 cells in the absence of TGF-[bgr] signalling. *Nature* 467(7318), 967-971 (2010).
- 48. Sutton C, Brereton C, Keogh B, Mills KH, Lavelle EC: A crucial role for interleukin (IL)-1 in the induction of IL-17–producing T cells that mediate autoimmune encephalomyelitis. *The J. Exp. Med.* 203(7), 1685-1691 (2006).
- 49. Yang L, Anderson DE, Baecher-Allan C *et al.*: IL-21 and TGF-&bgr; are required for differentiation of human TH17 cells. *Nature* 454(7202), 350-352 (2008).
- 50. Zielinski CE, Mele F, Aschenbrenner D *et al.*: Pathogen-induced human TH17 cells produce IFN-[ggr] or IL-10 and are regulated by IL-1. *Nature* 484(7395), 514-518 (2012).
- 51. Annunziato F, Cosmi L, Santarlasci V *et al.*: Phenotypic and functional features of human Th17 cells. *J. Exp. Med.* 204(8), 1849-1861 (2007).
- 52. Maggi L, Santarlasci V, Capone M *et al.*: Distinctive features of classic and nonclassic (Th17 derived) human Th1 cells. *Eur. J. Immunol.* 42(12), 3180-3188 (2012).
- 53. Wang Y, Godec J, Ben-Aissa K *et al.*: The Transcription Factors T-bet and Runx Are Required for the Ontogeny of Pathogenic Interferon-γ-Producing T Helper 17 Cells. *Immunity* 40(3), 355-366 (2014).
- 54. El-Behi M, Rostami A, Ciric B: Current views on the roles of Th1 and Th17 cells in experimental autoimmune encephalomyelitis. *J. Neuroimmune Pharm.* 5(2), 189-197 (2010).
- 55. Bailey SR, Nelson MH, Himes RA, Li Z, Mehrotra S, Paulos CM: Th17 cells in cancer: the ultimate identity crisis. *Front. Immunol.* 5, 276 (2014).
- 56. Yang XO, Pappu BP, Nurieva R *et al.*: T helper 17 lineage differentiation is programmed by orphan nuclear receptors RORα and RORγ. *Immunity* 28(1), 29-39 (2008).
- 57. Yang XO, Panopoulos AD, Nurieva R *et al.*: STAT3 regulates cytokine-mediated generation of inflammatory helper T cells. *J. Biol. Chem.* 282(13), 9358-9363 (2007).
- 58. Ivanov II, Mckenzie BS, Zhou L *et al.*: The Orphan Nuclear Receptor RORγt Directs the Differentiation Program of Proinflammatory IL-17+ T Helper Cells. *Cell* 126(6), 1121-1133 (2006).
- 59. Brüstle A, Heink S, Huber M *et al.*: The development of inflammatory TH-17 cells requires interferon-regulatory factor 4. *Nat. Immunol.* 8(9), 958-966 (2007).

- 60. Schraml BU, Hildner K, Ise W *et al.*: The AP-1 transcription factor Batf controls TH17 differentiation. *Nature* 460(7253), 405-409 (2009).
- 61. Zhang F, Meng G, Strober W: Interactions among the transcription factors Runx1, RORγt and Foxp3 regulate the differentiation of interleukin 17–producing T cells. *Nat. Immunol.* 9(11), 1297-1306 (2008).
- 62. Bauquet AT, Jin H, Paterson AM *et al.*: The costimulatory molecule ICOS regulates the expression of c-Maf and IL-21 in the development of follicular T helper cells and TH-17 cells. *Nat. Immunol.* 10(2), 167-175 (2008).
- 63. Quintana FJ, Basso AS, Iglesias AH *et al.*: Control of Treg and TH17 cell differentiation by the aryl hydrocarbon receptor. *Nature* 453(7191), 65-71 (2008).
- Dang EV, Barbi J, Yang H-Y *et al.*: Control of TH17/Treg Balance by Hypoxia-Inducible Factor 1. *Cell* 146(5), 772-784 (2011).
- 65. Harrington LE, Hatton RD, Mangan PR *et al.*: Interleukin 17–producing CD4+ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nat. Immunol.* 6(11), 1123-1132 (2005).
- 66. Stumhofer JS, Laurence A, Wilson EH *et al.*: Interleukin 27 negatively regulates the development of interleukin 17–producing T helper cells during chronic inflammation of the central nervous system. *Nat. Immunol.* 7(9), 937-945 (2006).
- 67. Guo B, Chang EY, Cheng G: The type I IFN induction pathway constrains Th17-mediated autoimmune inflammation in mice. *J. Clin. Invest.* 118(5), 1680-1690 (2008).
- 68. Chen Z, Laurence A, Kanno Y *et al.*: Selective regulatory function of Socs3 in the formation of IL-17-secreting T cells. *P. Natl. Acad. Sci. USA* 103(21), 8137-8142 (2006).
- 69. Moisan J, Grenningloh R, Bettelli E, Oukka M, Ho I-C: Ets-1 is a negative regulator of Th17 differentiation. *J. Exp. Med.* 204(12), 2825-2835 (2007).
- 70. Giannopoulos K, Schmitt M, Własiuk P *et al.*: The high frequency of T regulatory cells in patients with B-cell chronic lymphocytic leukemia is diminished through treatment with thalidomide. *Leukemia* 22(1), 222-224 (2007).

* The frequency and function of Treg cells were analyzed in CLL patients.

71. Jak M, Mous R, Remmerswaal EB *et al.*: Enhanced formation and survival of CD4+ CD25hi Foxp3+ T-cells in chronic lymphocytic leukemia. *Leuk. Lymphoma* 50(5), 788-801 (2009).

* The mechanisms by which Treg cells accumulated in CLL patients were introduced.

- 72. Weiss L, Melchardt T, Egle A, Grabmer C, Greil R, Tinhofer I: Regulatory T cells predict the time to initial treatment in early stage chronic lymphocytic leukemia. *Cancer* 117(10), 2163-2169 (2011).
- 73. D'arena G, Laurenti L, Minervini MM *et al.*: Regulatory T-cell number is increased in chronic lymphocytic leukemia patients and correlates with progressive disease. *Leuk. Res.* 35(3), 363-368 (2011).
- 74. Piper K, Karanth M, Mclarnon A *et al.*: Chronic lymphocytic leukaemia cells drive the global CD4+ T cell repertoire towards a regulatory phenotype and leads to the accumulation of CD4+ forkhead box P3+ T cells. *Clin. Exp. Immunol.* 166(2), 154-163 (2011).
- 75. Perry C, Herishanu Y, Hazan-Halevy I *et al.*: Reciprocal changes in regulatory T cells and Th17 helper cells induced by exercise in patients with chronic lymphocytic leukemia. *Leuk. Lymphoma* 53(9), 1807-1810 (2012).
- 76. Lad DP, Varma S, Varma N, Sachdeva MUS, Bose P, Malhotra P: Regulatory T-cells in B-cell chronic lymphocytic leukemia: their role in disease progression and autoimmune cytopenias. *Leuk. Lymphoma* 54(5), 1012-1019 (2013).
- 77. Biancotto A, Dagur PK, Fuchs JC, Wiestner A, Bagwell CB, Mccoy JP: Phenotypic complexity of T regulatory subsets in patients with B-chronic lymphocytic leukemia. *Mod. Pathol.* 25(2), 246-259 (2011).
 - * The different Treg cell subsets were analyzed in CLL patients.

- 78. Nunes C, Wong R, Mason M, Fegan C, Man S, Pepper C: Expansion of a CD8+ PD-1+ replicative senescence phenotype in early stage CLL patients is associated with inverted CD4: CD8 ratios and disease progression. *Clin. Cancer Res.* 18(3), 678-687 (2012).
- 79. Lindqvist CA, Christiansson LH, Thörn I *et al.*: Both CD4+ FoxP3+ and CD4+ FoxP3- T cells from patients with B-cell malignancy express cytolytic markers and kill autologous leukaemic B cells in vitro. *Immunology* 133(3), 296-306 (2011).
- 80. Lindqvist CA, Christiansson LH, Simonsson B, Enblad G, Olsson-Strömberg U, Loskog AS: T regulatory cells control T-cell proliferation partly by the release of soluble CD25 in patients with B-cell malignancies. *Immunology* 131(3), 371-376 (2010).
- 81. Rissiek A, Schulze C, Bacher U *et al.*: Multidimensional scaling analysis identifies pathological and prognostically relevant profiles of circulating T-cells in chronic lymphocytic leukemia. *Int. J. Cancer* 15;135(10), 2370-9 (2014).
- 82. Skórka K, Bhattacharya N, Własiuk P *et al.*: Thalidomide Regulation of NF-κB Proteins Limits Tregs Activity in Chronic Lymphocytic Leukemia. *Adv. Clin. Exp.Med.*23(1), 25-32 (2013).
- 83. Piechnik A, Dmoszynska A, Omiotek M *et al.*: The VEGF receptor, neuropilin-1, represents a promising novel target for chronic lymphocytic leukemia patients. *Int. J. Cancer* 133(6), 1489-1496 (2013).
- 84. Palma M, Hansson L, Choudhury A *et al.*: Vaccination with dendritic cells loaded with tumor apoptotic bodies (Apo-DC) in patients with chronic lymphocytic leukemia: effects of various adjuvants and definition of immune response criteria. *Cancer Immunol. Immun.* 61(6), 865-879 (2012).
- 85. Jadidi-Niaragh F, Jeddi-Tehrani M, Ansaripour B, Razavi SM, Sharifian RA, Shokri F: Reduced frequency of NKT-like cells in patients with progressive chronic lymphocytic leukemia. *Med. Oncol.* 29(5), 3561-3569 (2012).
- 86. Ghalamfarsa G, Hadinia A, Yousefi M, Jadidi-Niaragh F: The role of natural killer T cells in B cell malignancies. *Tumour Biol.* 34(3), 1349-1360 (2013).
- 87. Dhodapkar KM, Barbuto S, Matthews P *et al.*: Dendritic cells mediate the induction of polyfunctional human IL17-producing cells (Th17-1 cells) enriched in the bone marrow of patients with myeloma. *Blood* 112(7), 2878-2885 (2008).
- 88. Gnerlich JL, Mitchem JB, Weir JS *et al.*: Induction of Th17 cells in the tumor microenvironment improves survival in a murine model of pancreatic cancer. *J. Immunol.* 185(7), 4063-4071 (2010).
- 89. Prabhala RH, Pelluru D, Fulciniti M *et al.*: Elevated IL-17 produced by TH17 cells promotes myeloma cell growth and inhibits immune function in multiple myeloma. *Blood* 115(26), 5385-5392 (2010).
- 90. Yang Z-Z, Novak AJ, Ziesmer SC, Witzig TE, Ansell SM: Malignant B cells skew the balance of regulatory T cells and TH17 cells in B-cell non-Hodgkin's lymphoma. *Cancer Res.* 69(13), 5522-5530 (2009).
- 91. Giannopoulos K, Własiuk P, Dmoszyńska A, Roliński J, Schmitt M: Peptide vaccination induces profound changes in the immune system in patients with B-cell chronic lymphocytic leukemia. *Folia. Histochem. Cytobiol.* 49(1), 161-160 (2011).
- 92. Idler I, Giannopoulos K, Zenz T *et al.*: Lenalidomide treatment of chronic lymphocytic leukaemia patients reduces regulatory T cells and induces Th17 T helper cells. *Br. J. Haematol.* 148(6), 948-950 (2010).
- 93. Jadidi-Niaragh F, Ghalamfarsa G, Memarian A *et al.*: Downregulation of IL-17-producing T cells is associated with regulatory T cell expansion and disease progression in chronic lymphocytic leukemia. *Tumour Biol.* 34(2), 929-940 (2013).
 - * It demonstrates imbalanced Treg/Th17 cells during CLL progression.
- 94. Cha Z, Zang Y, Guo H *et al.*: Association of peripheral CD4+ CXCR5+ T cells with chronic lymphocytic leukemia. *Tumour Biol.* 34(6), 3579-3585 (2013).

- 95. Ghalamfarsa G, Jadidi-Niaragh F, Hojjat-Farsangi M *et al.*: Differential regulation of B-cell proliferation by IL21 in different subsets of chronic lymphocytic leukemia. *Cytokine* 62(3), 439-445 (2013).
- 96. Jain P, Javdan M, Feger FK *et al.*: Th17 and non-Th17 interleukin-17-expressing cells in chronic lymphocytic leukemia: delineation, distribution, and clinical relevance. *haematologica* 97(4), 599-607 (2012).
- 97. Hus I, Bojarska-Junak A, Chocholska S *et al.*: Th17/IL-17A might play a protective role in chronic lymphocytic leukemia immunity. *PLoS ONE* 8(11), e78091 (2013).
- 98. Tajima M, Wakita D, Satoh T, Kitamura H, Nishimura T: IL-17/IFN-γ double producing CD8+ T (Tc17/IFN-γ) cells: a novel cytotoxic T-cell subset converted from Tc17 cells by IL-12. *Int. Immunol.* 23(12), 751-9 (2011).
- 99. Hamaï A, Pignon P, Raimbaud I *et al.*: Human TH17 Immune Cells Specific for the Tumor Antigen MAGE-A3 Convert to IFN-γ–Secreting Cells as They Differentiate into Effector T Cells In Vivo. *Cancer Res.* 72(5), 1059-1063 (2012).
- 100. Gol-Ara M, Jadidi-Niaragh F, Sadria R, Azizi G, Mirshafiey A: The role of different subsets of regulatory T cells in immunopathogenesis of rheumatoid arthritis. *Arthritis* 2012, (2012).
- 101. Li MO, Wan YY, Flavell RA: T cell-produced transforming growth factor-β1 controls T cell tolerance and regulates Th1-and Th17-cell differentiation. *Immunity* 26(5), 579-591 (2007).
- 102. Tang D, Niu Q, Zeng T *et al.*: [Ratio balance of Th17 and Treg cells in peripheral blood of patients with chronic lymphocytic leukemia]. *Zhongguo shi yan xue ye xue za zhi* 21(2), 329-333 (2013).
- 103. Fletcher JM, Lonergan R, Costelloe L *et al.*: CD39+ Foxp3+ regulatory T Cells suppress pathogenic Th17 cells and are impaired in multiple sclerosis. *J. Immunol.* 183(11), 7602-7610 (2009).
- 104. Ye Z-J, Zhou Q, Zhang J-C *et al.*: CD39+ regulatory T cells suppress generation and differentiation of Th17 cells in human malignant pleural effusion via a LAP-dependent mechanism. *Respir Res* 12(77), 1-10 (2011).
- 105. Clayton A, Al-Taei S, Webber J, Mason MD, Tabi Z: Cancer exosomes express CD39 and CD73, which suppress T cells through adenosine production. *J. Immunol.* 187(2), 676-683 (2011).
- 106. Pulte D, Furman RR, Broekman MJ *et al.*: CD39 expression on T lymphocytes correlates with severity of disease in patients with chronic lymphocytic leukemia. *Clin. Lymphoma Myeloma Leuk.* 11(4), 367-372 (2011).
- 107. Lotz M, Ranheim E, Kipps TJ: Transforming growth factor beta as endogenous growth inhibitor of chronic lymphocytic leukemia B cells. *J. Exp. Med.* 179(3), 999-1004 (1994).
- 108. Huber S, Gagliani N, Esplugues E *et al.*: Th17 cells express interleukin-10 receptor and are controlled by Foxp3– and Foxp3+ regulatory CD4+ T cells in an interleukin-10 dependent manner. *Immunity* 34(4), 554 (2011).
- 109. Carter NA, Rosser EC, Mauri C: Interleukin-10 produced by B cells is crucial for the suppression of Th17/Th1 responses, induction of T regulatory type 1 cells and reduction of collagen-induced arthritis. *Arthritis Res. Ther.* 14(1), R32 (2012).
- 110. Giannoni P, Pietra G, Travaini G *et al.*: Chronic Lymphocytic Leukemia Nurse-like cells express the hepatocyte growth factor receptor (c-MET) and indoleamine 2, 3-dioxygenase and display features of immunosuppressive type 2 skewed macrophages. *haematologica*, haematol. 2013.091405 (2014).
- 111. Waickman AT, Powell JD: mTOR, metabolism, and the regulation of T-cell differentiation and function. *Immunol. Rev.* 249(1), 43-58 (2012).
- 112. Powell JD, Pollizzi KN, Heikamp EB, Horton MR: Regulation of immune responses by mTOR. *Annu. Rev. Immunol.* 30, 39 (2012).

- 113. Zent CS, Laplant BR, Johnston PB *et al.*: The treatment of recurrent/refractory chronic lymphocytic leukemia/small lymphocytic lymphoma (CLL) with everolimus results in clinical responses and mobilization of CLL cells into the circulation. *Cancer* 116(9), 2201-2207 (2010).
- 114. Shi LZ, Wang R, Huang G *et al.*: HIF1α–dependent glycolytic pathway orchestrates a metabolic checkpoint for the differentiation of TH17 and Treg cells. *J. Exp. Med.* 208(7), 1367-1376 (2011).
- 115. Shachar I, Cohen S, Marom A, Becker-Herman S: Regulation of CLL survival by hypoxia-inducible factor and its target genes. *FEBS lett.* 586(18), 2906-2910 (2012).
- 116. Michalek RD, Gerriets VA, Jacobs SR *et al.*: Cutting edge: distinct glycolytic and lipid oxidative metabolic programs are essential for effector and regulatory CD4+ T cell subsets. *J. Immunol.* 186(6), 3299-3303 (2011).
- 117. Tung S, Shi Y, Wong K *et al.*: PPARα and fatty acid oxidation mediate glucocorticoid resistance in chronic lymphocytic leukemia. *Blood* 122(6), 969-980 (2013).
- 118. Lee WH, Kim SG: AMPK-dependent metabolic regulation by PPAR agonists. *PPAR Res.* 2010, (2010).
- 119. Campàs C, López JM, Santidrián AF *et al.*: Acadesine activates AMPK and induces apoptosis in B-cell chronic lymphocytic leukemia cells but not in T lymphocytes. *Blood* 101(9), 3674-3680 (2003).
- 120. Yang Y, Lovett-Racke AE, Racke MK: Regulation of immune responses and autoimmune encephalomyelitis by PPARs. *PPAR Res.* 2010, (2010).
- 121. Klotz L, Burgdorf S, Dani I *et al.*: The nuclear receptor PPARγ selectively inhibits Th17 differentiation in a T cell–intrinsic fashion and suppresses CNS autoimmunity. *J. Exp. Med.* 206(10), 2079-2089 (2009).
- 122. Kanakasabai S, Chearwae W, Walline CC, Iams W, Adams SM, Bright JJ: Peroxisome proliferator-activated receptor δ agonists inhibit T helper type 1 (Th1) and Th17 responses in experimental allergic encephalomyelitis. *Immunology* 130(4), 572-588 (2010).
- 123. Wohlfert EA, Nichols FC, Nevius E, Clark RB: Peroxisome proliferator-activated receptor γ (PPARγ) and immunoregulation: enhancement of regulatory T cells through PPARγ-dependent and-independent mechanisms. *J. Immunol.* 178(7), 4129-4135 (2007).
- 124. Hontecillas R, Bassaganya-Riera J: Peroxisome proliferator-activated receptor γ is required for regulatory CD4+ T cell-mediated protection against colitis. *J. Immunol.* 178(5), 2940-2949 (2007).
- 125. Spaner D, Lee E, Shi Y *et al.*: PPAR-alpha is a therapeutic target for chronic lymphocytic leukemia. *Leukemia* 27(5), 1090-1099 (2012).
- 126. Li B, Reynolds JM, Stout RD, Bernlohr DA, Suttles J: Regulation of Th17 differentiation by epidermal fatty acid-binding protein. *J. Immunol.* 182(12), 7625-7633 (2009).
- 127. Repa JJ, Liang G, Ou J *et al.*: Regulation of mouse sterol regulatory element-binding protein-1c gene (SREBP-1c) by oxysterol receptors, LXRα and LXRβ. *Genes Dev.* 14(22), 2819-2830 (2000).
- 128. Cui G, Qin X, Wu L *et al.*: Liver X receptor (LXR) mediates negative regulation of mouse and human Th17 differentiation. *J. Clin. Invest.* 121(2), 658-670 (2011).
- 129. Bengoechea-Alonso MT, Ericsson J: SREBP in signal transduction: cholesterol metabolism and beyond. *Curr. Opin. Cell Biol.* 19(2), 215-222 (2007).
- 130. Geyeregger R, Shehata M, Zeyda M *et al.*: Liver X receptors interfere with cytokine-induced proliferation and cell survival in normal and leukemic lymphocytes. *J. Leukoc. Bio.* 86(5), 1039-1048 (2009).
- 131. Michalek RD, Gerriets VA, Nichols AG *et al.*: Estrogen-related receptor-α is a metabolic regulator of effector T-cell activation and differentiation. *P. Natl. Acad. Sci. USA* 108(45), 18348-18353 (2011).

- 132. Nishikawa M, Uemura Y, Komada F, Ohno T, Katayama N, Shirakawa S: Clinical Response to Busramustine (KM-2210) in Chronic Lymphocytic Leukemia: A Pilot Evaluation of Estrogen Receptor in Relation to Its Therapeutic Effect. *Jpn. J. Clin. Oncol.* 18(4), 327-333 (1988).
- 133. Cobbold SP, Adams E, Farquhar CA *et al.*: Infectious tolerance via the consumption of essential amino acids and mTOR signaling. *P. Natl. Acad. Sci. USA* 106(29), 12055-12060 (2009).
- 134. Sundrud MS, Koralov SB, Feuerer M *et al.*: Halofuginone inhibits TH17 cell differentiation by activating the amino acid starvation response. *Science* 324(5932), 1334-1338 (2009).
- 135. Kurebayashi Y, Nagai S, Ikejiri A *et al.*: PI3K-Akt-mTORC1-S6K1/2 axis controls Th17 differentiation by regulating Gfi1 expression and nuclear translocation of RORγ. *Cell Rep.* 1(4), 360-373 (2012).
- 136. Nakamura H, Makino Y, Okamoto K *et al.*: TCR engagement increases hypoxia-inducible factorlα protein synthesis via rapamycin-sensitive pathway under hypoxic conditions in human peripheral T cells. *J. Immunol.* 174(12), 7592-7599 (2005).
- 137. D'addio F, Riella LV, Mfarrej BG *et al.*: The link between the PDL1 costimulatory pathway and Th17 in fetomaternal tolerance. *J. Immunol.* 187(9), 4530-4541 (2011).
- 138. Palacios F, Abreu C, Prieto D *et al.*: Activation of the PI3K/AKT pathway by microRNA-22 results in CLL B-cell proliferation. *Leukemia*, (2014).
- 139. Amrein L, Shawi M, Grenier J, Aloyz R, Panasci L: The phosphatidylinositol-3 kinase I inhibitor BKM120 induces cell death in B-chronic lymphocytic leukemia cells in vitro. *Int. J. Cancer* 133(1), 247-252 (2013).
- 140. Mucida D, Park Y, Kim G *et al.*: Reciprocal TH17 and regulatory T cell differentiation mediated by retinoic acid. *Science* 317(5835), 256-260 (2007).
- 141. Joshi S, Pantalena L-C, Liu XK *et al.*: 1, 25-Dihydroxyvitamin D3 ameliorates Th17 autoimmunity via transcriptional modulation of interleukin-17A. *Mol. Cell. Biol.* 31(17), 3653-3669 (2011).
- 142. Aref S, Ibrahim L, Azmy E: Prognostic impact of serum 25-hydroxivitamin D [25 (OH) D] concentrations in patients with lymphoid malignancies. *Hematology* 18(1), 20-25 (2013).
- 143. Chen H, Qin J, Wei P *et al.*: Effects of leukotriene B4 and prostaglandin E2 on the differentiation of murine Foxp3+ T regulatory cells and Th17 cells. *Prostaglandins Leukot. Essent. Fatty Acids* 80(4), 195-200 (2009).
- 144. Runarsson G, Liu A, Mahshid Y *et al.*: Leukotriene B4 plays a pivotal role in CD40-dependent activation of chronic B lymphocytic leukemia cells. *Blood* 105(3), 1274-1279 (2005).
- 145. Ryan EP, Pollock SJ, Kaur K *et al.*: Constitutive and activation-inducible cyclooxygenase-2 expression enhances survival of chronic lymphocytic leukemia B cells. *Clin. Immunol.* 120(1), 76-90 (2006).
- 146. Ayyoub M, Deknuydt F, Raimbaud I *et al.*: Human memory FOXP3+ Tregs secrete IL-17 ex vivo and constitutively express the TH17 lineage-specific transcription factor RORγt. *P. Natl. Acad. Sci. USA* 106(21), 8635-8640 (2009).
- 147. Ichiyama K, Yoshida H, Wakabayashi Y *et al.*: Foxp3 inhibits RORγt-mediated IL-17A mRNA transcription through direct interaction with RORγt. *J. Biol. Chem.* 283(25), 17003-17008 (2008).
- 148. Koenen HJ, Smeets RL, Vink PM, Van Rijssen E, Boots AM, Joosten I: Human CD25highFoxp3pos regulatory T cells differentiate into IL-17–producing cells. *Blood* 112(6), 2340-2352 (2008).
- 149. Acebes-Huerta A, Huergo-Zapico L, Gonzalez-Rodriguez AP *et al.*: Lenalidomide Induces Immunomodulation in Chronic Lymphocytic Leukemia and Enhances Antitumor Immune Responses Mediated by NK and CD4 T Cells. *Biomed Res. Int.* 2014, (2014).
- 150. Tausch E, Mertens D, Stilgenbauer S: Advances in treating chronic lymphocytic leukemia. *F1000prime Rep.* 6, (2014).